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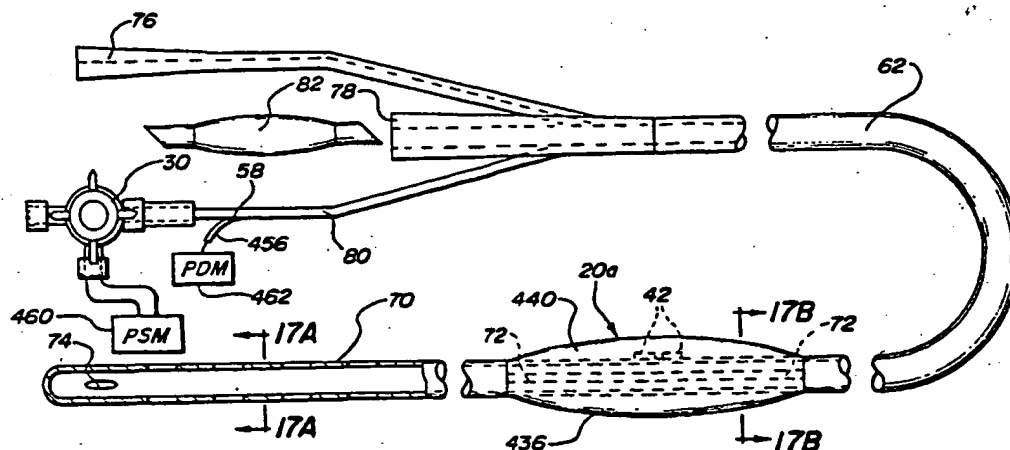


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(54) Title: INTRA-ABDOMINAL PRESSURE MEASUREMENT APPARATUS AND METHOD



(57) Abstract

An improved tonometric catheter (20) has tubing (22), luer lock (24), closed end (32), and perforations (34). A permeable membrane (36) encloses perforations (34) and forms sampling chamber (40). Also on tubing (22) are sensors (42) located within membrane (36) to sense a desired property.

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INTRA-ABDOMINAL PRESSURE MEASUREMENT APPARATUS AND METHODBackground and Summary of the Invention

This invention relates to medical diagnostic equipment and methods and is particularly concerned with hollow viscus tonometry and direct or remote sensing of pressure within organs such as intra-abdominal pressure (IAP) and related properties indicative of the condition of the internal organs.

Until the advent of the tonometric method (see U.S. Patent No. 4,643,192, issued February 17, 1987) few considered any aspect of acid-base balance when attempting to monitor or maintain the adequacy of tissue oxygenation. Yet acid-base balance is primarily determined by the balance between the protons released during the release of energy by ATP hydrolysis and the resynthesis of ATP by oxidative phosphorylation. The hydrolysis of ATP generates 150,000 mmols of H⁺ each day in a resting 70 Kg man. All, but the 1% of this fixed acid load excreted by the kidneys each day, is presumed to be consumed in the resynthesis of ATP by oxidative phosphorylation. When the delivery of oxygen fails to satisfy the energy needs of the tissue the rate of ATP hydrolysis exceeds the rate of synthesis and the pH falls as the degree of unreversed ATP hydrolysis increases.

It is now widely accepted that global measurements of oxygen delivery, consumption and extraction do not provide reliable information about the adequacy of local or even "global" tissue oxygenation in patients. The indirect measurement of gastric intramucosal pH (pHi) as described in U.S. Patent Nos. 4,643,192; 5,158,083; 5,186,172 provides clinicians with a minimally invasive yet sensitive means of detecting the development of a tissue acidosis, and hence inadequacy of tissue oxygenation, in a region of the body that is one of the first to exhibit an

inadequacy of tissue oxygenation in shock. Use of the measurement has revealed that some 50% to 60% of patients having major surgery and 80% of ICU patients develop an intramucosal acidosis during their illness despite the
5 conventional appearance of being adequately resuscitated.

The degree and duration of the presence of a gastric intramucosal acidosis are highly sensitive measures of the risk of developing ischemic gut mucosal injury and its putative consequences, namely the translocation of
10 bacteria and their toxins, cytokine release, organ dysfunctions and failures, and death from the organ failures. By providing an index of the adequacy of tissue oxygenation in one of the first parts of the body to exhibit dysoxia in shock the measurement of gastric
15 intramucosal pH improves the opportunity to obtain advanced and accurate warning of impending complications and to intervene in time to prevent them. More importantly timely therapeutic measures that restore the intramucosal pH to normality and "gut-directed" therapies incorporating
20 measures that reverse an intramucosal acidosis are associated with an improved outcome. "pH-directed" therapy has in addition been shown to improve outcome in a prospective randomized multicenter study of medical and surgical ICU patients.

25 The measurements of gastric intramucosal pH have revealed deficiencies in currently accepted practices. It has, for example, become apparent that empirical increases in global oxygen delivery may be redundant in some 40% to 50% of patients having major cardiovascular surgery who do
30 not develop a gastric intramucosal acidosis and whose prognosis is excellent. It is further apparent that the vogue of increasing global oxygen delivery to supranormal levels cannot be relied upon to prevent or to reverse the presence of an intramucosal acidosis. Of particular
35 concern is the intramucosal acidosis that may be induced by measures, notably the transfusion of red blood cells and dobutamine, that increase global oxygen delivery in

patients who do not have an intramucosal acidosis but whose global oxygen delivery is considered too low.

Intra-abdominal pressure (IAP) measurement is increasingly used in clinical practice as a guide to intra-peritoneal pathology and as a predictor of renal function. In particular, IAP monitoring may be useful in the increasing practice of minimally invasive procedures such as laparoscopic surgery. In laparoscopic surgery gas is insufflated into the peritoneum to provide an operating field and improve the field of vision. Steps are taken to monitor the IAP caused by the insufflated gas to reduce the chances of overinsufflation.

Increased IAP may be associated with a variety of clinical situations that may adversely affect cardiac, renal, respiratory, and metabolic functions. See Iberti, et al., The Determination Of Intra-Abdominal Pressure Using A Transurethral Bladder Catheter: Clinical Validation of the Technique Anesthesiology, Vol. 70, No. 1, Jan. 1989, p. 47. Diebel, et al., Effect Of Increased Intra-Abdominal Pressure On Hepatic Arterial, Portal Venous And Hepatic Microcirculatory Blood Flow, The Journal Of Trauma, Vol. 33, No. 2, 1992, p. 279 and references cited therein which are expressly incorporated by references herein.

Examples of situations where IAP monitoring may be useful include trauma, aneurism repair, laparoscopic surgery, massive edema, and bowel distention. The measurement of IAP may be complimentary to monitoring intramucosal pH as studies have demonstrated an increase in IAP may cause a reduction of blood flow to the microcirculation of the gut thus causing a fall in intramucosal pH.

A number of different techniques have been used to measure IAP. The measurement of IAP was performed in the last century using rectal and oesophageal sounds connected to a Marcy tambour. Wagoner GW, American Journal of Medical Science 1926; 171: 697-707. Intra-abdominal pressure has been measured transrectally in dogs --

Thorington JM, Schmidt CP, A Study of Urinary Output and Blood Pressure Changes Resulting in Experimental Ascites, Am J Med Sci 1923; 165: 880-890. Also used has been a Miller Abbott tube in the rectum and a Hamilton manometer
5 in the stomach to record pressure -- Bradley SE, Bradley GP, The Effect on Increased Intra-abdominal Pressure on Renal Function in Man, J Clin Invest 1947; 26:1010-1022. IAP has also been measured through invasive techniques namely via catheters inserted transcutaneously into the
10 peritoneal cavity Diebel, et al., Journal Of Trauma, 1992, p. 280. Direct cannulation of the peritoneal cavity and the injection of air have also been used -- Emerson H., Intra-abdominal Pressures, Arch Int Med 1911;7:754 -- Overholt RH, Intraperitoneal Pressure, Arch Surg
15 1931;22:691-703.

IAP monitoring techniques have been developed for intravesical methods where the bladder is partially filled with saline and the fluid pressure in the bladder has been monitored as an indicium of IAP. The original technique
20 has undergone some modification by Iberti, et al. -- Iberti TJ, Lieber CE, Benjamin E., Determination of Intra-abdominal Pressure Using a Transurethral Bladder Catheter, Clinical Validation of the Technique, Anesthesiology 1989;70:47-50. The intravesical technique has become an
25 accepted standard for IAP monitoring. While reliable, it is cumbersome to perform and interferes with the estimation of the patient's urinary output readings. Because the urinary tract is essentially sealed by the intravesical technique of IAP monitoring, it is also of limited value
30 for continuous IAP monitoring because urine output cannot be indefinitely obstructed.

THE TONOMETRIC METHOD

References to the tonometrics method, tonometrics
35 and tonometer in this application refers to the use of catheters or like devices as part of a system to measure

and monitor internal conditions of a subject, such as pH_i and/or IAP.

The measurement of pH in the most superficial layer of the mucosa is obtained indirectly by measuring the partial pressure of carbon dioxide (pCO_2 ; PCO_2) in the lumen of the gut and the bicarbonate concentration in arterial blood and substituting these two values in the Henderson-Hasselbalch equation or some modification thereof. See "Gastric Intramucosal pH as a Therapeutic Index of Tissue Oxygenation in Critically Ill Patients," *Lancet* 1992; 339; 195-99, incorporated herein by reference. The indirect measurement of the pH of the wall of the organ (pH indirect or pH_i) may be employed because it is believed or assumed that the pCO_2 in the most superficial layers of the mucosa is in equilibrium with that in the luminal contents with which it is in contact. It is further based upon the assumption that the bicarbonate concentration in the tissue is the same as that being delivered to it in arterial blood and that the pK_a , 6.1, is the same as that in plasma.

At present, measurements of pCO_2 in the lumen of the stomach are obtained by infusing saline into the silicone balloon of a gastrointestinal tonometer, allowing the pCO_2 in the saline to equilibrate with that in the lumen of the gut; recording the equilibration time; aspirating the saline; measuring the pCO_2 in the saline with a blood gas analyzer; using a nomogram to derive the steady-state adjusted pCO_2 from the equilibration time and the measured pCO_2 ; and then derive the intramucosal pH from the steady-state adjusted pCO_2 obtained and the bicarbonate concentration in a substantially contemporaneous sample of arterial blood. Again, see U.S. Patent Nos. 4,643,192, issued February 17, 1987; 5,174,290, issued December 29, 1992; and 5,186,172, issued February 16, 1993; as well as copending U.S. Applications, Serial Number 719,097, filed June 20, 1991; Serial Number 994,721, filed December 22, 1992 and Serial Number 014,624, filed February 8, 1993; all three issued patents being completely and expressly

incorporated herein by reference. The precision of the measurement of gastric intramucosal pH between healthy subjects is excellent, the gastric intramucosal pH in a healthy subject being the same as the pH in his arterial
5 blood.

The prior art (see U.S. Patent No. 4,643,192) has recognized that intestinal ischemia, and to a lesser degree, stress ulceration, are two problems that plague physicians involved in the management of patients in
10 intensive care units. Intestinal ischemia, in particular, has an insidious onset and may not be detected until days after the intestine has become completely and irreversibly compromised. A delay in the diagnosis of intestinal ischemia may have devastating consequences for a patient.
15 The availability of means for early diagnosis and management of patients with these problems would have immediate applicability in all intensive care units, especially where the procedure can be conveniently conducted with reasonable safety and reliability.

It has been established that a fall in the intramucosal pH may precede the development of intestinal ischemia and stress ulceration. As discussed in U.S. Patent No. 4,643,192, which is expressly incorporated
20 herein by reference, entitled "Hollow Viscus Tonometry" a fall in intramucosal pH also occurs within minutes of inducing intestinal ischemia in dogs. The fall in pH in
25 intestinal mucosa, and hence the likelihood of ischemia or stress ulceration, can be reliably calculated from a $p\text{CO}_2$ (partial pressure of CO_2), or other indicia of pH, in
30 lumenal fluid and the bicarbonate concentration in arterial blood. The method of calculating the pH in intestinal mucosal tissue, pursuant to principles of the prior patent no. 4,643,192, has been validated by directed measurements under a variety of conditions simulating clinical problems.
35 A correlation coefficient in the order of 0.92 to 0.95 has been obtained in each of 16 dogs. The validity of the procedure is inherently extensible to humans, and indeed

may also be useful in assessing the vitality of other hollow organs and tissue. See R.G. Fiddian-Green et al. "Splanchnic Ischemia and Multiple Organ Failure".

To measure the pCO_2 in the lumen of the gut it has heretofore been necessary to obtain and remove a sample of fluid that has been in contact with the wall of the gut for a certain time period, usually at least half an hour. It has now been observed that it is somewhat difficult to manually aspirate the sampling fluid or medium from a tonometric catheter located in the gut or other internal focus with any consistency. It is much easier to obtain such samples from the stomach, but samples obtained from the stomach frequently contain foreign material that can damage a gas analyzer.

As taught in the prior patent no. 4,643,192, the desired sample or samples can be obtained from the gut using a catheter tube (called a tonometric catheter) having a walled sampling chamber on the tube with the sampling chamber being in sample-specific communication with the hollow interior of the tube. The wall of the sampling chamber comprises a material which is substantially impermeable to liquid yet is highly permeable to gas. One suitable material is polydimethylsiloxane elastomer.

In use the catheter is introduced into a patient to place the sampling chamber at a desired site within the gut. An aspirating liquid or medium is employed to fill the interior of the sampling chamber. The sampling chamber is left in place at the desired sampling site long enough to allow the gases present to diffuse through the wall of the sampling chamber into the aspirating liquid. The time should be long enough for the gases to equilibrate. The liquid impermeable nature of the sample chamber wall material prevents both the aspirating liquid from leaking out of the chamber and also the intrusion of any liquids into the aspirating liquid. After the appropriate or desired amount of placement time has elapsed the aspirating liquid is aspirated along with the gases which have

diffused into it. The sample thus obtained is analyzed for gas content, in particular for pCO_2 . In this way the pCO_2 within the lumen of the gut can be reliably measured with the fluid being free from luminal debris.

5 In carrying out the diagnostic method taught in my prior patent the pCO_2 measurement is utilized in conjunction with a measurement of the bicarbonate ion concentration (HCO_3^-) in an arterial blood sample of the patient for determining the pH of the tract wall.

10 Depending upon the particular condition of a given patient, the catheter may be left in place and samples may be taken at periodic intervals so that pH values may be periodically calculated. The procedure has a high reliability in accurately determining the adequacy
15 of organ tissue oxygenation, and diagnosing intestinal ischemia in its incipient stages. Such determination or detection can be useful in treating the patient so that the potentially devastating consequences resulting from less timely detection may often be avoided.

20 While the sampling techniques taught in my prior patent have provided highly accurate and reliable results, it has now been observed that there are instances (in the care of the critically ill in intensive care units, for example) in which remote sensing of the organ or organ-wall
25 condition and automatic determination or calculation of the organ or organ-wall pH would be advantageous and easier to effectuate. This method would thus partially or totally eliminate the need for the somewhat cumbersome aspiration of the sampling fluid or medium which fills the sampling
30 chamber; it may also eliminate the need for the sampling chamber to be in sampling-medium communication with any other part of the device. There is also a need to extend the benefits of tonometric sampling and sensing to other internal hollow viscous organs. To this end, there is a
35 need for new and different tonometric devices specifically adapted to allow my sensing and sampling techniques to be

performed with ease in a clinical environment, and in combination with other procedures.

The importance and significance of determining the pH of the wall of a given hollow viscous organ has been recently dramatically magnified as a result of the recent recognition that the pH of the wall of a given organ can be employed to accurately evaluate the vitality and/or stability of that organ as well as others; this is in contrast to merely determining whether such an organ is experiencing an ischemic event. Further, certain organs can be selected for monitoring, either alone or in combination, and evaluation of this organ or these organs can aid in predicting the overall condition of the patient, or the onset of a multitude of pathologies, including predicting or identifying such events as multiple organ failure. Such a methodology can be employed to greatly enhance and supplement the monitoring of the critically ill, for example.

It has also been observed that an unusually large negative bias is encountered when measuring the pCO_2 in saline with certain blood gas analyzers (including those manufactured by Nova Biomedical, L. Eschweiller and Mallinckrodt) that have been standardized for blood but not for saline. The presence or absence of unacceptable bias may be determined by the use of reference samples of tonometered saline. The inter-instrumental bias encountered when measuring arterial blood gases and especially pCO_2 in saline with different blood gas analyzers requires that each institution derive its own normal values for meaningful use in clinical practice. It is reported that the precision of the measurements made within a static environment may be improved and unacceptable interinstrumental bias eliminated, in whole or in part, by using gelofusine, phosphate-buffered or bicarbonate-buffered saline, or mixtures thereof. Unfortunately the diffusional characteristics are altered and, in the case of the buffered saline, the time constant extended from about

18 to about 48 minutes. The nomograms provided for the determination of steady-state adjusted $p\text{CO}_2$ in saline cannot be used for the determination of intramucosal pH with these fluids.

5 The time constant may be reduced to seconds by using an electrochemical $p\text{CO}_2$ sensor directly in the lumen of the gut and measuring the $p\text{CO}_2$ in either liquid or gaseous luminal contents, as described herein. Unfortunately, $p\text{CO}_2$ sensors are known for their ability to
10 drift and cannot be easily recalibrated in vivo. The bias of a measurement of $p\text{CO}_2$ made with a conventional $p\text{CO}_2$ sensor is a function of the chemical composition of the gaseous or liquid fluid into which it is immersed and must be separately standardized for measurements made in
15 environments of different chemical composition such as blood and saline, as discussed above. In one set of experiments the bias observed in each of two instruments varied between 1% and 45%, the bias in each of the different solutions being consistent between the blood gas
20 analyzers used. The large and frequent changes in the chemical composition of luminal contents, especially in the stomach, almost certainly cause large and unpredictable changes in bias that render measurements made outside the protective environment of a silicone balloon tonometer
25 prone to unacceptable error. The deposition of luminal contents on the electrode membrane is an additional source of potential error with the use of a naked sensor, i.e., one without a tonometric walled sampling chamber.

 In one aspect, the present invention provides a
30 new apparatus and method for remotely sensing organ condition and conveying an electromagnetic signal, e.g. an electrical current or optical signal, to an electronic or optical apparatus located outside the organ under investigation. In one embodiment, a chemically sensitive
35 electronic transducer (or plurality of transducers), such as a field effect transistor, is attached to a tonometric catheter for introduction into the organ along with the

tonometric catheter. The first electronic sensor, preferably non-temperature, generates and conveys an electromagnetic signal indicative of some desired aspect of organ condition, e.g., indicative of the pCO_2 , pH and/or pO_2 level of the organ or organ-wall. For example, in one preferred embodiment, mean ambient pCO_2 , pH and/or pO_2 of lumenal fluid or the like is measured or monitored via wire or other suitable electromagnetic energy conveying means to an electronic circuit which interprets the electromagnetic signal and produces a report of the organ condition. The electronic circuit may include an input for receiving a separately determined signal indicative of the blood pH of the patient. Using this pCO_2 , pH and/or pO_2 measurement along with blood (preferably arterial) pH data, the electronic circuit determines the pH of the organ wall under test and thereby provides information for determining the organ's current condition or perhaps predicting the organ's future condition. The electronic circuit may be suitably constructed from analog components, digital components or both.

In another embodiment, a pH, pCO_2 or pO_2 sensitive colorimetric substance is injected into an area adjacent to the organ, e.g., into the sampling chamber of the tonometric catheter, and an optical sensor is employed to detect color change in order to determine the pH of the wall of that organ. The optical sensor can either be disposed in or on the tonometric catheter for introduction into the area adjacent the organ or it may be disposed outside the organ with fiber optic cable optically coupling the sensor to the tonometric catheter site at which the pH sensitive substance has been injected.

In another aspect the present invention provides a variety of new and different tonometric catheter devices for sensing and/or sampling a fluid or gas property (such as pH, pO_2 , pCO_2 , and the like) which is indicative of the condition of an internal organ, in conjunction or combination with a walled catheter tube adapted for

delivery or draining fluids, such as nasogastric tubes, urinary catheters, ureteric catheters, intestinal feeding tubes, wound or abdominal drains (suction or regular) and biliary tubes, catheters and stents, with or without remote
5 sensing means for pH, pCO₂ and/or pO₂.

In still another aspect or embodiment, the device employs two separate walled catheter tubes, one tonometric catheter tube for the measurement of a fluid or gas property, that is in communication with the sampling
10 chamber; and a second walled catheter tube adapted for delivering or draining fluids.

In yet another aspect or embodiment, the device employs a walled sampling chamber in communication with a sensing means, and a second walled catheter tube adapted
15 for delivering or draining fluids.

Optionally, when a non-temperature sensing-means is employed, a second sensing-means may be employed as well.

Although not originally thought to be feasible or
20 efficacious, the present invention in yet another embodiment has also accomplished improved accuracy and speed by the effective infrared sensor measurement of liquid or gaseous fluid parameters or compounds of interest, such as pO₂, pCO₂, pH_i, etc., admixed in a gaseous
25 sampling medium, preferably air. This was previously not believed to be possible due to the high gas volumes typically required for accurate infrared measurements, and because of erroneous measurements resulting from increased gas densities caused by higher tonometric sampling medium
30 pressures.

In view of all of the above, it will be appreciated that the tonometric method can now be modified in a fashion that provides the advantages of reduced equilibration time (with respect to saline) and without the
35 need to recalibrate the sensor *in vivo*, or remove it for recalibration. In the improved method, and very generally, air is employed as the sampling medium and sampling can be

done substantially continuously. The sampling medium air is aspirated from the walled sampling chamber of a tonometric catheter which has been inserted into the organ of interest (e.g., the gut). The $p\text{CO}_2$ of the aspirated sample is measured by employing a side-stream or main-stream, drift-free, non-dispersive infrared gas analyzer. The $p\text{CO}_2$ value obtained is then compared with either (1) the standard arterial bicarbonate value and/or (2) another direct or indirect measurement of a "global" or "systemic" physiologic value (e.g., arterial or venous pH; mixed venous bicarbonate; arterial oxygen saturation (e.g., as measured by pulse oximetry); arterial $p\text{CO}_2$; end tidal $p\text{CO}_2$; transcutaneous (TC $p\text{CO}_2$) $p\text{CO}_2$) in order to make a determination of the condition of the organ or if (A) a bicarbonate value must be obtained and/or (B) what, if any, clinical therapy or intervention may be necessary or appropriate with respect to oxygenation of the organ of interest.

In some embodiments, a Raman spectrometer may be employed, either in line or side stream, in place of the IR gas analyzer, as it will be appreciated by those skilled in the art that Raman spectroscopy offers distinct advantages over the more direct infrared-type measurements in certain applications.

A preferred indirect measurement of a "global" or "systemic" pH value is an end-tidal CO_2 value, or a transcutaneous CO_2 value.

However, the present invention has overcome these perceived and actual obstacles, and it successfully uses a gaseous sampling medium, such as air, along with known commercially available non-dispersive infrared spectrophotometry devices, resulting in high sample and measurement reliability, faster equilibration, thus allowing for faster and more frequent intermittent sampling, increased ease of use, and decreased sources of error, when compared to the prior use of a liquid sampling

medium (such as saline), and a blood gas analyzer, for example.

Those skilled in the art will readily recognize the kind of non-dispersive, infrared gas analyzing devices contemplated by the present invention. Examples of these devices are those commercially available and marketed by such companies as Datex Instrumentarium Corporation or Novamatrix Medical Systems, Inc., for example. Other examples of such devices and related equipment are discussed and disclosed in United States Patent Nos. 4,233,513; 4,423,739; 4,480,190; 4,596,931; 4,859,858; 4,859,859; 4,907,166; 4,914,720; 5,042,522; 5,067,492; 5,095,913, the disclosures and drawings of all of which are hereby incorporated by reference herein.

Non-dispersive infrared gas analyzers in general are typically manufactured in either "side-stream" or "main-stream" configurations. In one, a sample of a volume of gas is taken from a patient's gas flow (such as respiratory gas flow, a tonometric sampling chamber gas flow, or both) and conveyed through a sample tube to the infrared sensor and analyzer; in such a device, the sample is not typically returned to the patient's gas flow. The other common type is the so-called in-stream or main-stream type, which has a sensor apparatus that mounts directly within the patient's gas flow conduit and senses and takes measurements as the gas flows past the sensor.

In an embodiment of the present invention, a pressure transmitting chamber in the form of a balloon is disposed about a catheter and encloses a volume between the wall of the pressure transmitting chamber and the catheter wall. That volume is filled with a pressure transmitting fluid or medium such as air or saline solution. As the pressure in the transmitting chamber varies in reaction to changes in the patient's internal pressure, e.g. IAP, that change in pressure is transmitted by the pressure transmitting fluid via the lumen in the catheter to a point

outside of the patient for eventual monitoring and/or recordal by either medical personnel or machine.

In one embodiment, the pressure transmitting chamber communicates with a lumen inside the catheter via
5 a portal which allows the pressure transmitting medium, e.g. air or a saline solution, to pass through the portal as internal pressure of a patient's organ, e.g. IAP, bearing on the pressure transmitting chamber wall varies. The change in pressure on the pressure transmitting chamber
10 wall is thereby communicated through the pressure transmitting chamber, through the portal and into and through the lumen by a pressure transmitting medium. In the case of an open system such as a catheter hooked up to a manometer, a change in pressure on the pressure
15 transmitting chamber will cause a flow of pressure transmitting medium through the portal. By way of an example, an increase of pressure will compress the pressure transmitting chamber causing a change in volume. That change in volume is reflected by a flow of pressure
20 transmitting medium out of the pressure transmitting chamber through the portal and into the catheter lumen eventually resulting in the change in the fluid level in the manometer.

In the case of a closed system such as one with
25 a pressure transducer and saline solution as a pressure transmitting medium, a change in internal pressure may not produce a sizable change in volume of the pressure transmitting chamber, but rather compress the medium in the chamber with that change in pressure being transmitted
30 through the portal up through the catheter lumen to the pressure transducer. The pressure transmitting medium can transmit this pressure to a pressure sensing means such as a pressure transducer or a manometer located, for example, at the proximal end of the catheter. The pressure
35 transducer is used in the customary manner, known to those skilled in the art.

In an alternate embodiment, a pressure transducer can be installed in the pressure transmitting chamber and/or within the catheter lumen itself. A reading of the monitored pressure, e.g. the signal from the transducer, is transmitted through a portal communicating with the lumen and then through the lumen to a point outside the patient. There the signal from the transducer can be converted or translated, if necessary, to a machine or human readable form.

5 The invention can be used to monitor IAP by direct insertion into the gut by the use of conventional insertion techniques of a tonometric catheter. In one such use the invention is part of a nasogastric tonometer inserted into the stomach cavity. The pressure transmitting chamber is located in the organ or region where internal pressure is to be monitored.

10 The invention also encompasses the sensing of intra-abdominal pressure while at the same time measuring intramucosal pH or pHi such as pHi of the gut. In one embodiment of the invention the catheter containing the lumen via which pressure is monitored contains a separate lumen by which pHi is monitored as described elsewhere in the application. Another lumen may allow flow of the pressure transmitting medium or the pressure signal.

15 Alternately a single lumen may be shared for both pH and pressure monitoring.

The IAP may be monitored continuously while monitoring of the pHi is conducted. The preferred embodiment for this method is a catheter which comprises a pressure transmitting chamber communicating with a lumen for monitoring of the internal pressure and a separate walled sampling chamber to monitor pHi in fluid communication with the interior of a separate lumen within the same catheter.

20 In a separate embodiment of the invention a single chamber serves as both a pressure transmitting chamber for monitoring of internal pressure and as a

sampling chamber for monitoring of p_{Hi} . In such an embodiment the wall of the chamber, i.e. the combined pressure transmitting and sampling chamber, comprises a membrane which is relatively permeable to the fluid or gaseous component to be sampled but sufficiently non-permeable to the pressure transmitting medium so that there is minimal transmission of pressure transmitting medium from the chamber across the chamber wall. The pressure gradient and loss across the chamber wall is thereby minimized. In this embodiment, the pressure transmitting medium can be the same as a saline sampling medium used to monitor the p_{Hi} . The saline sampling medium will sample the concentration and/or composition of at least one liquid fluid or gaseous fluid which permeates the chamber wall. The saline sampling medium can also be used to charge the chamber and create an inflated chamber for use as the pressure transmitting chamber by insufflating the catheter lumen and chamber with a quantity of the saline sampling medium. The saline sampling medium can then act as the pressure transmitting medium for transmitting pressure to a pressure sensor. By this embodiment, the internal pressure of the patient as well as the p_{Hi} can be monitored for the same in vivo location. The monitored pressure can then be used to correct p_{Hi} readings based upon the predictable effects that changes in pressure will have upon the concentration of gases which determine p_{Hi} .

In an embodiment of the invention internal pressure monitoring is used in conjunction with the monitoring of p_{Hi} to arrive at a correction factor applied to the determination of the p_{Hi} . The correction factor allows adjustment of the p_{Hi} based upon changes in the pressure of the gas such as CO_2 which permeates a CO_2 monitoring chamber. In this embodiment, the concentration of CO_2 in the sampling chamber is monitored to arrive at p_{Hi} as discussed in this application. The pressure of the CO_2 in the sampling chamber will be a function of the pressure of the CO_2 outside of the sampling chamber. The pressure of

the CO₂ in the chamber will effect its concentration and hence the pHi. A change in the pressure of the CO₂ outside of the chamber will effect the pressure of the CO₂ measured inside the chamber. An increase in the internal pressure, 5 e.g. IAP, will be transmitted through the sampling chamber wall and effect a change in the pressure of the CO₂ inside the sampling chamber with a concomitant change in the monitored pHi. By monitoring internal pressure such as IAP while also monitoring for pHi, the calculation of pHi can 10 be adjusted based upon changes in internal pressure and the effect that those changes in internal pressures will have upon the gas pressures and hence concentration of measured gases.

For further understanding of the invention, its 15 objects and advantages, reference may be had to the following specification, the accompanying drawings, and the information incorporated herein by reference. Also, see commonly assigned applications serial no. 719,097, filed June 20, 1991; serial no. 994,721, filed December 22, 1992; 20 serial no. 014,624, filed February 8, 1993; and serial no. 08/035,020, filed March 22, 1993, all of which are completely and expressly incorporated herein by reference.

Brief Description of the Drawings

25 Figure 1 is a plan view of an embodiment of a tonometric catheter;

Figure 2A is a partial cross-sectional view of the tonometric catheter illustrating a first means for attachment of an electronic field effect transistor sensor;

30 Figure 2B is a partial cross-sectional view of the tonometric catheter illustrating a second means of attachment of the field effect transistor sensor;

Figure 3 illustrates the method of use of the tonometric catheter in measurement of the pH of the colon 35 and also of the stomach, the specific embodiment illustrated for colonic measurement being that of Figure 5

and the specific tonometric catheter for gastric measurement being that of Figure 4;

Figure 4 is another embodiment of the tonometric catheter with nasogastric tube;

5 Figure 4A is a cross-sectional view of the tonometric catheter of Figure 4 taken substantially along the line 4A-4A of Figure 4;

 Figure 4B is a cross-sectional view of the tonometric catheter of Figure 4 taken substantially along
10 the line 4B-4B of Figure 4;

Figure 5 is yet another embodiment of the tonometric catheter having multiple sensing/sampling portions;

 Figure 5A is a cross-sectional view of the
15 tonometric catheter of Figure 5, taken substantially along the line 5A-5A of Figure 5;

Figure 6 is a detailed view illustrating the tonometric catheter of Figure 4 in use within the stomach;

 Figure 7 is a detailed view illustrating the
20 tonometric catheter of Figure 5 in use within the colon;

Figure 8 is a similar view illustrating the tonometric catheter of Figure 1 in use within the colon;

 Figure 9 is an electrical schematic diagram illustrating one embodiment of electronic circuit in
25 accordance with the invention;

Figure 10 is an electrical schematic diagram illustrating another embodiment of the optical measurement of pH in accordance with the invention;

 Figure 11 is another embodiment of a tonometric
30 catheter with a urinary catheter;

Figure 11A is a cross-sectional view of the tonometric catheter/urinary catheter of Figure 11, taken substantially along the line 11A-11A of Figure 11.

 Figure 12 illustrates one preferred example of
35 the application of a tonometric catheter device, with remote sensing and recording apparatuses for monitoring and recording certain critical properties of interest.

Figure 13 is a diagrammatic representation of an exemplary non-dispersive infrared gas analyzer system usable in the present invention.

5 Figure 14 is a schematic representation of an alternate variation of a tonometric catheter usable in the invention.

Figure 15 is a diagrammatic representation of a manual syringe, modified to provide for sample pressure equalization usable in the present invention.

10 Figure 16 is a plan view of an embodiment of a pressure monitoring catheter with a cut away section.

Figure 16A is a partial cross-sectional view of the tonometric catheter illustrating the attachment of a pressure transducer.

15 Figure 17 is an embodiment of a catheter for internal pressure sensing with a nasogastric tube.

Figure 17A is a cross sectional view of the catheter of Fig. 17 taken substantially along line 17a-17a.

20 Figure 17B is a cross sectional view of the tonometric catheter of Fig. 17 taken substantially along line 17b-17b.

Figure 18 illustrates a catheter with dual use pressure sensing and pH sensing tonometric catheter.

25 Figure 18A is a cross-sectional view taken along line 18a-18a of Figure 18.

Figure 19 illustrates a tonometric catheter with a pressure sending apparatus installed.

Figure 20 is a chart showing the relationship of intra-vesical and gastric pressure in a subject patient.

30 Figure 21 is a plan view of a wound drain catheter modified to embody the invention.

Figure 21A is a side elevational view of the wound drain catheter of Figure 21 taken along line 21A-21A of Figure 21.

35

DETAILED DESCRIPTION

This application hereby expressly incorporates by reference, the disclosures and drawings of the following issued U.S. patents: United States Patent Nos. 4,221,567; 5 4,233,513; 4,273,636; 4,423,739; 4,576,590; 4,480,190; 4,596,931; 4,643,192; 4,671,287 4,859,858; 4,859,859; 4,907,166; 4,914,720; 5,042,522; 5,067,492; 5,095,913; 5,158,083; 5,174,290; and 5,186,172.

Figure 1 illustrates a first embodiment of 10 tonometric catheter 20. The tonometric catheter comprises a length of suitable tubing 22, one end 32 of which is closed, and the opposite end of which has a connector such as a luer-lock 24. Luer-lock 24 is adapted to receive a complementary fitting 26, which in turn couples through a 15 second length of tubing 28 to a three-way stopcock 30. Three-way stopcock 30 may be used to selectively connect tubing 28 to various sources of irrigation or aspiration.

Adjacent the closed end 32, tubing 22 is perforated as at 34. A balloon-like tonometric catheter 20 membrane 36 is fitted over the closed end so that the perforations 34 are enclosed, as illustrated. The tonometric catheter membrane 36 has an internal sleeve diameter at 38 which forms a tight fit with tubing 22. The preferred form of tonometric catheter membrane is 25 polydimethylsiloxane elastomer. The membrane may be sealed to the tubing 22 with appropriate adhesive so that the tonometric catheter membrane is sealed in a closed relationship to the outer wall of tubing 22, thereby forming a sampling chamber 40 adjacent closed end 32. The 30 tonometric catheter membrane has a certain elasticity to allow the membrane to expand when filled with an aspirating fluid in order to contact the wall of the organ under examination, as will be explained below.

The membrane 36 is preferably constructed such 35 that at least a portion of it is selectively permeable to the gas or fluid property of interest. In a preferred embodiment, it is selectively permeable to hydrogen,

oxygen, or H^+ , so that pH, pCO_2 , and/or pO_2 can be measured. It is also preferably impermeable to other materials that would interfere with the desired measurements, such as other gases, proteins, and the like. In a highly preferred
5 embodiment, an ion-selective membrane is employed.

Bonded to either the inner wall or the outer wall of tubing 22 are one or more sensors 42 for detecting a property indicative of pH and/or temperature. Two such sensors are illustrated in Figure 1, bonded to the outside
10 wall of tubing 22 with suitable adhesive. Figures 2A and 2B illustrate two alternate means of sensor attachment, Figure 2A illustrating the sensor attached to the inner wall of tubing 22 and Figure 2B illustrating the sensor attached to the outer wall of tubing 22.

In a preferred embodiment, at least a portion of the tubing, but not all of it, is made of a CO_2 impermeable material, such as those based on polyurethanes, PVC's, or polyester elastomers derived from the reaction of dimethylterephthalate 1,4-butanediol and α -hydro- Ω -
15 hydroxypoly (oxytetramethylene). In a highly preferred embodiment, this is a material such as Hytrel, sold by duPont.

For purposes of sensing temperature, thermistor devices are presently preferred. For sensing properties
25 indicative of pH chemically responsive field effect transistors or "Chemfets" may be employed. In this regard, Chemfet sensors 44 have been illustrated in Figures 2A and 2B. Chemfet sensor 44 comprises a field effect semiconductor device 46, which is encapsulated in a
30 solution impervious material 48, such as a polymerized epoxy resin. The encapsulation material 48 in turn may be encapsulated in a housing 50 (Figure 2A). Semiconductor device 46 is electrically coupled by bonding wires 52 to a terminal 54. Suitable electrical conductors such as
35 conductor 56 are attached to terminal 54 for electrically communicating between the Chemfet device 44 and the electronic circuitry described below in connection with

Figure 9. Conductor 56 is preferably routed through tubing 22 and exits through a sealed aperture at or near the luer-lock end of tubing 22, as at 58. A more detailed description of a suitable electronic sensor may be found in
5 U.S. Patent No. 4,020,830 to Johnson, entitled "Selective Chemical Sensitive FET Transducers," incorporated herein by reference. In order to allow a solution to contact the chemically sensitive surface of semiconductor device 46, tubing 22 may be provided with an aperture 60 when
10 implementing the embodiment of Figure 2A. Such an aperture is not needed in the embodiment of Figure 2B, since the semiconductor device 46 is exposed to sampling chamber 40 by virtue of the external mounting configuration.

The sampling chamber 40 can be filled with an
15 aspiration or sampling medium (gaseous or liquid) that is used to absorb or otherwise provide a means for incorporating and delivering or measuring the fluids or gases of interest. Such a medium is selected depending upon many factors, including the properties of the fluids
20 or gases of interest, the type of sensor 42 employed, and the type of calibration that is necessary. Such mediums include air, bicarbonate solutions, bicarbonate-buffered solutions, phosphate-buffered solutions and saline solution. It might be noted that gases often behave as
25 fluids and are therefore frequently considered to be fluids.

As noted above, when the sensor employed does not require frequent recalibration, the need for the sampling chamber 40 to be in communication with the proximate end of
30 the tonometric catheter (that remains outside the patient) may be eliminated since no aspiration is needed. However, in many instances such communication may still be desirable as aspiration may be required to calibrate the sensor or sensors, to replace the aspirating or sampling medium with
35 a fresh medium, and to incorporate the gas or gases of interest.

An embodiment of the tonometric catheter configured for the sensing of internal pressures, especially IAP, is illustrated in Figure 16. Elements in common with Figure 1 are given the same reference numerals.

- 5 The pressure sensing catheter 420 comprises a length of suitable tubing 22, one end 32 of which is closed and the opposite end of which has a connector such as a luer-lock 24. Luer-lock is adapted to receive a complimentary fitting 26 which in turn couples to a second length tubing 10 28 to a three way stopcock. Three way stopcock 30 may be used to selectively connect tubing 28 to various sources of irrigation, aspiration or to a pressure sensing means or monitor.

- Adjacent to closed end 32, tubing 22 is 15 perforated as at 434. A balloon like tonometric catheter membrane 436 is fitted over the closed end so that the perforations 434 are enclosed as illustrated. Pressure transmitting catheter membrane 436 has an internal sleeve diameter at 438 which forms a tight fit with tubing 22.
- 20 The preferred form of tonometric catheter membrane for pressure sensing applications is a relatively gas impermeable and saline impermeable material such as a conventional balloon used in angioplasty. The balloon wall can be made of a latex, silicone, polyurethane or 25 polyethylene. For certain applications a PVC material may also be used. Tonometric catheter balloons of the type suitable for use in the present invention are available from Tonometrics, Inc., Worcester, Massachusetts, U.S.A. The membrane may be sealed to tubing 22 with appropriate 30 adhesive known to the art such as silicone RTV type adhesive so that the tonometric catheter membrane is sealed in a closed relationship to the outer wall of tubing 22 therefore forming a pressure transmitting chamber 440 adjacent closed end 32. The pressure transmitting 35 tonometric catheter membrane has a certain elasticity to allow the membrane to expand when filled with a pressure transmitting medium such as air, or a saline solution. The

pressure transmitting medium allows the transmission of pressure changes as reflected by the movement of pressure transmitting tonometric catheter membrane 436 to be transmitted through the pressure transmitting chamber 440 and registered upon a pressure sensing means as explained below. As an alternative to the balloon-type chamber illustrated in Figure 16 a cuff-type chamber may be used, similar to the chamber on a pulmonary arterial catheter.

A membrane 436 is preferable constructed such that in one embodiment it is relatively impermeable to the pressure transmitting medium as well as to normally occurring gaseous and liquid fluids encountered *in vivo*. For another embodiment, the membrane 436 can be silicone which is permeable by CO₂ through diffusion *in vivo* but relatively impermeable to a saline solution used both for CO₂ sampling and pressure transmitting purposes.

The pressure transmitting medium is in pressure transmitting communication with the interior of tubing 22 such as through perforation 434. Tubing 22 is filled with pressure transmitting medium along its entire length. Tubing 28 is also filled with pressure transmitting medium throughout its length. By filled it is meant that the system from pressure transmitting chamber 440 through tubing 22 through tubing 28 up to pressure sensing means 460 is filled with a single uninterrupted volume of pressure transmitting medium, substantially devoid of discontinuities or impurities having different compressibilities than the pressure transmitting medium. By way of example, when the pressure transmitting medium is a saline solution, the system should be substantially devoid of gas bubbles or gas pockets which would inhibit the accurate transmission of pressure through the pressure transmitting medium from pressure transmitting chamber 440 to pressure sensing means 460. To monitor the temperature and temperature changes a thermistor may be located either within the pressure transmitting chamber or the catheter

lumen. Temperature readings can be used to correct changes in pressure based on temperature dependent effects.

Preferred embodiments for pressure sensing means 460 are pressure sensing transducers such as Gould
5 disposable model TXXR or a manometer such as a CVP manometer by Abbott Laboratories, North Chicago, Illinois, USA which is designed for intermittent or continuous monitoring of central venous pressure.

The preferred method of use of the tonometric
10 catheter of Figure 16 involves its insertion after assembly into the patient so that the pressure sensing chamber 440 is within the area to be monitored for pressure, e.g. the stomach. The tonometric catheter 20 is then charged with pressure transmitting medium in the case of a gaseous
15 pressure sensing medium. In the embodiment utilizing liquid pressure transmitting medium, such as a saline solution the system may be precharged before insertion with pressure transmitting medium so as to be substantially free of gaseous impurities which may diminish the systems
20 ability to accurately transmit pressure changes throughout the system. The system should not be charged before insertion to a pressure which would significantly expand chamber 440 and hinder insertion.

In an embodiment utilizing a gaseous pressure
25 transmitting medium, the system is charged after insertion by inflation of the balloon with gas such as via stopcock portal 462 so as to inflate pressure transmitting chamber 440 sufficiently to allow pressure transmitting tonometric catheter membrane 436 to be impinged upon and transmit
30 changes in pressure to the pressure transmitting medium in chamber 440. The chamber should not be over inflated so as to add unnecessary volume to chamber 440 which may add unwanted pressure to the internal organs of the patient. In adult patients it has been found that inflation of the
35 system of up to three milliliters of gaseous pressure transmitting medium has provided satisfactory results. The volume of medium inflated should be carefully measured.

Should inflation be necessary the same volume may be inflated allowing comparisons of pressure readings. By having the same volume in the systems, comparisons can be made according to the gas law $PV = nRT$. Once the system
5 has been inflated and stopcock opening 462 sealed, pressure sensing means 460 may be calibrated and/or zeroed. Changes in internal pressure of the patient act upon membrane 436 which in turn alters the volume and/or pressure of chamber 440. The change in volume or pressure of chamber 440 is
10 transmitted by pressure transmitting medium upon the pressure sensing means 460 which will thereby reflect changes in the internal pressure of the patient.

The pressure sensing means 460 need not be operably connected to the stopcock and can be connected
15 anywhere along the system where it is in pressure sensing communication with the pressure transmitting medium. In one embodiment pressure sensing means 460 may be connected to the luer-lock 24 after the system has been charged with pressure transmitting medium.

20 In an alternative embodiment also illustrated in Figure 16 by way of phantom line additions, pressure sensing means may be placed within the system such as by way of a pressure transducer 442 placed within the pressure transmitting chamber 440. Turning to Figure 16A, pressure
25 transducer 442 would then be operably connected to conduit 56 so as to communicate the pressure transducer's signal via conduit 56 outside of the system via aperture 58 and then to a proper translation and/or display apparatus (not shown). Aperture 58 should be such that pressure sealing
30 contact with conduit 56 is maintained by tubing 22 so that tubing 22 may be charged with pressure transmitting medium without leakage at aperture 58. In a preferred embodiment, pressure transducer 442 is a microelectronic device whose output is communicated via a conduit 56 in the form of an
35 electrically conductive wire.

The preferred method of use of the pressure sensing catheter is to have the catheter assembly completed

before insertion into the patient. The catheter is inserted via conventional nasogastric techniques into the gut of the patient. The catheter assembly in the proper configuration can be utilized for monitoring pressure in a variety of locations. These locations include the stomach, the large and/or small intestines and the rectum. In addition, the pressure sensing chamber may be located so as to rest against the surface of an organ such as the liver or kidney to monitor the pressure exerted upon that organ to assist in the diagnosis of the effect of pressure upon that organ's function. In addition, the system could be used to monitor esophageal pressure. In general, the system can be used to monitor whether or not the various sphincters throughout the body are working to seal off chambers by monitoring, e.g. pressures upon either side of the various sphincters. Areas other than the gut can be monitored via this apparatus and method including intercranial pressure.

For certain monitoring applications, the configuration of the catheter can be changed, for example the pressure sensing chamber may be applied for use on a feeding catheter. A pressure sensing chamber could also be installed on a wound drain catheter such as a Jackson-Pratt catheter where the pressure sensing chamber was resting against the liver. In such an embodiment a single catheter can be simultaneously draining as well as performing pH, pressure, and/or temperature monitoring.

Figure 21 shows a closed wound drainage catheter modified to also monitor internal pressure and/or pH. Tubing 522 has a plurality of internal lumens. One internal lumen 524, in communication with the drainage area 528, can be hooked up to a conventional drainage reservoir for a closed wound drainage system such as the Jackson-Pratt closed wound drainage system supplied by American Hospital Supply Corporation as is well known to those skilled in the art. Lumen 524 is in fluid communication with wound drainage apertures 530. Turning to Figure 21A,

lumen 524 is non-communicating with lumen 526. Lumen 526 communicates with combined pressure transmitting and sampling chamber 540. Chamber wall 542 is preferably made of a silicone membrane which allows for diffusion of CO₂ for measurement of pHi as discussed above. Lumen 526 will therefore communicate with both a pressure sensing means such as a manometer or pressure transducer as well as sensor means to monitor and calculate the pHi of the subject. A single medium such as saline solution can serve as both a sampling medium and a pressure transmitting medium. The flat surface of the bottom of the chamber 540 allows the membrane 542 to lie against an organ, such as the liver. While wound drainage occurs the pressure and pHi of the monitored liver surface can be monitored.

Pressure may be continuously or intermittently monitored. The catheter may be left in the patient for a period of time and the pressure continuously monitored such as by using a plotter to record a graph of pressure over time. Alternately the pressure readings may be taken at discrete time intervals, e.g. every 30 minutes. Use of the described techniques and apparatus in the stomach can avoid transcutaneous monitoring of pressure allowing longer term monitoring while avoiding invasive techniques and their attendant risks such as infection.

The use of a pressure transmitting chamber on a catheter can also serve as a positioning mechanism. The pressure transmitting chamber can be inserted into the patient and its location confirmed through various techniques such as its inflation and monitoring of the pressure, manipulation of the patient such as manually applying pressure to a selected location and monitoring any result in increase in pressure or auscultation of air as it is being insufflated. The use of the pressure monitoring system as a position mechanism may eliminate the need for an x-ray to confirm the location of either the pressure transmitting chamber or other apparatus on the same catheter.

In the preferred method, the catheter is inserted during a period of quiescent motor activity with no evidence of the migrating motor complex. Certain phases of the migrating motor complex are characterized by rapid convulsions of the stomach and/or intestine which reduce the effectiveness of the pressure monitoring apparatus. In certain situations, the pressure monitoring apparatus can be used to monitor the phase of migrating motor complex. During the phases that have rapid convulsions, the pressure readings can be run through a signal detection algorithm where rapid fluctuations and peaks trigger a reading that a certain phase of the complex is present. The algorithm can be programmed to either mark certain pressure readings as unusable due to excessive motor activity or be used itself as a monitor of motor activities.

Another embodiment of the tonometric catheter is illustrated in Figures 4, 4A and 4B. As illustrated, the tonometric catheter is appropriately configured to also serve as a nasogastric sump, either with or without gastric suction. With reference to Figure 4, the tonometric catheter 20a comprises a multipassage tubing 62 which defines three individual noncommunicating (between each other) passageways or lumens, an air lumen 64, an optional suction lumen 66 and a tonometric catheter lumen 68. A tonometric catheter membrane, similar to that previously described, is attached at an intermediate location on tubing 62, allowing a portion of the tubing to extend beyond the end of membrane 36 to define the nasogastric sump 70. Tubing 62 is provided with a plurality of perforations 72 which communicate between tonometric catheter lumen 68 and the sampling chamber 40 defined by membrane 36. If desired, one or more sensors 42 can be included in accordance with the above teachings, in which case a suitable conductor 56 may be routed through tonometric catheter lumen 68 to exit at sealed aperture 58.

The nasogastric sump portion 70 is suitably provided with a plurality of openings 74 through which the stomach may be aspirated.

At the opposite end of tubing 62 the tubing
5 splits to form three separate connections. Air lumen 64 communicates with air lumen passageway 76, suction lumen 66 connects with suction lumen passageway 78 and tonometric catheter lumen 68 communicates with tonometric catheter lumen passageway 80. The tonometric catheter lumen
10 passageway is fitted with three-way stopcock 30, similar in function and purpose to the three-way stopcock 30 described in connection with Figure 1. If desired, a quick connect fitting 82 may be used to couple the suction lumen
15 passageway 78 with an aspiration source. As illustrated, the quick connect fitting preferably has angularly cut ends and a slightly enlarged midsection, making it easy to insert into the end of passageway 78 and also into the aspiration hose coupling (not shown). The enlarged
20 midsection helps form a seal with the adjoining passageways. Preferably the quick connect fitting is fabricated of disposable plastic.

Turning to Figure 17 another embodiment of a pressure sensing tonometric catheter is illustrated for sensing the internal pressure such as the IAP of a patient.
25 The catheter illustrated in Fig. 17 is configured to also serve as a nasogastric sump either with or without gastric suction. Identical elements of the tonometric catheter of Figure 4 are identified with the same reference numerals. Multi-passage tubing 62 defines three individual non-
30 communicating (between each other) passageways or lumens, air lumen 64, an optional suction lumen 66, and a pressure transmitting medium lumen 468. A tonometric catheter membrane, similar to that previously described with
reference to Figure 16 is attached in an intermediate
35 location in tubing 62, along a portion of the tubing so as to allow tubing 62 to extend beyond the end of membrane 436 to define the nasogastric sump 70. Tubing 62 is provided

with a plurality of perforations 72 which communicate between pressure transmitting lumen 468 and the pressure transmitting chamber 440 defined by membrane 436. If desired, one or more pressure transducers or sensors 42 can
5 be included in accordance with the above teachings, in which case a suitable conductor or conduit 456 may be routed through pressure transmitting lumen 468 to exit at sealed aperture 58. Conductor 456 can be figured so as not to compromise pressure transmitting lumen 468 in its
10 ability to allow a flow of pressure transmitting medium through lumen 468.

The nasogastric sump portion 70 is suitably provided with a plurality of openings 74 through which the stomach may be aspirated.

15 At the opposite end of the tubing 62 the tubing splits to form three separate connections. Air lumen 64 communicates with air passageway 76. Suction lumen 66 connects with suction lumen passageway 78 and pressure transmitting lumen 468 communicates with pressure
20 transmitting lumen passageway 80. The pressure transmitting lumen passageway 80 is fitted with a three way stopcock 30 similar in function and purpose to the three way stopcock described above especially in connection with Figure 16.

25 In use the pressure transmitting lumen 468 is purged of potential contaminants and either precharged with pressure transmitting medium before insertion or preferably charged after insertion. Once pressure transmitting lumen 468 is properly charged along with pressure transmitting
30 passageway 80, pressure sensing means 460 may be operably associated via the stopcock 30 with pressure transmitting passageway 80 so as to register changes in the IAP of the patient. Changes in the internal pressure, such as the IAP, of the patient will be communicated to pressure
35 sensing means 460 in the manner described in connection with Figure 16. Alternately to pressure sensing means 460, pressure transducer or sensor 42 may be operably associated

with pressure display means 462 via conduit or conductor 56 to display changes in the patients internal pressure. Pressure display means 462 may be of the type known to those skilled in the art such as a cathode ray tube which
5 can display in graphic or numerical format or a plotter which will graph on paper in X-Y coordinates.

Yet another embodiment of the tonometric catheter is illustrated in Figures 5 and 5A. This embodiment is a multiple tonometric catheter embodiment employing a
10 tubing 84 having a plurality of passageways or lumen as shown in the cross-sectional view of Figure 5A. Specifically, tubing 84 includes an air lumen 86a which communicates with the endmost tonometric catheter 36a and three additional tonometric catheter lumens 86b, 86c and
15 86d, which communicate respectively with tonometric catheters 36b, 36c and 36d. As with the other embodiments, each tonometric catheter may be provided with one or more sensors such as sensors 42. A radiopaque tungsten plug 88 is positioned within each of the three tonometric catheter
20 lumen 86b, 86c and 86d adjacent the distal end of each tonometric catheter, serving to block the remainder of the tonometric catheter lumen passageway and thereby ensuring that fluid pressure introduced into each tonometric catheter lumen will cause the associated tonometric
25 catheter to balloon outwardly as required during use. Similarly, a radiopaque tungsten rod 90 is fitted as a plug in the end of air lumen 86a, serving to terminate the end of the air lumen passageway. Being radiopaque, the tungsten plugs and tungsten rod aid in properly positioning
30 the tonometric catheters by being visible under fluoroscope or x-ray. In addition, if desired, tubing 84 can be provided with a radiopaque stripe along all or part of its length.

At the proximal end of tubing 84 the lumen 86a-
35 86d diverge to define four separate tubes 92a-92d. Each tube is fitted with a three-way stopcock similar to those described above. Each sampling connector may optionally be

coded numerically by color, etc. While four approximately equally spaced tonometric catheters have been illustrated in Figure 5, it will be understood that the invention can be modified to include a greater or fewer number of tonometric catheters at different spacing as required for a particular application. It will also be understood that some or all of the tonometric catheters can include one or more sensors coupled to conductors 56, each preferably routed through the corresponding lumen passageway.

Referring now to Figure 19, an alternate embodiment of the invention is illustrated. With reference to Figure 5, the same elements are given the same reference numerals as in Figure 19. The device of Figure 5 has been modified as shown in Figure 19 to include means for sensing internal pressure of a patient such as IAP. In this embodiment, a multiple tonometric catheter is shown employing tubing 84 having a pluralities of passageways or lumens which do not communicate with one another as shown in the cross sectional view of Figure 19A. Specifically, tubing 84 includes an air lumen 86 which communicates with the end most tonometric catheter 36A and two additional tonometric lumens 86b and 86c which communicate respectively with tonometric catheters 36b and 36c. As explained above, each tonometric catheter may be provided with one or more sensors such as sensors 42. A radiopaque plug 88 is positioned within each of the tonometric catheter lumens 86b and 86c adjacent the distal end of each tonometric catheter serving to block the remainder of the tonometric catheter lumen passageway and thereby insuring that the fluid pressure introduced into each tonometric catheter lumen will cause the associated tonometric catheter to balloon outwardly as required during use.

Tubing 84 includes pressure transmitting lumen 486 which communicate with pressure transmitting chamber 440 created by membrane 436 to create a pressure sensing tonometric catheter similar to that described in Figure 16.

At the proximal end of tubing 84, pressure transmitting lumen 486 diverges from lumens 86a, 86b, and 86c. As described with respect to other embodiments the pressure sensing tonometric catheter may be operably
5 connected to a pressure sensing means 460 via pressure transmitting lumen 486 by means of a pressure transmitting medium. Alternatively, a pressure transducer 442 may be located within the pressure transmitting chamber 440 and operably connected with a signal converting and display
10 means 462 via conductor 456. The radiopaque plug 88 is positioned in pressure transmitting lumen 486 adjacent the distal end of the pressure transmitting chamber.

Referring now to Figure 9, a suitable electronic monitoring circuit will now be described. In Figure 9
15 CHEMFET semiconductor device 46 has been shown schematically by the equivalent circuit model enclosed in dotted lines. The device 46 thus comprises drain electrode 150, source electrode 152 and reference electrode 154. The chemically selective system, such as a
20 membrane system is depicted diagrammatically at 156. The substrate is grounded as at 158.

Source electrode 154 is coupled to an input lead of operational amplifier 160 which includes feedback network diagrammatically depicted at 162. Operational
25 amplifier 160 senses the drain source current flowing through device 46 and converts this signal into a voltage signal which is output on lead 164. The drain source current changes in accordance with changes in the chemical system under test. More specifically, as the pCO_2 level
30 changes in the fluid exposed to device 46, the drain source current changes accordingly. Hence the output voltage signal on lead 164 is likewise an indication of the pCO_2 level of the organ under test. This voltage signal on lead 164 is coupled to an input of comparator 166 which
35 also receives a reference voltage V_{ref} , which may be supplied using a voltage divider network (not shown) or which may alternatively be provided by a digitally controlled voltage

source 168. The output of comparator 166 is fed to reference electrode 154 to provide a stable reference bias voltage. If a digitally controlled voltage source is used, this reference voltage can be adjusted and calibrated
5 by a computer circuit yet to be discussed. The voltage signal on lead 164 is also fed to an analog to digital convertor 170, which is in turn coupled to a microprocessor-based microcomputer 172.

In order to automatically determine the pH of the
10 wall of the hollow viscous organ under test, a separate gas analyzer sensor 174 is used to determine the bicarbonate concentration in the arterial blood of the patient. The output of sensor 174 is coupled through analog to digital convertor 176 to microcomputer 172. Microcomputer 172 is
15 preprogrammed to determine or calculate the pH of the organ wall using the values provided by analog to digital convertors 170 and 176. Conversion of pCO_2 measurements can be converted into pH measurements automatically by microcomputer 172 using various equations and references
20 disclosed herein or others well-known in the art.

Although many different types of output devices may be employed, strip chart recorder 178 and CRT monitor 180 have been illustrated. Strip chart recorder 178 and monitor 180 are coupled as output devices
25 to microcomputer 172. Strip chart recorder 178 offers the advantage of developing an easily readable, permanent record of the fluctuations in organ wall pH. Monitor 180 offers the advantage of providing digital readout of the pH value as well as displaying the upper and lower excursions
30 of pH fluctuation. If desired, microcomputer 172 can be instructed and/or preprogrammed using keyboard 182 to compare the instantaneous pH value with doctor-selected upper and lower alarm limits. If the measured instantaneous pH fluctuates outside those limits,
35 microcomputer 172 can sound an alarm to alert hospital staff.

While a single semiconductor device 46 has been illustrated in conjunction with the electronic circuit of Figure 9, the circuit may be readily adapted for use with a plurality of semiconductor devices in order to measure the pH at different locations substantially simultaneously. In such an embodiment, the data coming from each sensor can be fed to a separate I/O port of microcomputer 172. In the alternative, a single I/O port can be used with the individual input signals being time multiplexed.

As an alternative to electronic pH sensors, the invention may also be practiced using optical sensor technology. Referring to Figure 10, the presently preferred optical sensor embodiment uses a first fiber optic cable 94 which is optically coupled through a series of lenses 96, selectable color filters 98 and heat absorber 100 to an illumination source 102, such as a 100 watt tungsten-halogen lamp. Fiber optic cable 94 is routed through the tonometric catheter lumen in a fashion similar to the conductor 56 of the above-described embodiments, with the end thereof protruding through the tubing and into the sampling chamber 40. A second fiber optic cable 104 is routed parallel to the first fiber optic cable 94, with one end protruding through the tubing and held in place adjacent the end of first cable 94 with a collar 106. Collar 106 may be adhesively bonded to the outside wall of the tubing. The opposite end of second fiber optic cable 104 is positioned for optically coupling with a phototransistor 108 which is electrically connected to an operational amplifier circuit 110. The operational amplifier circuit can be coupled to an analog to digital converter, such as A/D converter 170 of Figure 7.

In use, fiber optic cable 94 illuminates a region within the sampling chamber 40 which is filled with a sampling fluid containing a colorimetric pH indicator. The illumination from fiber optic cable 94 reflects from the molecules suspended in the pH indicator solution, with some of the reflected illumination passing back through second

fiber optic cable 104 to the phototransistor. By selecting the appropriate filter 98, a monochromatic illumination or illumination of otherwise known spectral content is employed to illuminate the colorimetric pH indicator solution. When the color of the filtered illumination matches that of the indicator, the illumination is absorbed and a low illumination signal is received at the phototransistor. When a pH change causes a color change in the indicator away from the color of the filtered illumination, more illumination is reflected back to the phototransistor, with an attendant increase in detected signal output. In this fashion, the proper selection of indicator dye and illumination filtration can be used to detect pH ranges. For a further description of fiber optic pH sensor technology, refer to G. G. Vurek "A Fiber Optic pCO₂ Sensor," Annals of Biomedical Engineering, Vol. 11, pp. 499-510, 1983, which is available from Pergamon Press, Ltd., and is expressly incorporated herein by reference.

While the preferred embodiments have been disclosed in connection with monitoring of the gastrointestinal tract and the urinary and ureteric tracts it will be appreciated that its principles are applicable to other hollow internal organs to monitor pH and hence perfusion of those organs. Also while several presently preferred detailed constructions for tonometric catheters have been disclosed, it will be appreciated that other constructions may be developed which are equally suitable. The disclosed constructions are presently preferred for the reason that they are readily fabricated using existing available materials. Other embodiments may include other, but equivalent materials for the tonometric catheter membrane and/or connective tubing. They may also differ in the specific fabrication details. As an example, the sampling chamber may be eccentric rather than symmetric about the connective tubing.

In still another embodiment, conventional gas analyzers may be employed externally. A device such as

that shown in Figure 1 may be used in combination with a pump or aspiration means (not shown) for continuous or regular intermittent aspiration of a sample of the aspirating liquid or medium that is used to fill the sampling chamber 40. The sample removed by pump or aspiration means via attachment to the luer-lock 24 can be optionally designed so that the sample aspirated at each sampling interval can be brought in contact with an exterior, separate gas analyzing means or sensor (not shown) to determine the pH, pO_2 , pCO_2 , and/or the like, of the sample. Such automatic sampling can be conducted employing a system as shown in Figure 12. In the assembly a sampling system employs a personal computer to conduct evaluations and analysis of the samples withdrawn from the tonometric catheter 299.

Pump 203 is loaded with the sampling or aspirating medium such as saline. Next, valve 201 is activated to withdraw a desired amount of the sampling fluid. The valve 201 is deactivated and pump 203 is used to enforce the sampling chamber of the tonometric catheter 299 using a calibrated amount or optionally a pressure transducer 215. The sampling fluid or medium is allowed to come to equilibrium with the wall of the organ or area of interest. Next the "dead space," i.e., the area of the lumen filled with the sampling fluid that is not in equilibrium, is removed by activating valve 205, activating pump 207, activating valve 209 and infusing pump 207; the waste 219 is discarded. A sample for analysis is then withdrawn by deactivating valve 209, activating pump 207 to then deliver the sampling to a gas analyzer such as an infrared or Raman gas analyzer (not shown) that provides data from the sample to the PC 217, and the evaluation is conducted as described herein.

The sample gas analyzer or a separate gas analyzer may be employed to determine the bicarbonate concentration in the arterial blood of the patient, as described above.

Another embodiment of the tonometric catheter is illustrated in Figures 11 and 11A. As illustrated, the tonometric catheter is appropriately configured to also serve as a urinary or ureteric catheter, either with or without suction, which optionally employs sensors. With reference to Figures 11 and 11A, the tonometric catheter 220 comprises a multipassage tubing 262 which defines three individual noncommunicating (between each other) passageways or lumens, an optional air or irrigation lumen 264, a drainage or suction lumen 266 and a tonometric catheter lumen 268. A tonometric catheter membrane, similar to that previously described, is attached at a distal location on tubing 262, allowing an intermediate portion of the tubing not extending beyond the end of membrane 236 to define the urinary or ureteric catheter 270. Tubing 262 is provided with a plurality of perforations 272 which communicate between tonometric catheter lumen 268 and the sampling chamber 240 defined by membrane 236. If desired, one or more sensors 242 can be included in accordance with the above teachings, in which case a suitable conductor 256 may be routed through tonometric catheter lumen 268 to exit at sealed aperture 258.

The urinary catheter or ureteric catheter portion 270 is suitably provided with a plurality of openings 274 through which the bladder or ureters may be aspirated or irrigated.

At the opposite end of tubing 262 the tubing splits to form three separate connections. Air or irrigation lumen 264 optionally communicates with air lumen passageway 276, urinary lumen connects with suction or drainage lumen passageway 278 and tonometric catheter lumen 268 communicates with tonometric catheter lumen passageway 280. The tonometric catheter lumen passageway is fitted with three-way stopcock 230, similar in function and purpose to the three-way stopcock 30 described in connection with Figure 1. If desired, a quick connect fitting 82 as seen in Figure 4 may be used to couple the

suction urinary passageway 278 with an aspiration source. As illustrated, the quick connect fitting preferably has angularly cut ends and a slightly enlarged midsection, making it easy to insert into the end of passageway 278 and
5 also into the aspiration hose coupling (not shown). The enlarged midsection helps form a seal with the adjoining passageways. Preferably the quick connect fitting is fabricated of disposable plastic.

Yet another embodiment of the urinary
10 catheter/tonometric catheter combination illustrated in Figures 11 and 11A may employ a multiple tonometric catheter embodiment employing a tubing having a plurality of passageways or lumens as shown in the cross-sectional view of Figure 5A.

15 In another embodiment of the present invention, a tonometric catheter may be adopted to deliver a pharmaceutically-active agent, either for systemic, local or topical activity, or a combination thereof. For example, an additional lumen may be added such as that and
20 for irrigation or aspiration, to deliver the active. For example, the irrigation/aspiration lumen 264 shown in Figure 11 and 11A, may be used to deliver an active agent. In another embodiment, a portion of the device may be modified so as to provide sustained release of the active
25 agent of interest.

Thus, for example, the problems of nosocomial infection associated with catheter insertion can be overcome by incorporating an antimicrobial into at least a portion of the polymeric material used to manufacture the
30 tonometric catheter, or by coating at least a portion of the device with a sustained release composition, or by delivering the antimicrobial via the tonometric catheter. Such modifications are well known to those skilled in the art. See U.S. Patent No. 4,677,143, incorporated herein by
35 reference.

Classes of useful agents include antimicrobial agents, nonsteroidal anti-inflammatory agents, topical

anesthetics, topical vasodilators, metabolic suppressants, and other agents that could be delivered for absorption at the sites of the tonometric catheter.

In another embodiment as shown in Figure 18, the catheter is appropriately configured to monitor both pH and IAP. Pressure transmitting medium also serves as sampling fluid which may be charged into sampling chamber 40 so as to cause it to act as a pressure transmitting chamber. Once the system is charged, pressure sensing means 460 may be operably connected to stopcock 30 to monitor IAP.

Sensor 42 detects a property indicative of pH and/or temperature as discussed above. Electronic pressure transducer 442 can also be mounted on the catheter within the chamber 40 for sensing pressure and communicating its output via conductor 456 running through the lumen 68 of the catheter 22 as shown in 18A. The output of the pressure transducer 442 can be communicated via conductor 456 to a pressure display means such as an X-Y plotter or cathode ray tube.

In lieu of an internal pressure transducer 442 communicating via conductor 456, the chamber 40 can be charged with the sampling fluid which will also serve as the pressure transmitting medium. The pressure transmitting medium is in fluid communication with lumen 68 via opening 34 and can be monitored by a pressure sensing means such as a manometer or pressure transducer at a location external to the patient such as via one opening of the stopcock 30.

Where a single balloon is used as both a sampling chamber and as a pressure transmitting chamber, the membrane 236 should be selected to be permeable by the element which is to be sampled but relatively impermeable to the pressure transmitting medium.

In numerous clinical settings it is now common to monitor the carbon dioxide concentration of the arterial blood of patients, particularly those who are critically

ill or under anesthesia; this measurement has been determined to bear a usually predictable relationship to pH. One of the most common non-invasive techniques for measuring arterial CO₂ is doing so indirectly by measuring the CO₂ concentration of the last gas expired from a patient (so called "end tidal") during normal respiration. The arterial CO₂ concentration is then calculated by employing the known correlation between the end tidal pCO₂ and pCO₂ of the arterial blood.

It has been discovered in another aspect of the present invention that end tidal CO₂ (as well as the underlying correlation between end tidal CO₂ and the pCO₂ of arterial blood) may also be useful in making clinical determination of the condition of an organ of interest when the end tidal CO₂ is compared and contrasted with the pCO₂ of air aspirated from a tonometric catheter having a walled sampling chamber inserted into an organ of interest. These measurements having the added convenience of both being measurable by IR or Raman gas analyzers.

However, in order to fully appreciate this, a detailed understanding of the general tonometric method is useful. This background is helpful primarily for the skilled artisan to fully appreciate the relationship of moving from the general tonometric method (which employs pCO₂ associated with the wall of the organ of interest and the bicarbonate concentrations of arterial blood) to even more indirect but useful measurements.

In accordance with one preferred embodiment of the present invention, the condition of an organ of interest is determined in a patient in need of such determination when the pCO₂ associated with the wall of the organ of interest is sampled and compared to substantially contemporaneous arterial or venous pCO₂ values or, in a highly preferred embodiment, end tidal pCO₂ value(s); the pCO₂ of the wall of the organ may also be compared to: venous or arterial pH; mixed venous bicarbonate values;

transcutaneous pCO_2 ; arterial oxygenation (saturation), and the like.

While not intending to be bound by theory, the following is offered to put these aspects and embodiments
5 of the present inventions in proper context.

The assumptions upon which the indirect measurement of intramucosal pH (pHi) are based are valid in normally perfused tissues. In these circumstances, the indirect measurement of intramucosal pH is identical to
10 that measured directly in the submucosal space with a microprobe.

The indirect measurement of pH falls in parallel with the pH made directly in the submucosal space when an intramucosal acidosis is induced by endotoxemia, low-flow
15 or no-flow. In those circumstances in which the intramucosal acidosis is induced by endotoxin and flow to the gut is maintained at control levels the measurements are in close agreement ($r=0.945$). When induced by low-flow and especially no-flow the indirect measurements
20 underestimate the severity of acidosis present in the submucosal space. The disparity between indirect and direct measurements observed in low-flow and no-flow states disappears when blood flow is reestablished and the pH is allowed to return towards normality. Inspection of the 20
25 minute values obtained in Antonsson et al's study reveals that the degree of dissociation observed between indirect and direct measurements is a linear function of the rate of change in intramucosal pH induced.

An additional primary assumption upon which the
30 validity of the tonometric measurement of the adequacy of tissue oxygenation is that the bicarbonate concentration in tissue fluid is the same as that being delivered to it in arterial blood. It has been postulated that the dissociation between calculated and measured pH in low-flow
35 and especially no-flow states may be due to a dissociation between arterial and interstitial bicarbonate induced by the buffering of metabolic acids by tissue bicarbonate.

The hypothesis does not account for the law of mass action which dictates that the fall in bicarbonate concentration induced by the addition of a fixed acid load to a "closed system" from which CO_2 cannot escape, such as the extracellular fluid compartment, is inhibited by the accumulation of CO_2 . The addition of even large amounts of fixed acid to a "closed system" does not produce a significant reduction in bicarbonate concentration but does produce a significant rise in pCO_2 . A fall in bicarbonate occurs only when venous blood enters the pulmonary circulation, an "open system" from which the CO_2 added to the venous blood by the buffering of the fixed acid load in the dysoxic tissue bed is able to escape. The fall in arterial bicarbonate thus induced causes the tissue bicarbonate to fall by equilibration with the lowered bicarbonate concentration in arterial blood returning to the tissue bed. The fall in arterial bicarbonate induced by the escape of CO_2 from the lungs cannot cause a reduction in tissue bicarbonate concentration in a no-flow state for it is unable to enter the tissue bed.

The tissue bicarbonate should be the same as that in arterial bicarbonate perfusing the tissue bed in all circumstances except perhaps very transiently after a sudden and large change in arterial bicarbonate induced by an intravenous bolus of bicarbonate or sudden change in pulmonary ventilation.

As a precaution, however, it is wise to wait until the arterial bicarbonate has been stable for some 10 to 15 or better yet 30 minutes before measuring the intramucosal pH after an intravenous bolus bicarbonate or sudden changes in ventilation regimes.

It is therefore suggested that the primary assumption upon which tonometric measurement of intramucosal pH is based, namely that the tissue bicarbonate is the same as that in arterial blood, is valid in many relevant clinical settings, including those in which the dissociation between measured and calculated

intramucosal pH was greatest. The indirect measurement of intramucosal pH appears to be an accurate measure of the pH in interstitial fluid in the most superficial layers of the intestinal mucosa especially in those circumstances in which the measurement is of greatest value, namely patients who appear by all conventional criteria to be adequately resuscitated. The only circumstance in which the measurement might be inaccurate for an extended period is a no-flow state. In this circumstance, the indirect measurement is so abnormal that the presence of the intramucosal acidosis should not be missed even if there is a large discrepancy between actual and assumed measurements. Transient inaccuracies may be expected following an intravenous bolus of bicarbonate or sudden change in pulmonary ventilation.

Stoichiometric Analysis of Determinants of Tissue Acidosis

During aerobic metabolism the pH of tissue fluid is determined by the bicarbonate concentration in tissue fluid, the CO_2 released by oxidative phosphorylation, and the balance between ATP hydrolysis and resynthesis. In gastric glands the intracellular pH is the same as the extracellular pH in acidotic states. The pH of the extracellular fluid (ECF) is determined by the amount of metabolic acid present and the ability of the ECF to buffer the acid. The pCO_2 attained following the buffering of a volatile (H_2CO_3 from oxidative phosphorylation) or fixed acid load (protons from ATP hydrolysis) in a closed system, (Fig. 5) such as the ECF, may be calculated in the manner described by Gattinoni and Feriani.

In normoxic tissues 6 mmol of CO_2 are produced for every mmol of glucose consumed in the generation of 38 mmol ATP. 13.5% of a volatile carbonic acid load added to ECF remains after being buffered by proteins and determines the pCO_2 present in the ECF. Assuming that the bicarbonate concentration in ECF is 25 mEq/l the metabolism of one mM glucose is 27 mmHg ($6 \times 0.135/0.03$). In normoxic and

resting healthy subjects with a tissue bicarbonate of 25 mEq/l the $p\text{CO}_2$, determined tonometrically, is 40 mmHg and the intramucosal pH 7.40. If it is assumed that the protons released by ATP hydrolysis are exactly balanced by the protons consumed by ATP resynthesis in oxidative phosphorylation then the aerobic metabolism of 1.48 mM glucose is required to generate the volatile carbonic acid necessary to attain the $p\text{CO}_2$ of 40 mmHg ($27 \times 1.48 = 40\text{mmHg}$) and pH of 7.40 found in normoxic ECF when the tissue bicarbonate concentration is 25 mEq/l.

The $p\text{CO}_2$ attained from the buffering of the volatile acids released into normoxic ECF in a tissue bed should increase as the metabolic rate increases, the increased demand for oxygen in the absence of replenishment by flowing blood being met exclusively by an increase in oxygen extraction ratio. A rise in metabolic rate of the magnitude seen in an exercising athlete, which may be as great as 900%, can be expected to cause a rise in equilibrium $p\text{CO}_2$ and hence fall in intramucosal pH in normoxic tissues. The magnitude of the fall in pH induced by the rise in $p\text{CO}_2$ is offset by the rise in tissue bicarbonate also induced by the buffering of carbonic acid (a volatile acid). The rise in metabolic rate observed in the critically ill is a fraction of that seen in an exercising athlete. Furthermore the oxygen extraction ratio is unchanged and more often decreased in septic patients who exhibit the highest metabolic rate in the critically ill. In any event, the increased metabolic demand for oxygen in the critically ill, especially in those who are septic, is primarily met by an increase in oxygen delivery, oxygen delivery being "demand-dependent" in these circumstances. The $p\text{CO}_2$ attained by the buffering of the volatile acid load generated in normoxic ECF should not, therefore, be significantly influenced by changes in metabolic rate of the order encountered in the critically ill.

Aerobic glycolysis and associated generation of CO_2 by oxidative phosphorylation decreases in dysoxic states as the availability of oxygen relative to demand decreases. Thus the fall in tissue pH in severely dysoxic states is
5 due almost exclusively to the protons released by adenine nucleotide hydrolysis and their interaction with the body buffers.

If it is assumed that the intramucosal pCO_2 and pH are solely determined by the amount of volatile and fixed
10 metabolic acid being buffered in the ECF at the time, the intramucosal pH can be expected to remain constant as oxygen delivery is reduced with or without a reduction in blood flow until the point at which supply-dependency or dysoxia develops. Below this point the pCO_2 in ECF should
15 rise and the intramucosal pH fall as the contributions by aerobic metabolism to volatile acid decreases and by anaerobic metabolism to proton release increases with further reductions in oxygen delivery.

20 Actual Intramucosal pH

The buffering of the protons by tissue bicarbonate in dysoxic states causes the pCO_2 to rise. As the bicarbonate concentrations in a "closed system", such as the ECF, is not significantly reduced by the addition of
25 a fixed acid load, the fall in pH must be inversely related to the rise in log pCO_2 at any given concentration of tissue bicarbonate. The constant bicarbonate line at 25 mEq/l on a pH-log pCO_2 diagram will show that the pCO_2 in normoxic ECF at a point A to be 40 mmHg and the pH to be 7.40. The
30 bicarbonate line moves to the right as the equilibrium pCO_2 rises above 40 mmHg to a point B in dysoxic states and the tissue pH falls below 7.40. The pH in the dysoxic state may be determined by extrapolation from the pCO_2 intercept on the constant bicarbonate line at 25 mEq/l.

35 The fall in pH induced by dysoxia alone in a tissue with a known bicarbonate concentration may be computed from the difference between the pH in the normoxic

and dysoxic states determined from the same constant bicarbonate line (pH-gap), log of the ratio pCO_{2t}/pCO_{2a} (B - A) or their antilog equivalents (pCO_2 -gap and H^+ -gap). These determinations of the magnitude in fall in pH induced by dysoxia are all dependent upon the assumption that the bicarbonate concentration in the dysoxic ECF is the same as that present in normoxic ECF. If it is assumed that the pCO_2 in normoxic ECF is the same as that in arterial blood (pCO_{2a}) and the tissue pCO_2 in dysoxic ECF is the same as the intramucosal pCO_2 measured from the lumen of the gut with a walled sampling chamber tonometer (pCO_{2t}) then the actual pH in dysoxic ECF may be calculated from the following formula (with pH_a = pH of arterial blood):

$$\begin{aligned} \text{Actual intramucosal pH} &= pH_a - (\log pCO_{2t} - \log pCO_{2a}) \\ &= pH_a - \log pCO_{2t}/pCO_{2a} \end{aligned}$$

and displayed in a perceptible form, such as human readable or audible form, or machine readable form.

20 Standard Intramucosal pH

The normoxic isobar shifts in one direction with a metabolic acidosis and to the other direction with a metabolic alkalosis, but the dysoxic component is still indicated by the difference between the dysoxic and normoxic isobars at the same bicarbonate concentration. This confounding influence of the presence of a metabolic acidosis or alkalosis in the determination of the degree of dysoxia present may be eliminated by standardizing the pH_i to that present if the bicarbonate concentration were 25 mEq/l:

$$\text{Standard intramucosal pH} = 7.40 - \log pCO_{2t}/pCO_{2a}$$

The standard intramucosal pH is easily interpreted by clinical staff who all know that 7.40 is a normal pH. It is the same as the actual pH in patients whose arterial pH is normal but differs from the actual pH with systemic disturbances in acid-base balance. As indicated above, the "pH-gap", log of the ratio pCO_{2t}/pCO_{2a} ,

and their antilog equivalents may also be used as indices of the degree of dysoxia present independently of changes in tissue bicarbonate provided that the bicarbonate concentration in the tissue is the same as that in arterial blood and remains constant. The pCO_{2t} alone does not provide a reliable measure of either the actual tissue pH or the degree of dysoxia present for interpretation of the measurement is confounded by alterations in arterial bicarbonate.

This analysis suggests that the actual measurement of intramucosal pH may be capable of detecting the presence of dysoxia whenever the delivery of oxygen fails to meet more than about 20% of the tissue's needs especially if the arterial pH is normal. The analysis further suggests that the use of standard intramucosal pH in addition to actual intramucosal pH may improve the sensitivity of measures of the degree of dysoxia present.

Clinical Implications

The indirect measurement of intramucosal pH (pHi) provides an accurate diagnostic test for the presence of macroscopic and clinical evidence of gastric, small intestinal and large intestinal ischemia in patients. The sensitivity of the intramucosal pH as a diagnostic test for gastric ischemia in man is reported to be 95% and the specificity 100%. For severe ischemic colitis after abdominal aortic surgery the sensitivity is reported to be 100% and the specificity 87%. Of particular relevance to patients who are critically ill is the inability of those with an intramucosal acidosis to secrete acid in response to pentagastrin. Those patients who have a normal gastric intramucosal pH secrete acid in response to this stimulus. It has been suggested that the inability to secrete acid in patients with an intramucosal acidosis may be due to an energy deficit secondary to a dysoxic state. An energy deficit is a known cause of stress ulceration in animals

and an impairment of gastric mucosal oxygenation the likely cause of stress ulceration in patients.

The gastric intramucosal pH, measured following the administration of an H_2 receptor antagonist to avoid
5 confounding influence of the back diffusion of acid and/or CO_2 , is inversely related to the hepatic venous lactate concentrations in patients having cardiac surgery ($r=-0.71$) and correlates closely with this and other indices of splanchnic tissue oxygenation ($r=0.92$). The gastric
10 intramucosal pH provides, therefore, an index of the adequacy of splanchnic tissue oxygenation.

The gastric intramucosal pH correlates very well and inversely with systemic blood lactate when it is abnormally elevated. In many circumstances, however, blood
15 lactate is normal when the intramucosal pH is low and no correlation between the variables can be demonstrated. Indeed a fall in gastric intramucosal pH may precede a rise in blood lactate in a deteriorating patient by many hours or even days. Changes in actual pH influence the pH
20 dependent enzymes regulating carrier mediated afflux of lactate from muscle and the pH dependent enzyme phosphofructokinase which regulates the rate of anaerobic glycolysis. In addition blood lactate is the net effect of both production by anaerobic glycolysis and consumption by
25 tissues such as the myocardium. The overall correlation between the two variables is thus rather poor ($r=-0.40$) but nevertheless statistically significant ($p=0.026$). Thus in addition to providing indices of gastric mucosal and splanchnic tissue oxygenation the indirect measurement of
30 gastric intramucosal pH provides an index of the adequacy of global tissue oxygenation.

The indirect measurement of intramucosal provides a measure of the adequacy of tissue oxygenation in the most superficial layer of the mucosa, a region of the gut
35 rendered relatively hypoxic by the counter current exchange system within the mucosal vasculature and hence especially

sensitive to alterations in the adequacy of tissue oxygenation. It also provides a measure of the adequacy of tissue oxygenation in a region of the body that is among the first to develop an inadequacy of tissue oxygenation or
5 dysoxia in shock and the last to be restored to normality with resuscitation. Splanchnic vasculature is selectively constricted by the endogenous vasoconstrictors released in shock. For these reasons a fall in intramucosal pH may occurs hours to days in advance of any other conventional
10 evidence of an inadequacy of tissue oxygenation, most specifically arterial acidosis, elevation in blood lactate, hypotension and oliguria.

It is concluded that the indirect measurement of gastric intramucosal pH provides a sensitive measure of the
15 adequacy of splanchnic and even global tissue oxygenation in patients in addition to providing an index of the adequacy of superficial gastric mucosal oxygenation.

Correlations With Acid-base Balance and Clinical Events

20 The indirect measurement of gastric intramucosal pH may correlate very closely with the arterial pH ($r=0.67$) and other systemic indices of a disturbance in acid-base balance such as arterial bicarbonate ($r=0.50$), the base deficit in extracellular fluid ($r=0.60$) and base
25 deficit in blood ($r=0.63$). This is consistent with the deduction that gastric intramucosal pH provides an index of the balance between the protons released by ATP hydrolysis and consumed in the resynthesis of ATP by oxidative phosphorylation. As with global measurements of blood
30 lactate changes in systemic acid-base balance provide a very dampened signal of disturbances in the adequacy of tissue oxygenation. A fall in intramucosal pH will often precede a fall in arterial pH by hours or even days.

The predictive value of measurements of gastric
35 intramucosal pH for outcome are superior to those of the systemic measures of acid-base balance. Maynard et al, for

example, compared the predictive value of measurement of gastric intramucosal pH with those of arterial pH and base excess for death in ICU patients. The likelihood ratio for pHi was 2.32, for arterial pH 1.52 and base excess 1.47.

5 Logistic regression showed only pHi to independently predict outcome. In Boyd et al's study, the gastric intramucosal pH was likewise of better predictive value for outcome than base excess. Clinical experience has shown that changes in gastric intramucosal pH correlate far

10 better with the passage of clinical events than either the arterial pH or base excess. Indeed abnormalities in these systemic measures of acid-base imbalance will often occur only as the intramucosal acidosis is being reversed and the patient's condition is improving.

15 Reperfusion after the low-flow and particularly no-flow states induced in Antonsson et al's validation study in pigs caused the intramucosal pH to rise and the arterial bicarbonate to fall. Similarly in patients the reversal of a severe intramucosal acidosis may be

20 accompanied by a fall in arterial pH and base excess at abnormally low levels. These observations are consistent with the consequences described above of reestablishing perfusion in a dysoxic tissue bed in patients. The pCO_2 in the venous effluent leaving the dysoxic tissue bed is

25 elevated but the bicarbonate concentration is not significantly reduced by the buffering of the fixed acid in the tissue bed. The bicarbonate is only reduced by the loss of CO_2 during the passage of the venous effluent through the pulmonary circulation (an open system). As dissociation

30 between the direction of change in the intramucosal and systemic pH is to be expected after flow is reestablished through a dysoxic tissue bed.

Intramucosal pH as a Therapeutic Target

35 "Gut-directed" and "pHi-directed" therapies may improve outcome. These therapies use a normal pHi or pHi

greater than 7.35 as an additional therapeutic goal in the resuscitation of patients. This pH was chosen to ensure the pH was maintained well within the normal limits reported for normal subjects. The normal limits may, however, differ from institution to institution with the use of saline and different blood gas analyzers, a problem solved by the air sampling medium/IR pCO₂ analysis embodiments of the instant invention. It is furthermore possible that an end-point other than 7.35 might be more appropriate. Values such as 7.25; 7.30; 7.35; 7.37 etc. may also be useful.

While it is clearly desirable to maintain a normoxic state by maintaining the standard pHi at 7.40, it is not necessarily desirable to maintain the actual pHi at 7.40. There is a considerable body of evidence indicating that mild degrees of cellular acidosis protect cells in anoxia and ischemia possibly by limiting the activity of the autolytic enzymes responsible for cell injury and death. A cellular acidosis may in addition facilitate carrier-mediated afflux of lactate from cells and bring the intracellular pH to an optimal range for anaerobic glycolysis during anaerobic metabolism. Furthermore the addition of bicarbonate to the extracellular environment attenuates the fall in intracellular pH during ATP depletion and accelerates cell death. The presence of an actual intramucosal acidosis may, therefore, be desirable and efforts to correct a metabolic acidosis with bicarbonate potentially harmful. Indeed the practice of correcting a metabolic acidosis induced by a cardiac arrest by the administration of bicarbonate is no longer recommended.

It is concluded that acid-base balance is intimately related to the adequacy of tissue oxygenation in so far as it relates to the balance between the protons released by ATP hydrolysis and consumed by ATP synthesis from oxidative phosphorylation. The tissue pH is

determined by the $p\text{CO}_2$ attained following and buffering of the metabolic acid released into the ECF and the bicarbonate concentration in ECF at the time--- the "buffer hypothesis". The intramucosal pH is related to blood flow only in so far as it relates to the adequacy of tissue oxygenation. The assumption that tissue bicarbonate is the same as that in arterial bicarbonate is only valid in the absence of the generation of an alkaline tide and associated secretion of acid. The indirect measurement of actual gastric intramucosal pH is the sum of the effects of several determinants of an intramucosal acidosis. It is relevant to activity of pH-dependent enzymes especially as they might relate to cellular injury in dysoxic states. By eliminating the confounding effects of disturbances in systemic acid-base balance the standard gastric intramucosal pH provides a measure of the acidosis attributable to an imbalance between ATP hydrolysis and resynthesis, or degree of dysoxia present. Systemic measures of acid-base balance may be dissociated from the adequacy of tissue oxygenation upon reperfusion of a dysoxic tissue bed and correlate poorly with clinical events relative to the measurement of gastric intramucosal pH.

In light of all the above, it will be appreciated that one series of embodiments of the present methods relate to the use of arterial carbon dioxide concentrations (measured directly or indirectly, preferably as an end tidal carbon dioxide value) as a predictive indicator of the pH of the most superficial layer of the mucosa of the wall of an internal solid organ, particularly the gut. In recognizing that

$$\text{pHi} = \text{pH}_a + \log \frac{\text{pCO}_{2a}}{\text{pCO}_{2t}}$$

and that pCO_{2a} is approximately equal to $\text{pCO}_{2\text{-end tidal}}$, thus

$$pH_i = pH_a + \log \frac{pCO_{2\text{-end tidal}}}{pCO_{2t}}$$

- 5 Either or both of these may be employed.

In accordance with the practice of the methods of the present invention, the pCO_2 of the wall of the organ is determined. This is preferably done by inserting a tonometric catheter with a walled sampling chamber into or
 10 adjacent the organ of interest. The sampling chamber is filled with a gaseous or liquid sampling medium such as air or saline. The sampling medium is allowed to come to equilibrium (equilibrate) with the area so that the pCO_2 concentration of the sampling medium reflects the pCO_2 of
 15 the superficial layer of the mucosa of the organ of interest. The pCO_2 concentration of the sampling medium is determined, giving pCO_{2t} .

In conjunction with the determination of the pCO_2 of the mucosa, the carbon dioxide concentration in arterial
 20 (pCO_{2a}) or venous blood is determined directly or indirectly. (A highly preferred indirect measure is end tidal pCO_2 , or $pCO_{2\text{-end tidal}}$). The two values (e.g., pCO_{2a} and pCO_{2t}) are then subjected to a nomogram, such as those described in equations above, to determine for example, a
 25 pHi value or pH gap.

In a highly preferred embodiment, the sampling medium for the walled sampling chamber is air. The air is aspirated to an IR or Raman spectrometer. In combination, the measurement of end tidal pCO_2 is employed as a
 30 substitute for the arterial $pCO_{2(a)}$. The end tidal respiratory air is likewise aspirated to an IR or Raman spectrometer. Both gas analyzing devices are controlled by a microcomputer, which also effects the selected nomogram

or nomograms which compare the $p\text{CO}_2$ of the wall of the organ (gut) with the end tidal $p\text{CO}_2$ value. The gas analyzing devices may operate on a single channel, or via multiple channels.

5 Additional detection techniques may be performed on the air aspirated from the patient, either via respiration or from the tonometric walled sampling chamber. For example, IR or Raman analyses may be performed to determine the level of anesthetic gases, such as N_2O . The
10 results of the nomogram are displayed on a monitor (not shown) in human or machine readable form.

In a highly preferred embodiment, the operation of one example of an infrared gas analyzer is controlled by a microcomputer. The microcomputer itself is not, by
15 itself, part of the present invention. For this reason and because one skilled in the relevant arts could routinely program a general purpose computer to follow the routines required for this application, the microcomputer will not be described in detail herein. (See the U.S. patents
20 incorporated herein by reference.)

Referring now to Figure 13, a gas analyzer or detector 320 is shown in accord with the principles of the present invention. Analyzer 320 is specifically designed to monitor the concentration of carbon dioxide in the
25 exhalations of a medical patient--e.g., a patient being ventilated during a surgical procedure.

The major components of the infrared gas analyzer 320 are a powered unit 322 and a sensor assembly 324 of a transducer head 326 and an airway adapter 328. The
30 transducer head 326 is connected to the unit 322 of the gas analyzer 320 by a conventional electrical cable 330.

In the application of the invention depicted in Figure 13, the gas analyzer 320 is employed to measure fluid parameters of interest, similar to the apparatuses
35 shown and discussed above, except that a gaseous sampling medium, such as air, is conveyed, either manually or automatically, as shown above, and analyzed by the infrared

sensor assembly 324, where the sampling medium is conveyed to the assembly 324 via one of the above described tonometric catheter devices. This information can be effectively employed by medical personnel to monitor the condition of a patient's internal organ more accurately and more quickly than before.

Figure 13 depicts an in-stream type of infrared gas analyzer, shown merely for purposes of illustration, but one skilled in the art will appreciate that the same principles apply to the use of a side-stream type IR gas analyzer.

Figure 14 schematically illustrates a biological filter (bio-filter) apparatus 340 being employed in-line, between an exemplary tonometric catheter apparatus 342 and the above-discussed exemplary infrared sensor assembly 324. The bio-filter 340 can be any of a number of biological filters known to those skilled in the art and is especially useful to allow side-stream systems to allow sample return or in-stream infrared gas analyzer apparatuses to be used in multi-patient applications.

Due to the sensitivity of the current commercial infra red sensors or detectors to moisture content, and due to the high moisture content of air sampling-medium based pCO_2 samples coming from both an *in vivo* tonometric walled sampling chamber and end tidal samples, a moisture filter or other dehumidifying means is preferably employed.

For example, air-based pCO_2 sample can be passed through dehumidification tubing, such as Nafion tubing.

Other methods of eliminating moisture problems include employing a heat sink around part or all of the IR optical path, particularly the lens window where the IR source passes light. Yet another means includes employing a barrier or filter which is selectively permeable to water vapor (moisture) and/or the gases of interest, particularly pCO_2 .

It should therefore also be noted that the filter 340 can also optionally include a dehumidifying means,

e.g., a water vapor filter or removal medium, either alone or in addition to the biological filter, for allowing any water vapor in the sampling medium or the sampling chamber to disperse in the environment by delivering the mixture thereof past a water-vapor-permeable wall or medium.

Figure 14 also schematically illustrates the addition of a gaseous sampling medium pressure sensor and/or regulator 350 (optional) for measuring the pressure of a gas sampling medium, such as air, for example, and/or for regulating such pressure to be substantially at some predetermined pressure level, such as atmospheric pressure, for example, at which the IR gas analyzer is designed to operate and give accurate, reliable results.

It will be appreciated that Raman spectrometers (gas analyzers) offer advantages over IR in the present invention, including the fact that such dehumidifying means need not be employed. Additionally, the use of this combination (a tonometric catheter plus a Raman spectrometer) would allow the measurement of oxygen gas; nitrogen gas; water; N_2O and other anesthetic agents such as halothane, enflurane, isoflurane, serroflurane, and desflyrane, all of which exhibit Raman scattering. Raman devices not only measure pCO_2 more accurately, but can measure N_2 , O_2 and H_2O directly. This may reduce the potential error associated with certain IR techniques, especially where other material (N_2O ; O_2 ; H_2O) may effect the pCO_2 IR measurement. It also would allow detection of air leaks.

Another important advantage would be the ability to employ a fiberoptic probe within the sampling chamber 40 of the tonometric catheter.

Significant deviations from this "design" gaseous sample pressure level can result in inaccurate readings due to density changes resulting from the gas being over-pressurized or under-pressurized.

In a manual system, wherein a syringe is used to draw the gaseous sample into the sensor assembly, it has

been found to be useful to provide one or more holes 360 in the syringe body 362 shown in Figure 15 in order to allow for pressure regulation and equalization with the atmosphere. Alternatively, a pressure-difference caused by a pump, with or without a pressure regulation or connection device, as needed, can be employed. In addition, as is clear from the foregoing discussion, the system can use a single-tube catheter device or a dual-tube version, wherein one tube delivers the sampling medium to the sampling chamber and the other is used to extract it for measurement.

Clinical Example

In the following clinical study, approval was received by the appropriate hospital ethics committee and informed consent was obtained from the patients. Recordings were performed in ten patients undergoing laparoscopic cholecystectomy at Liverpool Hospital, Sidney Australia. All patients underwent a general anesthetic with muscle relaxation and artificial ventilation. The patients were placed supine upon the operating table. All patients had nasogastric insertions of a nasogastric tube with a pressure transmitting chamber attached. The balloon on the nasogastric tube was one usually employed for tonometry, namely indirectly measuring intramural pH, i.e. a TRIP TGS catheter from Tonometrics, Inc., Worcester, Massachusetts U.S.A. The intragastric position of the tonometer was confirmed by aspiration of gastric fluids, auscultation of air insufflation of the stomach, and confirmation of a rise of IAP following external epigastric pressure. None of the patients had intra-abdominal adhesions. The stomach was visualized at laparoscopy to confirm its intra-abdominal position.

The pressure volume curve of the gastric balloon at 37° centigrade confirms that inflation of up to 3.0 ml of air allowed the balloon to reflect changes in IAP. Each balloon was individually checked prior to insertion. The

balloon on the nasogastric tube was completely emptied prior to injection of 2.5 ml of air. Particular attention was paid to make sure that the stomach was in a period of quiescent motor activity with no evidence of the phase II or phase III of the migrating motor complex Vantrappen G. The migrating myoelectric complex. In Motility of the digestive tract. Weinbeck M, New York, Raven Press 1982;157-167.

A size 16 Foley catheter was inserted into the bladder. A T-piece bladder pressure attachment was attached to the in-dwelling catheter and a primed pressure transducer was connected to the system. The pressure transducer connected to the urinary catheter was placed at the same level in the mid-axillary line as a transducer which was primed and connected to the nasogastric tube. 50 ml of normal saline was inserted into the bladder via a three way stopcock. The urinary catheter was clamped.

Simultaneous IAP recordings were obtained by means of an intragastric balloon catheter and urinary catheter transduced to a Datex AS3 three channel recorder. After zeroing the transducer, the pressures from both the stomach and bladder were recorded simultaneously with a two chart recorder. The volume of intraperitoneal gas was recorded. Insufflation pressure was limited to 20 millimeters Hg. The CO₂ was insufflated at a rate of two liters per minute. The nasogastric and urinary catheters were removed at the end of the procedure.

Statistical analysis was carried out using SAS version 6.04. Pearson product moment correlations coefficients and Spearman rank coefficients were used to assess the relationships between the intravesical and gastric pressures in each patient. Adjustments for age and body mass index were not attempted. The mean 95% confidence interval of the 9 slopes were calculated.

There were 8 females and 2 males aged 43 plus or minus 15 years. The mean body mass index was 30 plus or minus 5. Insufflation of the abdominal cavity was achieved

with carbon dioxide to a preset pressure of 20 mmHg as regulated by the insufflator (Wisap). Mean volume of gas insufflated was 8.8 plus or minus 4.3 liters. Baseline pressures were 0-2 mmHg in all patients rising to a mean
5 maximum pressure of 14 plus or minus 4 mmHg range 8-20 mmHg.

Simultaneous IAP recordings were obtained by means of the intra-gastric balloon and urinary catheter transduced to a Datex AS3 three channel recorder. The
10 pressure volume response was variable, due to the different peritoneal capacity of individual patients. Transduced pressures had a rapid response time facilitating recording of changes during insufflation. The mean insufflation time to these pressures was 4.7 plus or minus 1.9 minutes.
15 There was a very strong correlation for all pressures up to 17 mmHg. Individual correlations coefficients range from 0.90 to 0.99 (p less than 0.01). Figure 20 shows the relationship between intravesical and gastric pressures in one of the patients. The results of one patient were
20 excluded due to protocol violation as the balloon required the installation of 2.5 ml to allow it to record pressures, however, the correlation for this patient was 0.95.

IAP measured by the gastric technique can be predicted by the following equation:

25
$$\text{IAP} = 0.17 + 1.05 \times \text{intravesical pressure.}$$

The 95% confidence intervals for the intercept were -1.22 to 1.56, and for the slope 0.86 to 1.24.

Accordingly, while several preferred embodiments of the invention have been disclosed, it will be
30 appreciated that principles of the invention, as set forth in the following claims, are applicable to other embodiments.

IN THE CLAIMS

What is Claimed is:

- 5 1. A tonometric catheter apparatus for measuring the internal pressure of a human or other mammal comprising:
- (a) an elongated tonometric catheter tube having at least one lumen extending longitudinally therethrough;
- (b) at least one walled pressure transmitting
10 chamber on said catheter tube in pressure transmitting communication with the interior of said lumen, said walled pressure transmitting chamber generally surrounding a portion of said catheter tube and being sealingly interconnected therewith, the wall of said pressure
15 transmitting chamber being composed of a wall material being substantially impermeable to liquid fluids or gaseous fluids, said pressure transmitting chamber forming an interior space with said catheter tube in order to allow the transmission of change in pressure through said
20 pressure transmitting chamber to the interior of said lumen via a pressure transmitting medium;
- (c) pressure sensor means for measuring the pressure of said pressure transmitting medium.
- 25 2. An apparatus according to Claim 1 wherein said walled pressure transmitting chamber is defined by a balloon member generally surrounding a portion of said catheter tube and sealingly interconnected therewith.
- 30 3. The apparatus according to Claim 1 wherein said pressure transmitting chamber is in fluid continuous communication with said lumen.
4. An apparatus according to Claim 1 wherein said
35 pressure transmitting medium is air.

5. An apparatus according to Claim 1 wherein said pressure transmitting medium further comprises a saline solution.

5 6. An apparatus according to Claim 1 wherein said pressure sensor means further comprises a manometer.

7. An apparatus according to Claim 1 wherein said pressure sensor means further comprises a pressure
10 transducer.

8. A tonometric catheter apparatus for measuring a liquid fluid or gaseous fluid property indicative of the internal pressure of an internal cavity of a human or other
15 mammal *in vivo* comprising;

(a) an elongated tonometric catheter tube having at least one lumen extending longitudinally therethrough;

(b) at least one walled pressure transmitting chamber on said catheter tube in pressure transmitting
20 communication with the interior of said lumen, said walled pressure transmitting chamber being defined by a balloon member generally surrounding a portion of said catheter tube and being sealingly interconnected therewith, said wall of said balloon member being composed of a wall
25 material substantially impermeable to *in vivo* gaseous fluids or liquid fluids, said balloon member capable of being inflated for forming a pressure sensing diaphragm for acting upon pressure transmitting medium within said walled pressure sensing chamber;

30 (c) pressure sensor means for sensing the pressure of said pressure transmitting medium;

9. An apparatus according to Claim 8 wherein said pressure sensor means includes a interface means for
35 converting an output of said pressure sensor means into a human readable format.

10. An apparatus according to Claim 8 wherein said pressure sensing means further comprises a pressure transducer.
- 5 11. An apparatus according to Claim 8 wherein said pressure sensing means further comprises a manometer.
12. An apparatus according to Claim 10 wherein said pressure transducer is mounted within said pressure transmitting chamber.
- 10 13. An apparatus according to Claim 9 wherein said pressure transmitting medium is air.
- 15 14. An apparatus according to Claim 9 wherein said pressure transmitting medium further comprises a saline solution.
- 20 15. An apparatus according to Claim 8 wherein said internal cavity is the human gut.
16. An apparatus according to Claim 8 wherein said internal cavity is the cranium.
- 25 17. A tonometric catheter apparatus for measuring at least one liquid fluid or gaseous fluid property indicative of the condition of an internal organ of a human or other mammal *in vivo*, comprising:
- 30 (a) an elongated tonometric catheter tube comprising a first lumen and a second lumen extending longitudinally therethrough;
- (b) at least one walled sampling chamber on said catheter tube in fluid communication with the interior of said first lumen, said walled sampling chamber generally surrounding a portion of said catheter tube and being
- 35

sealingly interconnected therewith, at least a portion of the wall of said sampling chamber being composed of a wall material that is freely and selectively permeable to one or more liquid fluids or gaseous fluids of interest, said wall material being substantially impermeable to other interfering liquid fluids or gaseous fluids, and said sampling chamber forming an interior space with said catheter tube in order to allow said one or more liquid fluids or gaseous fluids of interest from the tissue of the wall portion of the internal organ to permeate into said sampling chamber with a sampling medium, with said catheter tube extending to a position outside of the body of the human or other mammal;

(c) first sensor means for sensing the level of at least one of said liquid fluids or gaseous fluids of interest permeated from the tissue of the wall portion of the internal organ with said fluid sampling medium in sampling chamber;

(d) at least one walled pressure transmitting chamber on said catheter tube in pressure transmitting communication with the interior of said second lumen, said pressure transmitting chamber generally surrounding a portion of said catheter tube and being sealingly interconnected therewith, the wall of said pressure transmitting chamber being composed of a wall material that is substantially impermeable to liquid fluids or gaseous fluids normally encountered *in vivo*;

(e) a pressure transmitting medium within said pressure transmitting chamber for transmitting pressure received by said walled pressure transmitting chamber to said second lumen;

(f) pressure sensor means for sensing the pressure of said pressure transmitting medium.

18. The apparatus according to Claim 17, wherein said sensor means further comprises a manometer located external to said human or other mammal.

5 19. The apparatus according to Claim 17, wherein said pressure sensor means further comprises a pressure transducer located external to said human or other mammal.

10 20. The apparatus according to Claim 19 wherein said pressure transmitting chamber and said sampling chamber share a common wall.

15 21. The apparatus of Claim 20 wherein said common wall creates a single chamber for both pressure transmitting and fluid sampling.

22. A tonometric catheter apparatus for measuring the internal pressure of a human or other mammal comprising:

20 (a) an elongated tonometric catheter tube having at least one lumen extending longitudinally therethrough;

25 (b) at least one walled pressure transmitting chamber in pressure transmitting communication with the interior of said lumen, so as to allow the transmission of change in said internal pressure through said pressure transmitting chamber to the interior of said lumen via a pressure transmitting medium;

30 (c) pressure sensor means for measuring the pressure of said pressure transmitting medium.

35 23. An apparatus according to Claim 22 wherein said wall of said pressure transmitting chamber is defined by a balloon member in sealing contact with a portion of said catheter.

24. A tonometric catheter apparatus for measuring at least one liquid fluid or gaseous fluid property indicative of either the internal pressure or the pH of an internal organ of a human or mammal *in vivo* comprising:

5 (a) an elongated tonometric catheter tube comprising lumen means extending longitudinally therethrough;

(b) a walled pressure transmitting sampling chamber on said catheter tube in fluid communication with
10 the interior of said lumen means, said walled sampling pressure transmitting chamber generally surrounding a portion of said catheter tube and being sealingly interconnected therewith, said sampling chamber forming an interior space with said catheter tube in order to allow
15 one or more liquid fluids or gaseous fluids of interest from the tissue of the walled portion of the internal organ to permeate into said pressure transmitting sampling chamber;

(c) a first sensor means for sensing the level
20 of at least one of said liquid fluids or gaseous fluids of interest permeated from a tissue of the wall portion of the internal organ with a sampling medium;

(d) second sensor means for sensing the pressure
of said sampling medium in said pressure transmitting
25 sampling chamber.

25. The apparatus according to Claim 24 wherein said second sensor means further comprises a pressure transducer for sensing the pressure of said sampling medium within
30 said lumen means.

26. The apparatus according to Claim 25 wherein said first sensor means is located *in vivo* and transmits a signal external to said human or other mammal via said
35 lumen means.

27. An apparatus according to Claim 26 wherein said sampling fluid comprises a saline solution.

28. A method of monitoring the internal pressure of
5 a human or other mammal comprising:

(a) inserting an elongated tonometric catheter tube having at least one lumen extending longitudinally therethrough into said human or other mammal;

10 said catheter having at least one walled pressure transmitting chamber in transmitting communication with the interior of said lumen, said walled chamber surrounding a portion of said catheter tube and being sealingly interconnected therewith, said pressure transmitting
15 chamber forming an interior space with said catheter tube in order to allow the transmission of change in pressure through said pressure transmitting chamber to the interior of said lumen via a pressure transmitting medium;

20 (c) receiving a pressure indicating signal through said lumen;

(d) providing pressure sensor means for receiving said pressure indicating signal and converting it
25 into a human readable form.

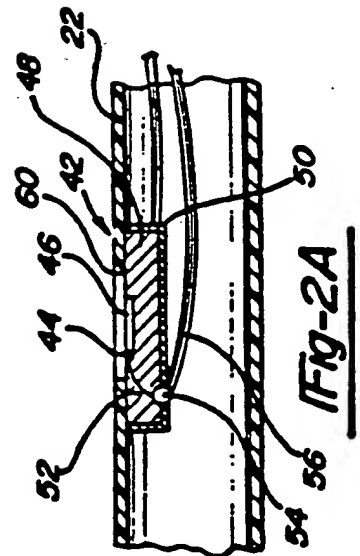
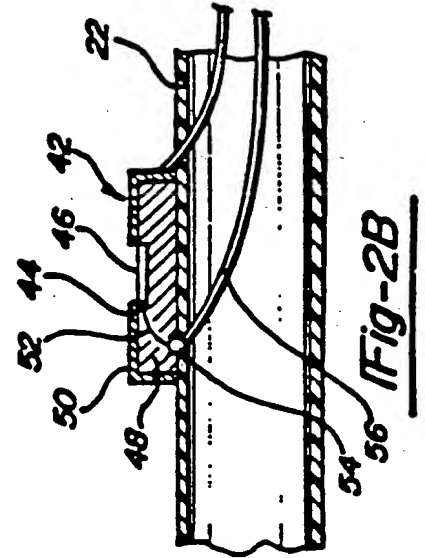
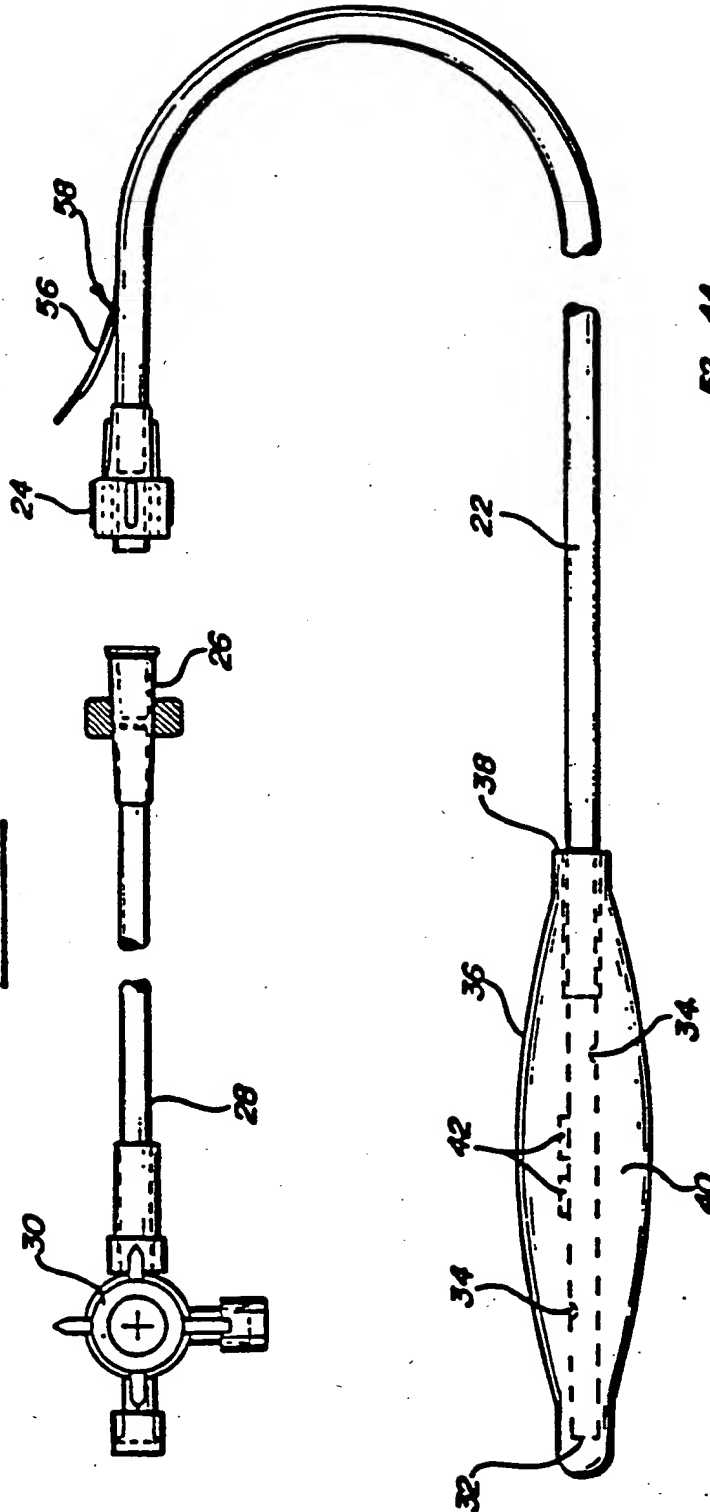
29. The method according to Claim 28 wherein said receiving step further comprises receiving pressure transmitting medium on a pressure transducer.

30

30. The method according to Claim 28 further comprising insufflating said pressure transmitting chamber with pressure transmitting medium after insertion in said human or other mammal; reading changes in said internal pressure over a period of time by repeatedly sensing the changes in the pressure transmitting medium pressure without removal or reinsertion of said catheter.

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Fig-1



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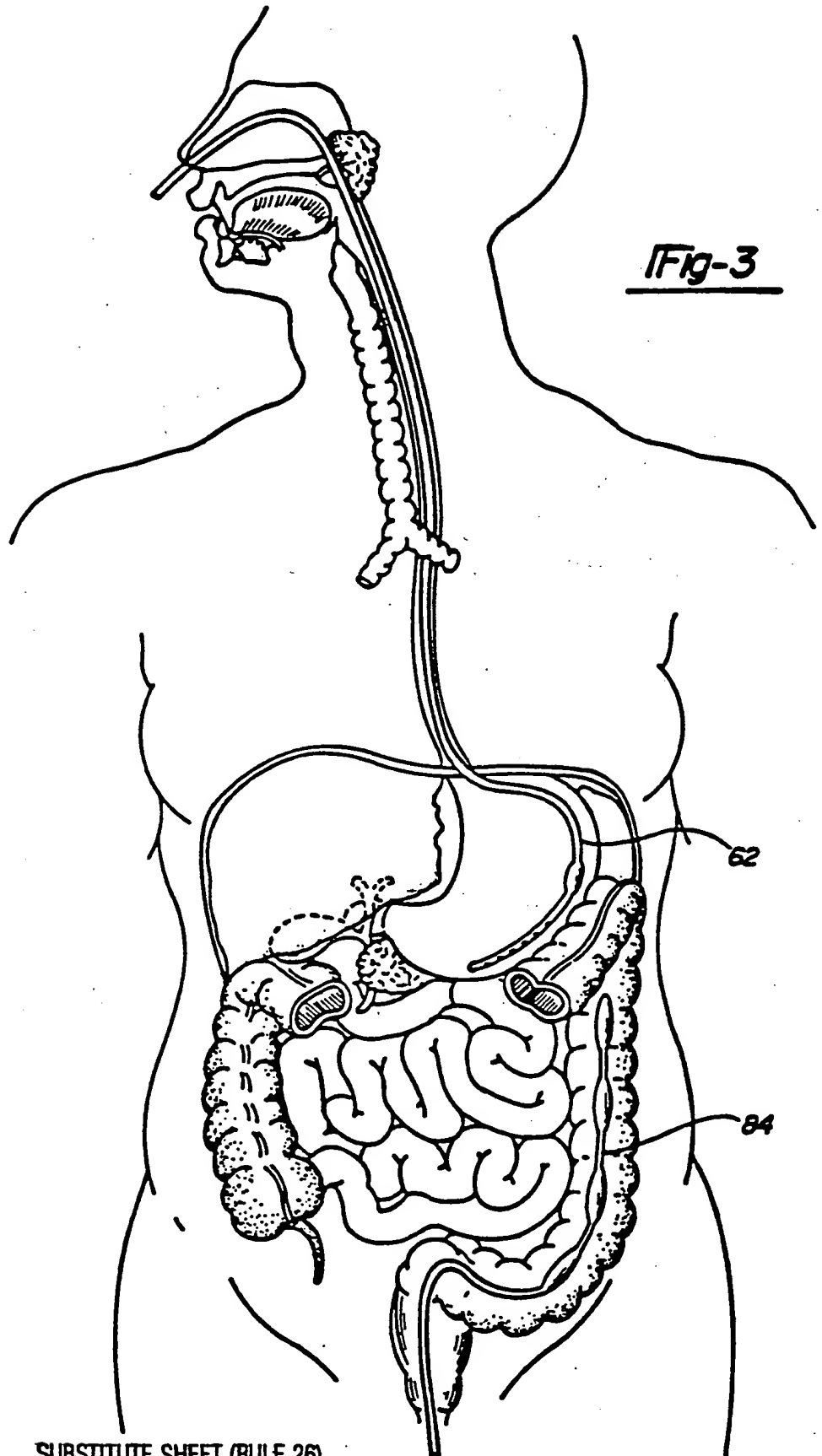
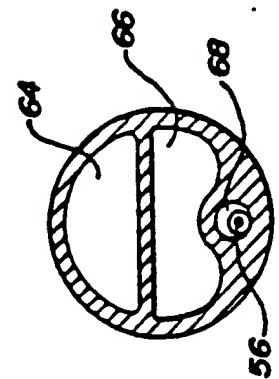
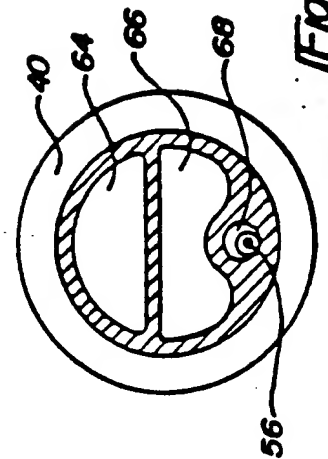
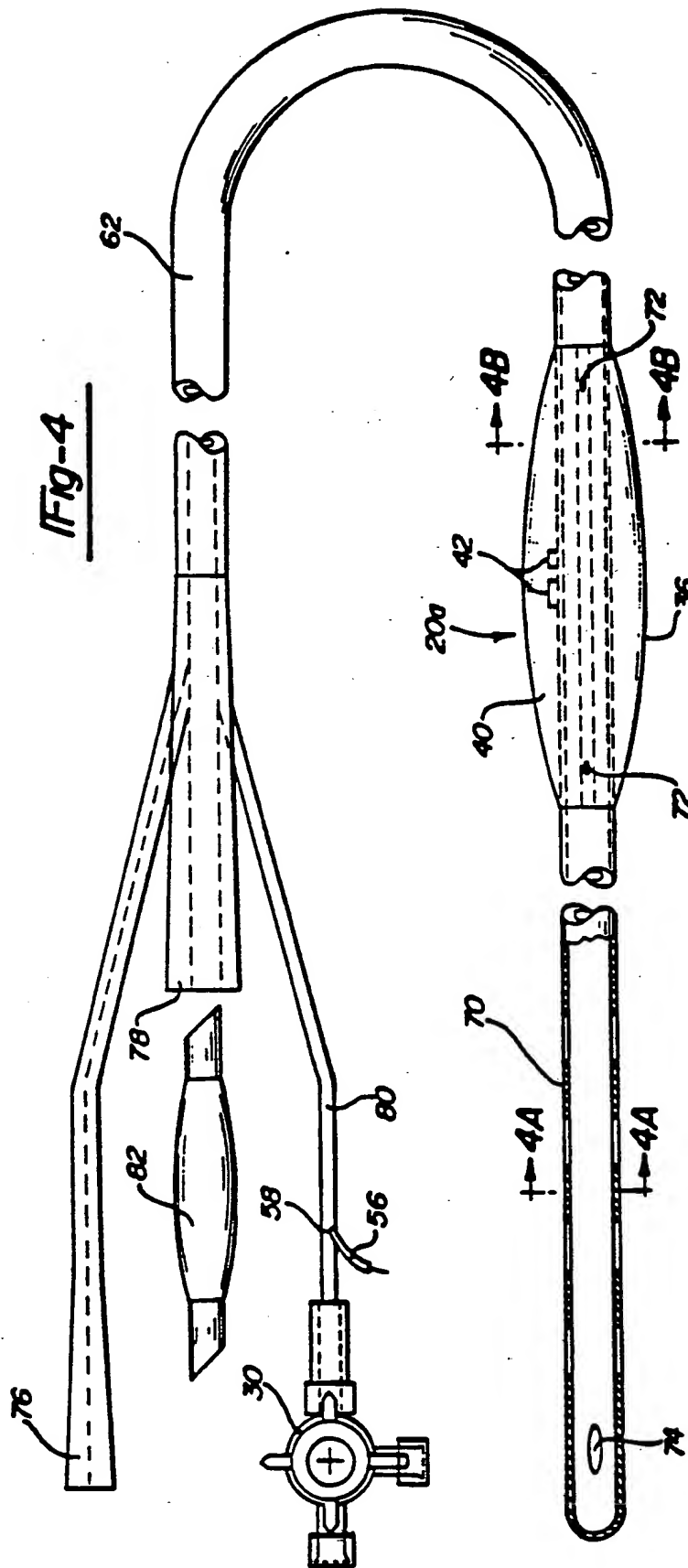


Fig-3

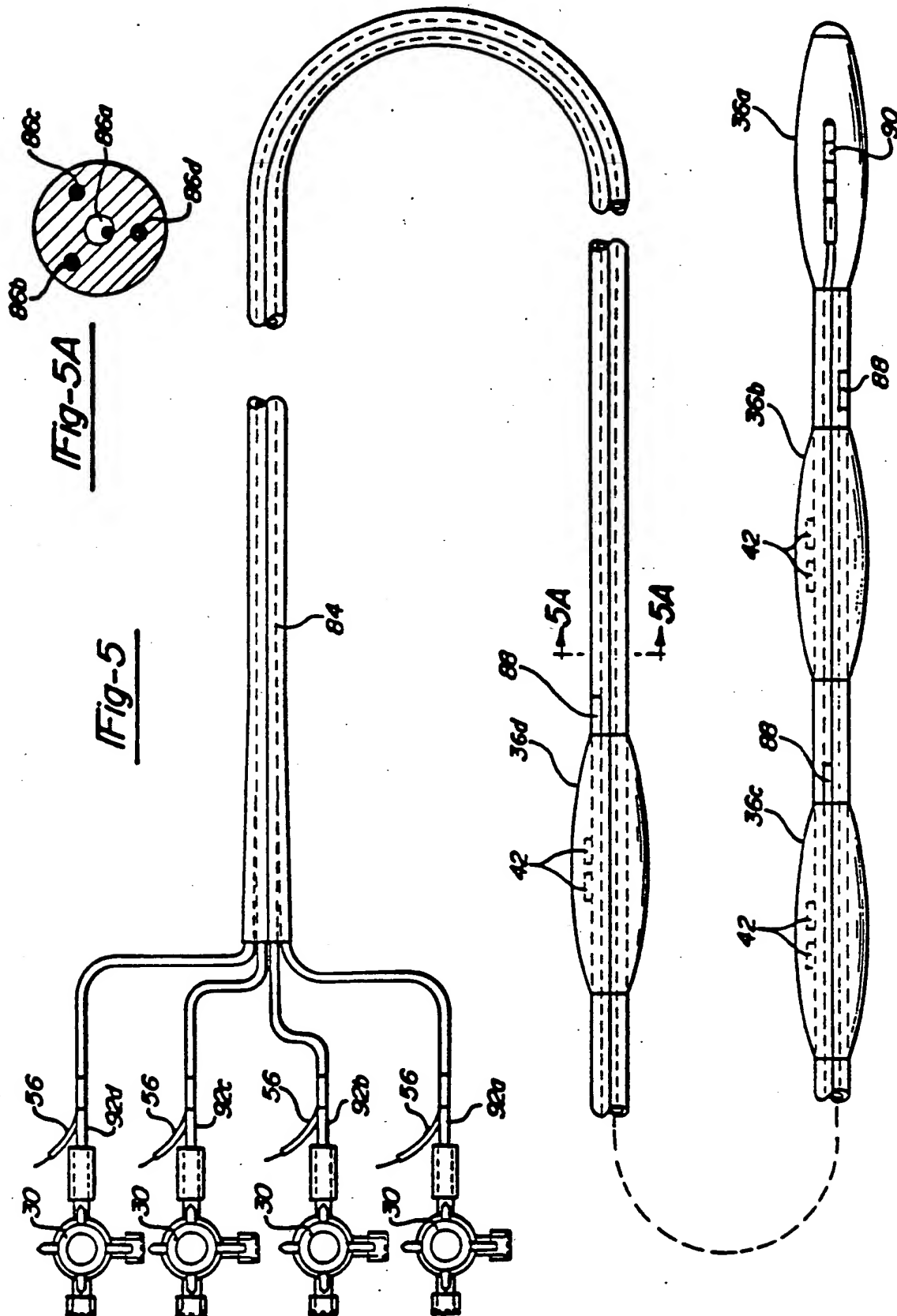
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Fig-6

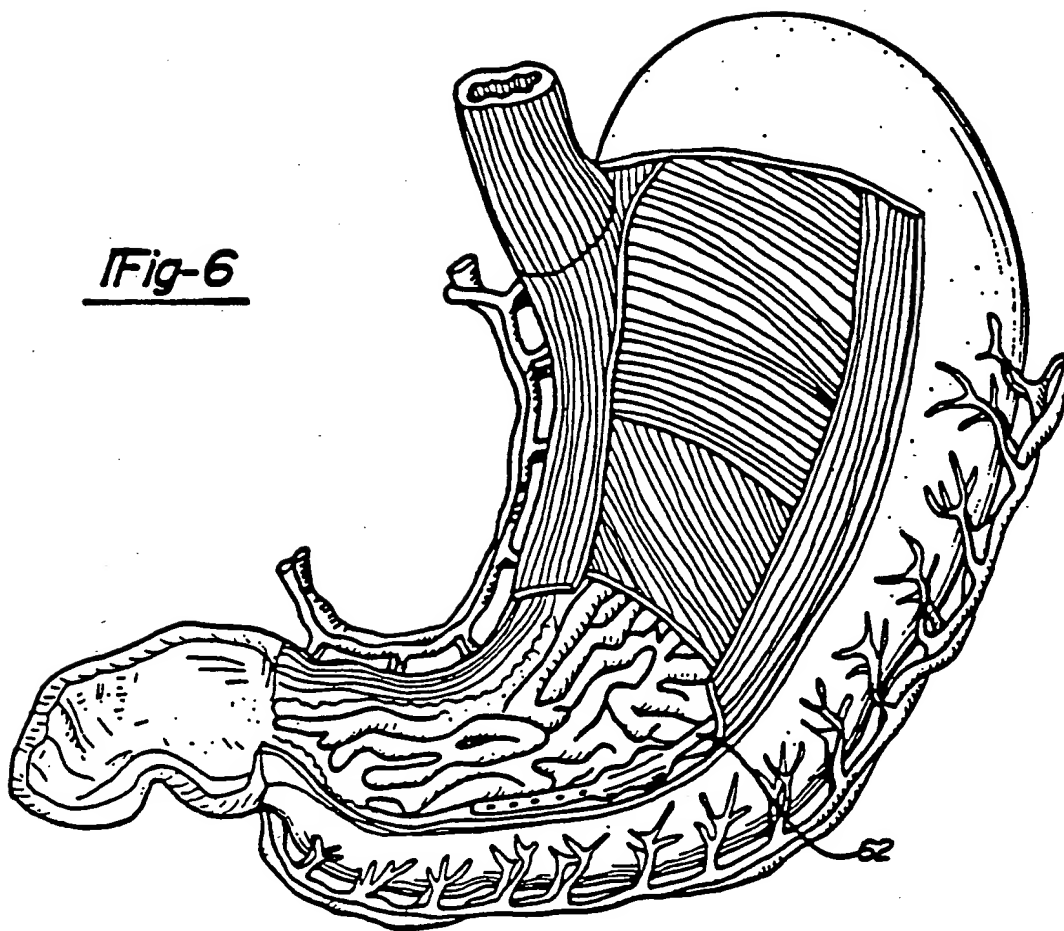
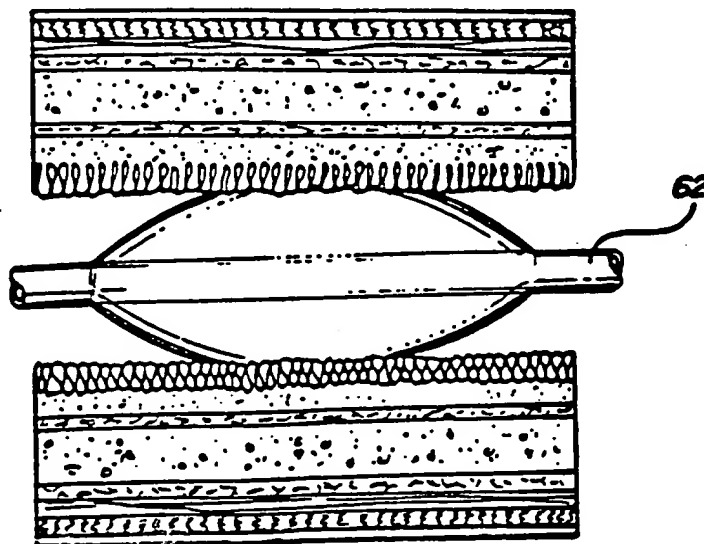
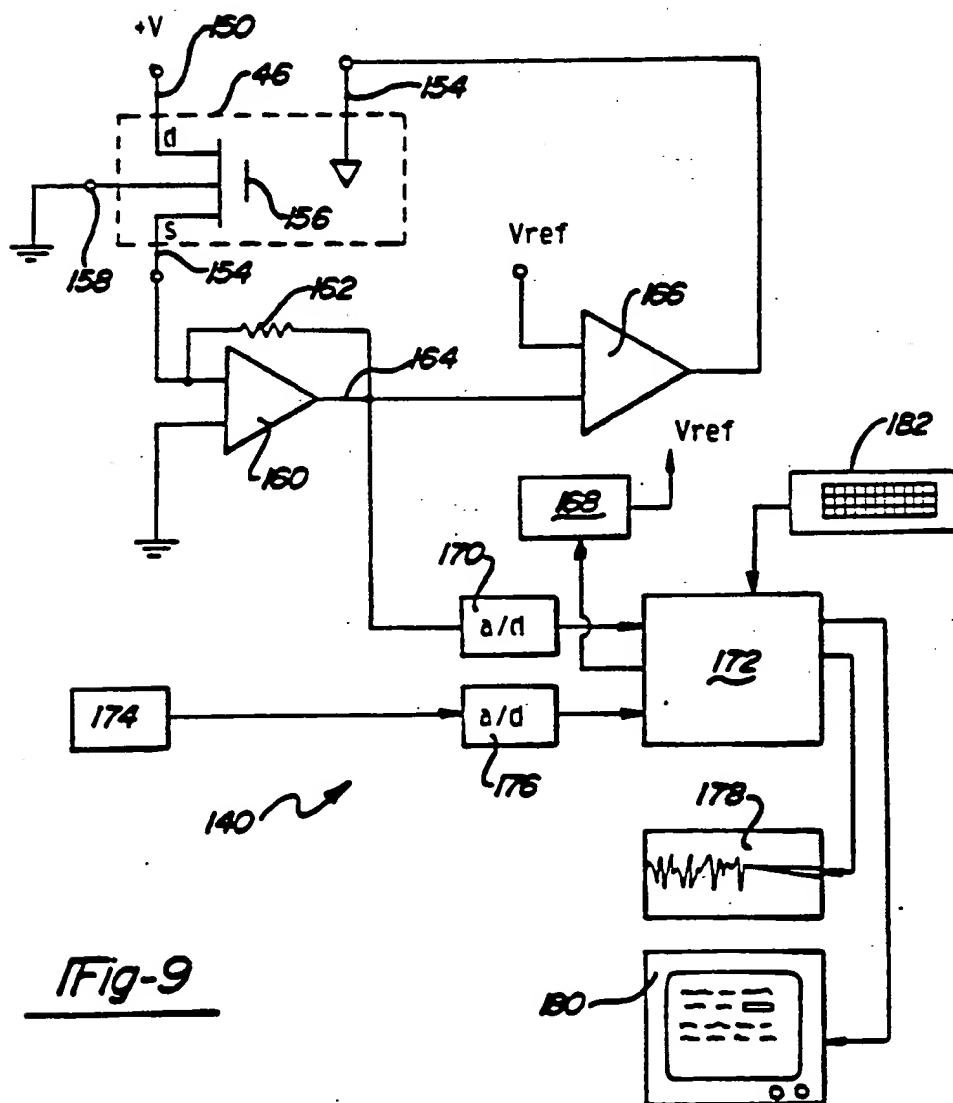
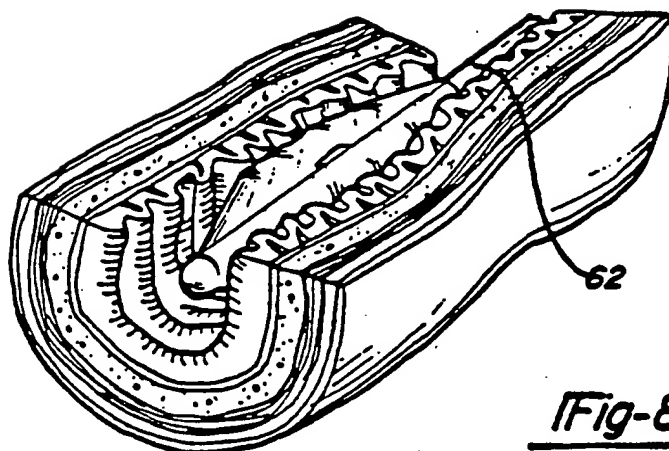


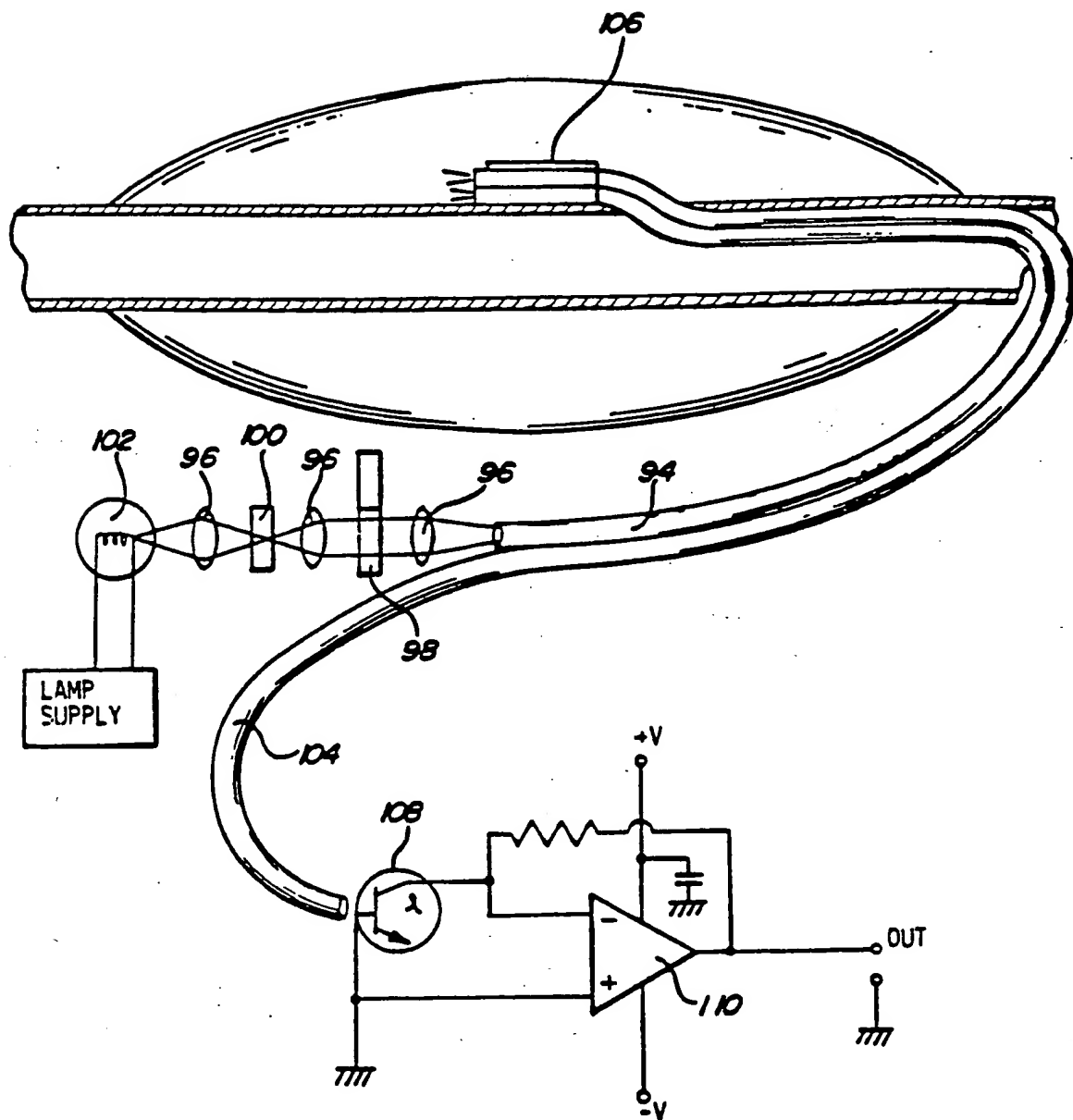
Fig-7



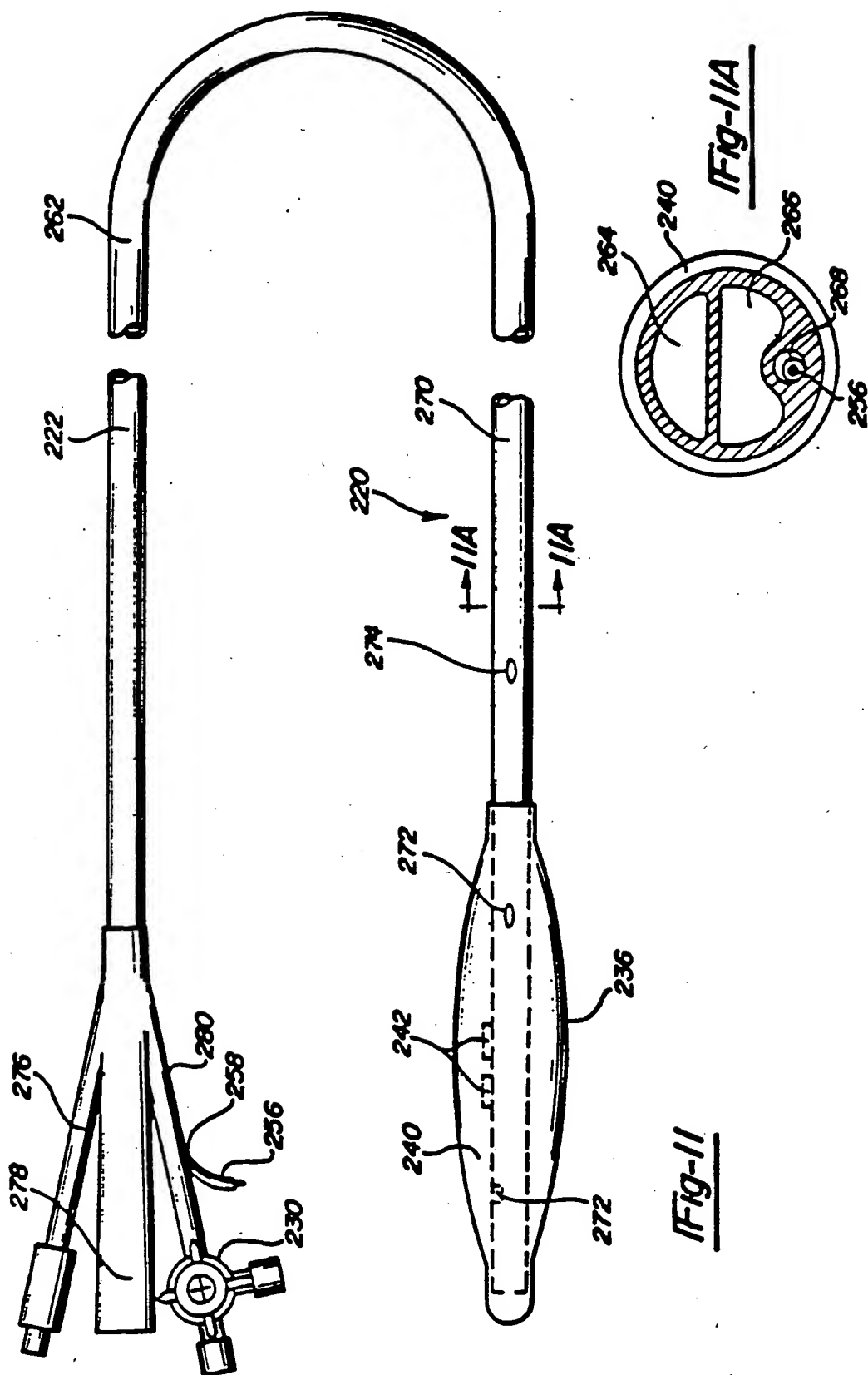
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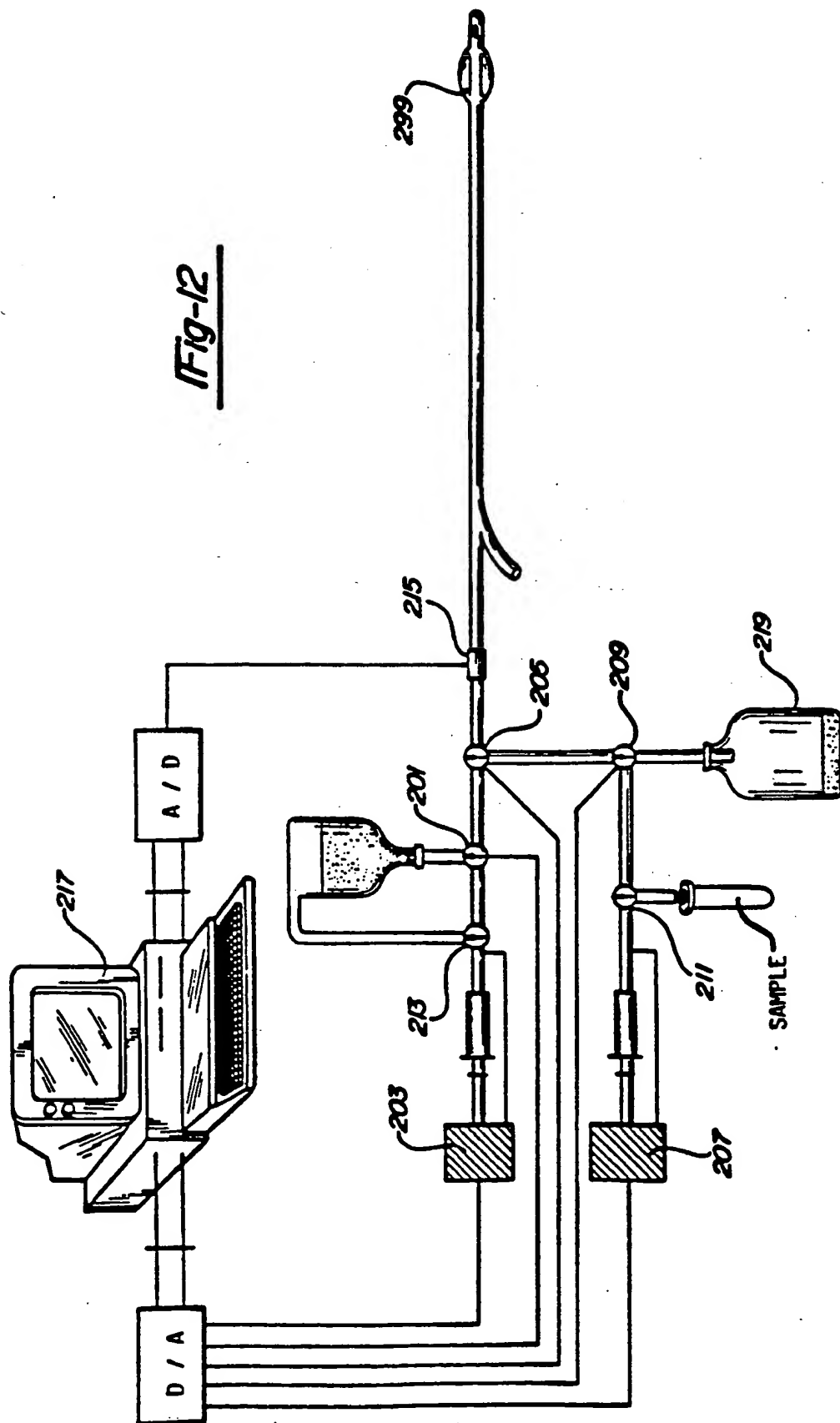
Fig-10

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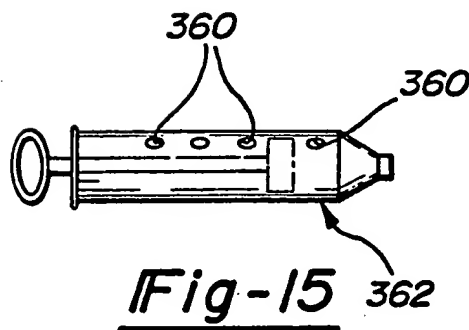
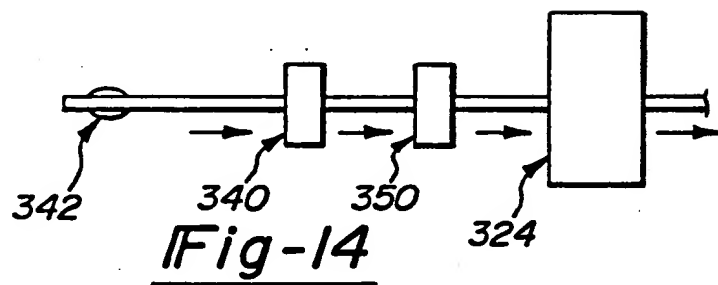
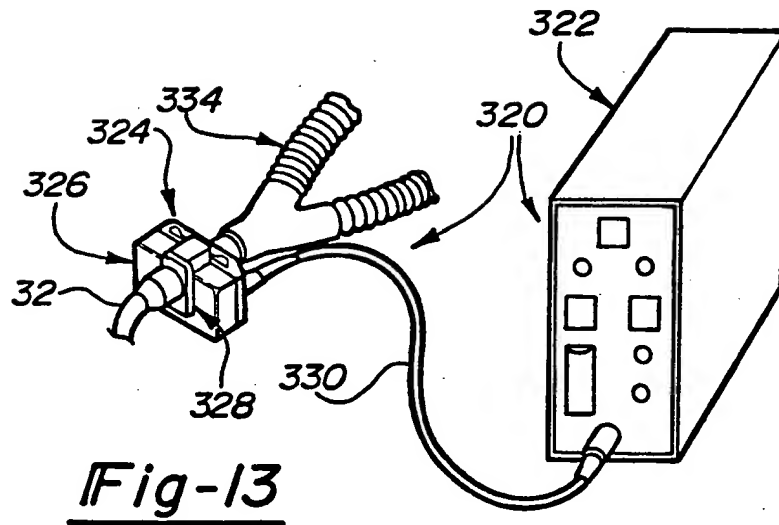


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Fig-12



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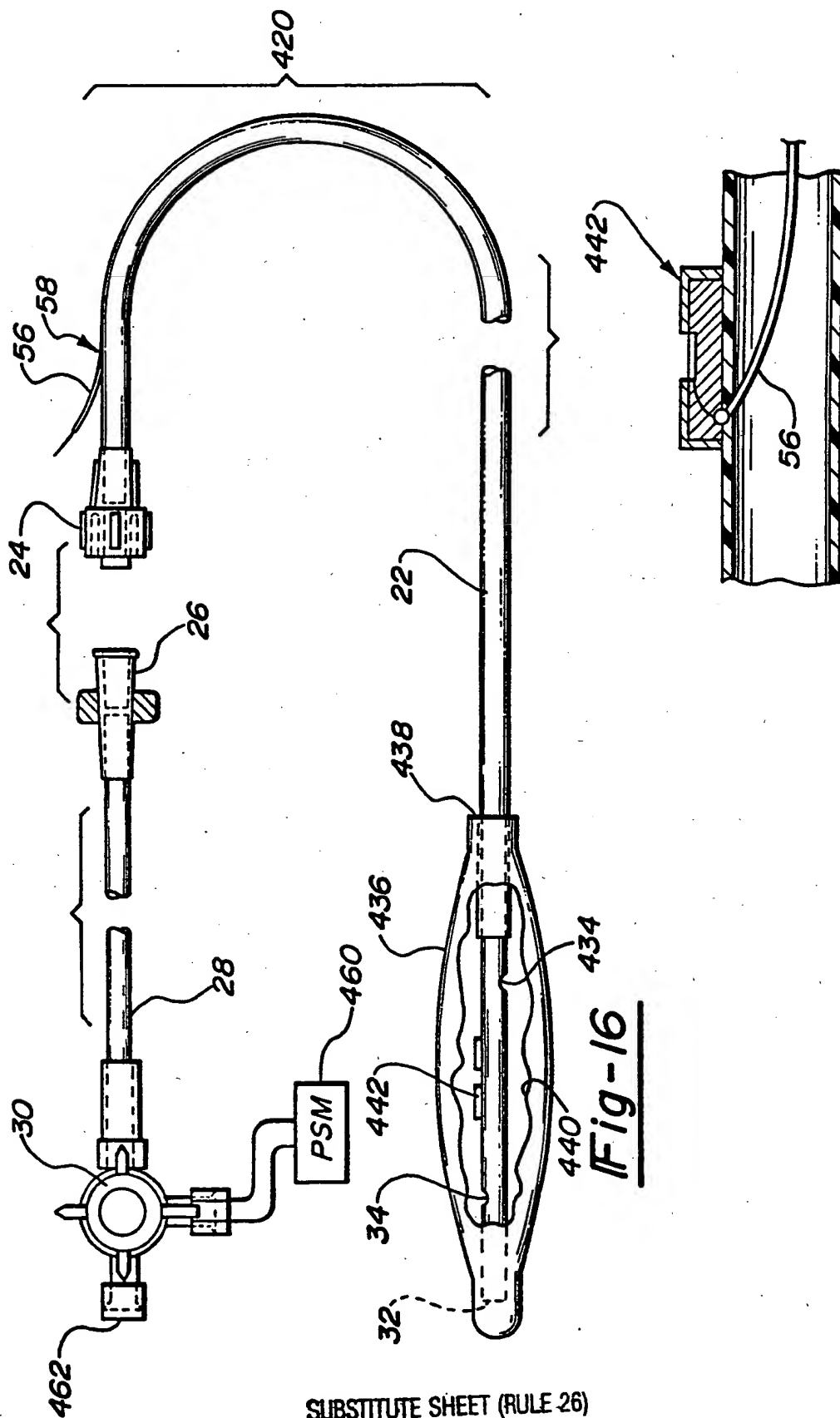
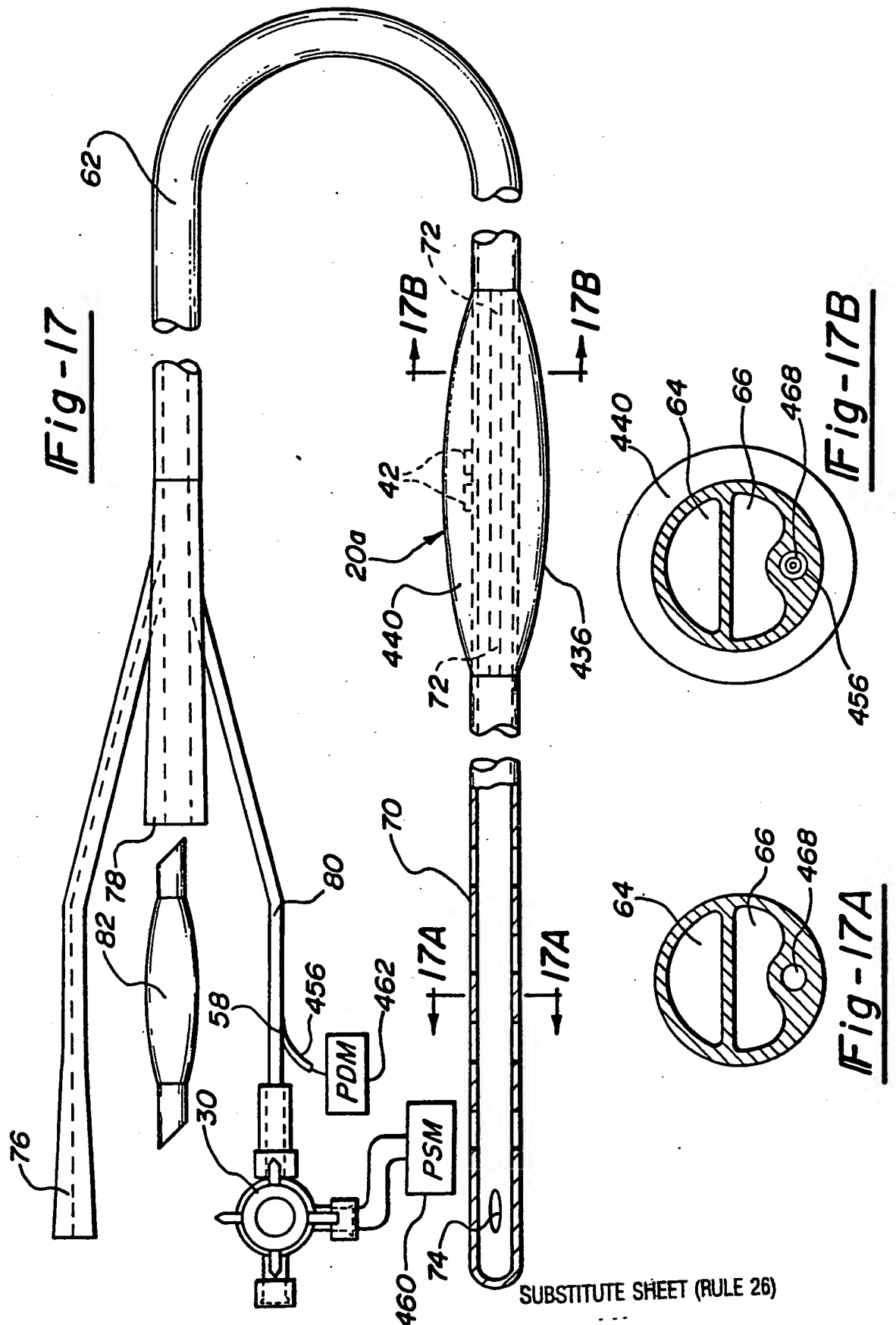


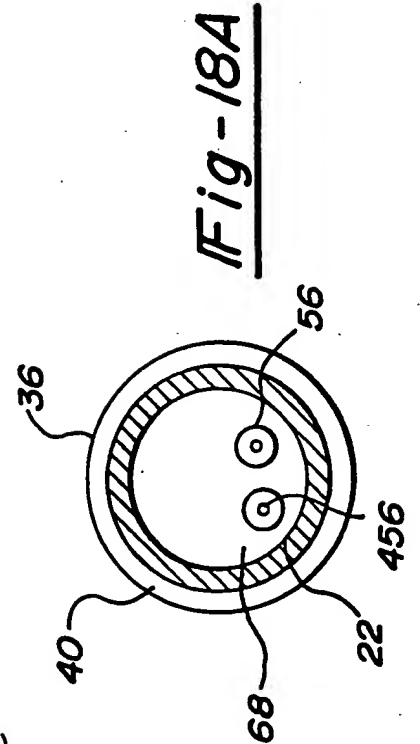
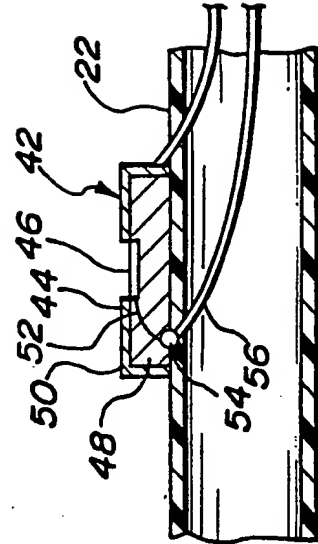
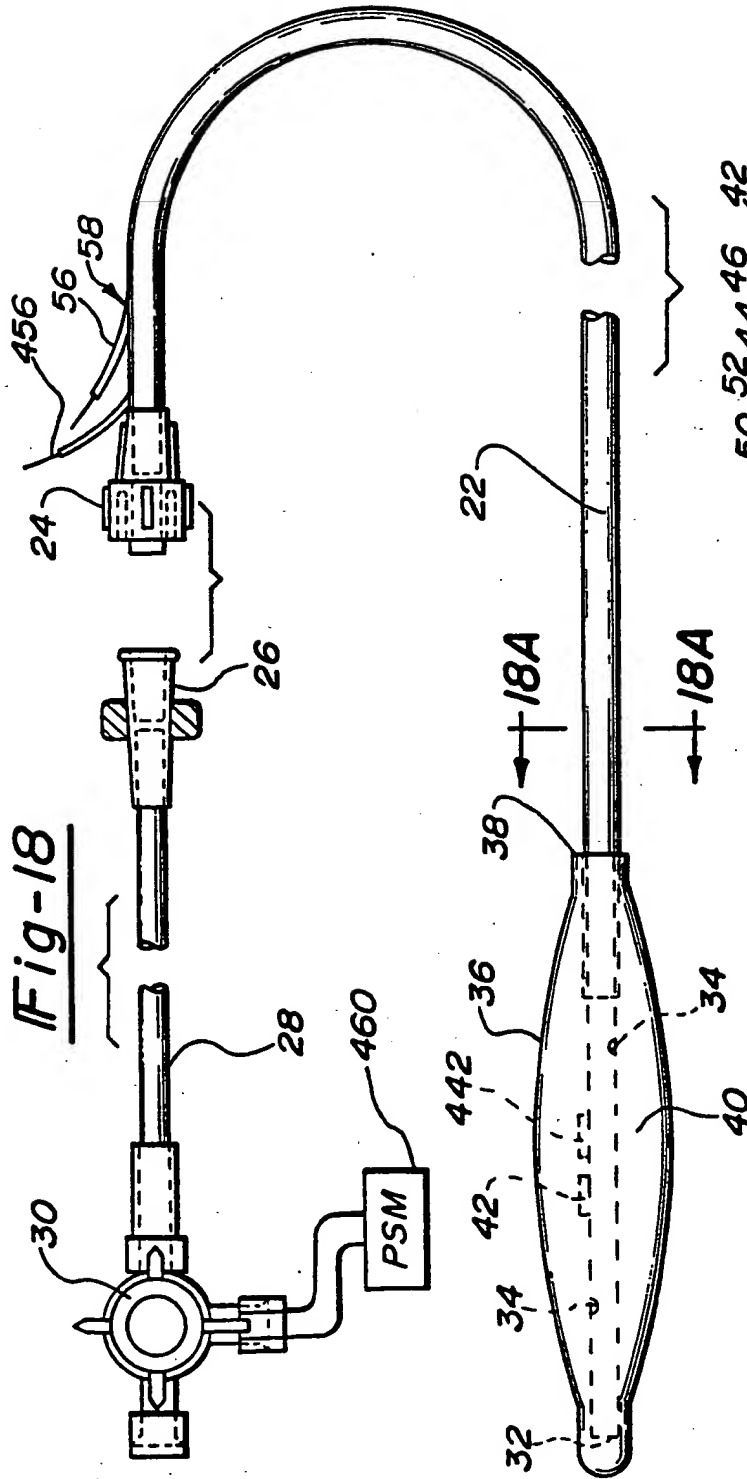
Fig-16A

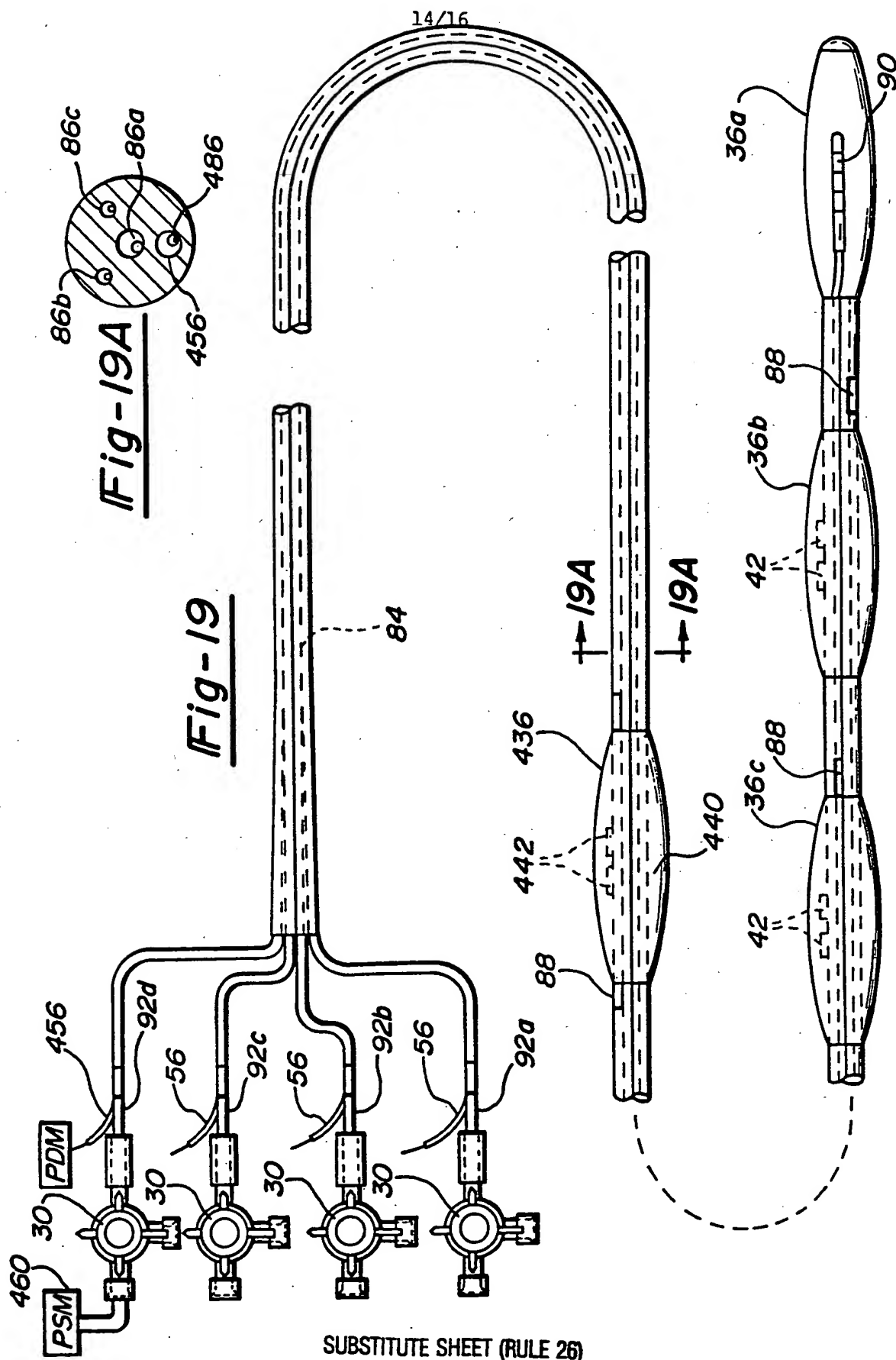
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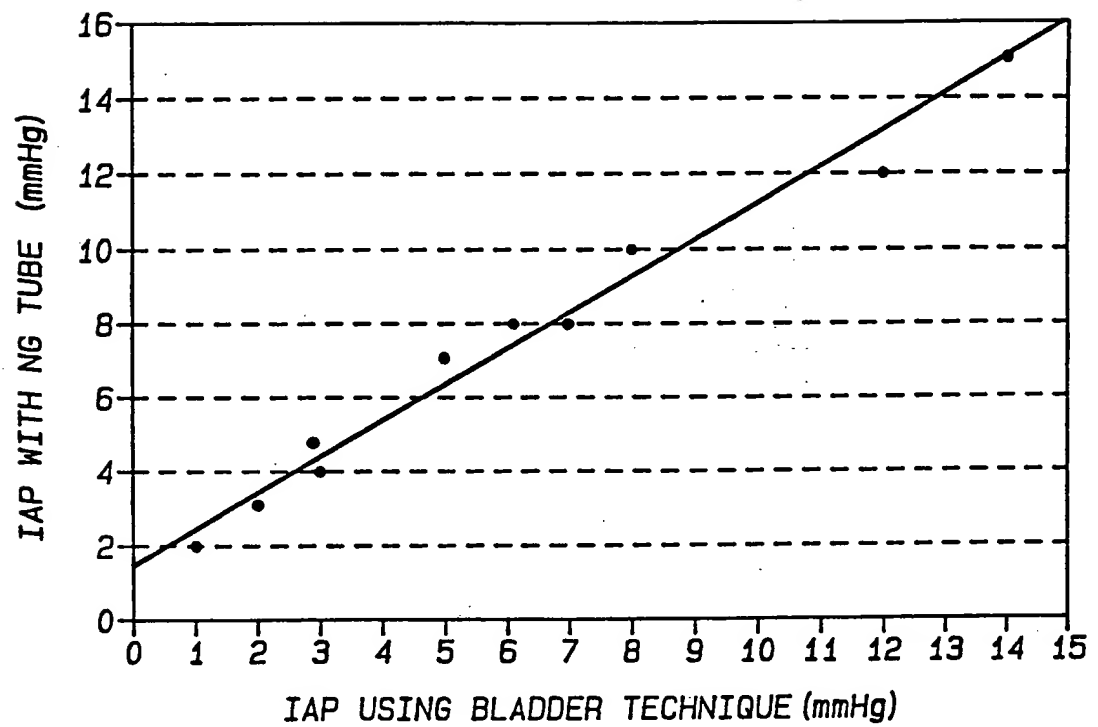
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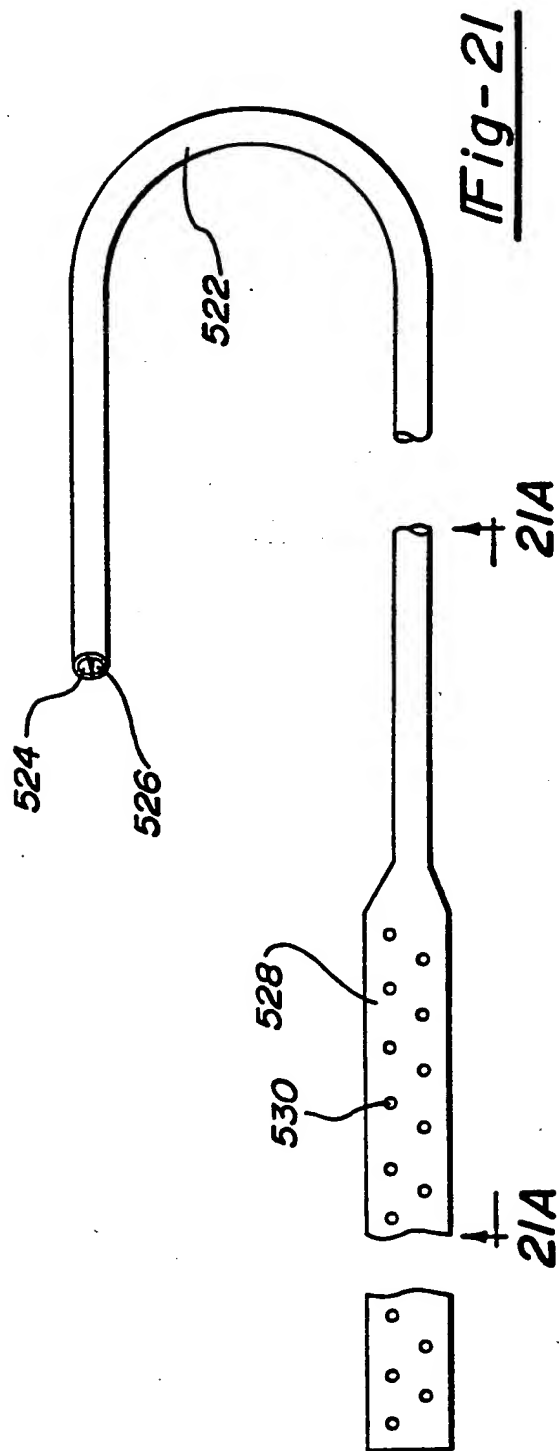
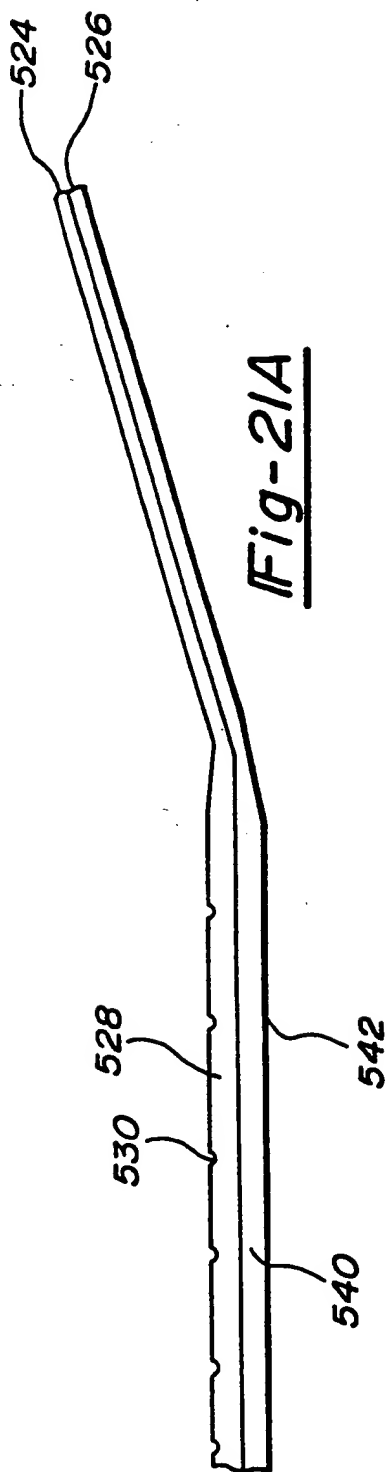
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Fig-20

$r=0.99$
$a=1.37$
$b=0.98$

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/06535

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :A61B 5/103

US CL :128/780

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 128/778, 780; 604/100; 606/191-194

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
NONEElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)
NONE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X --- Y	US, A, 4,981,470, (BOMBECK, IV), 01 January 1991. See entire referemce.	1-4, 8, 9, 13, 22, 23, 28, 30 ----- 5-7, 10-12, 14- 21, 24-27, 29
Y	US, A, 5,174,290, (FIDDIAN-GREEN), 29 December 1992. See Fig. 1.	12, 17-21, 24- 27

☐

Further documents are listed in the continuation of Box C.

☐

See patent family annex.

* Special categories of cited documents:	T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be part of particular relevance	X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	&*	document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means		
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Date of the actual completion of the international search

28 JULY 1994

Date of mailing of the international search report

26 AUG 1994

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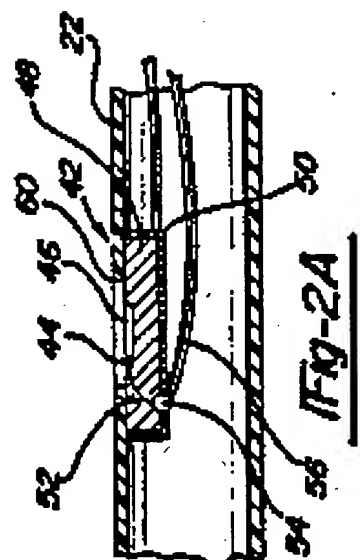
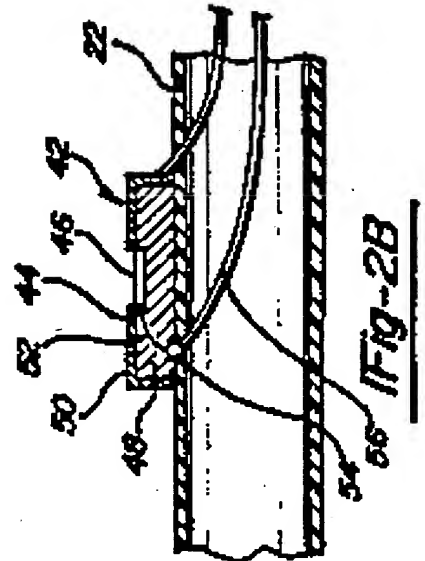
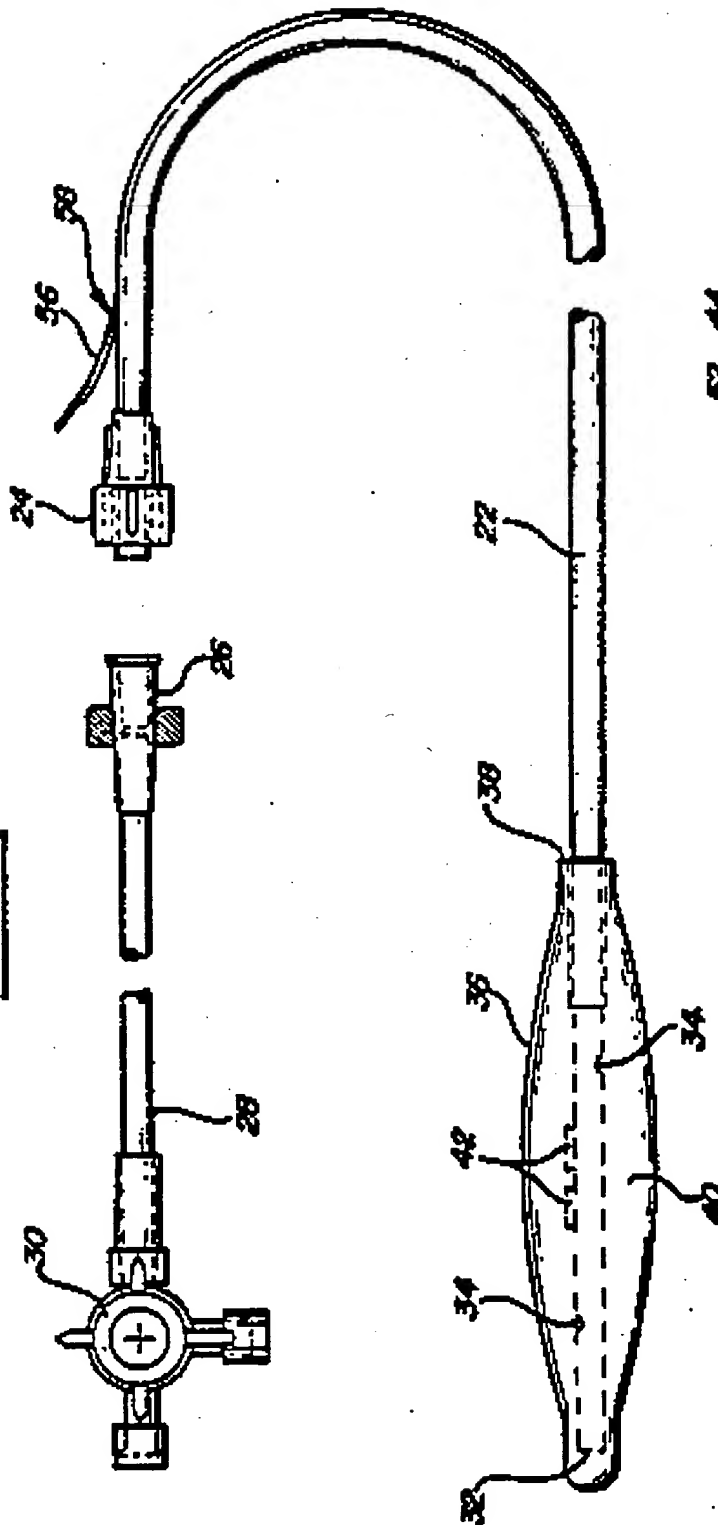
for MAX HINDENBURG

Telephone No. (703) 308-3130

Form PCT/ISA/210 (second sheet)(July 1992)*

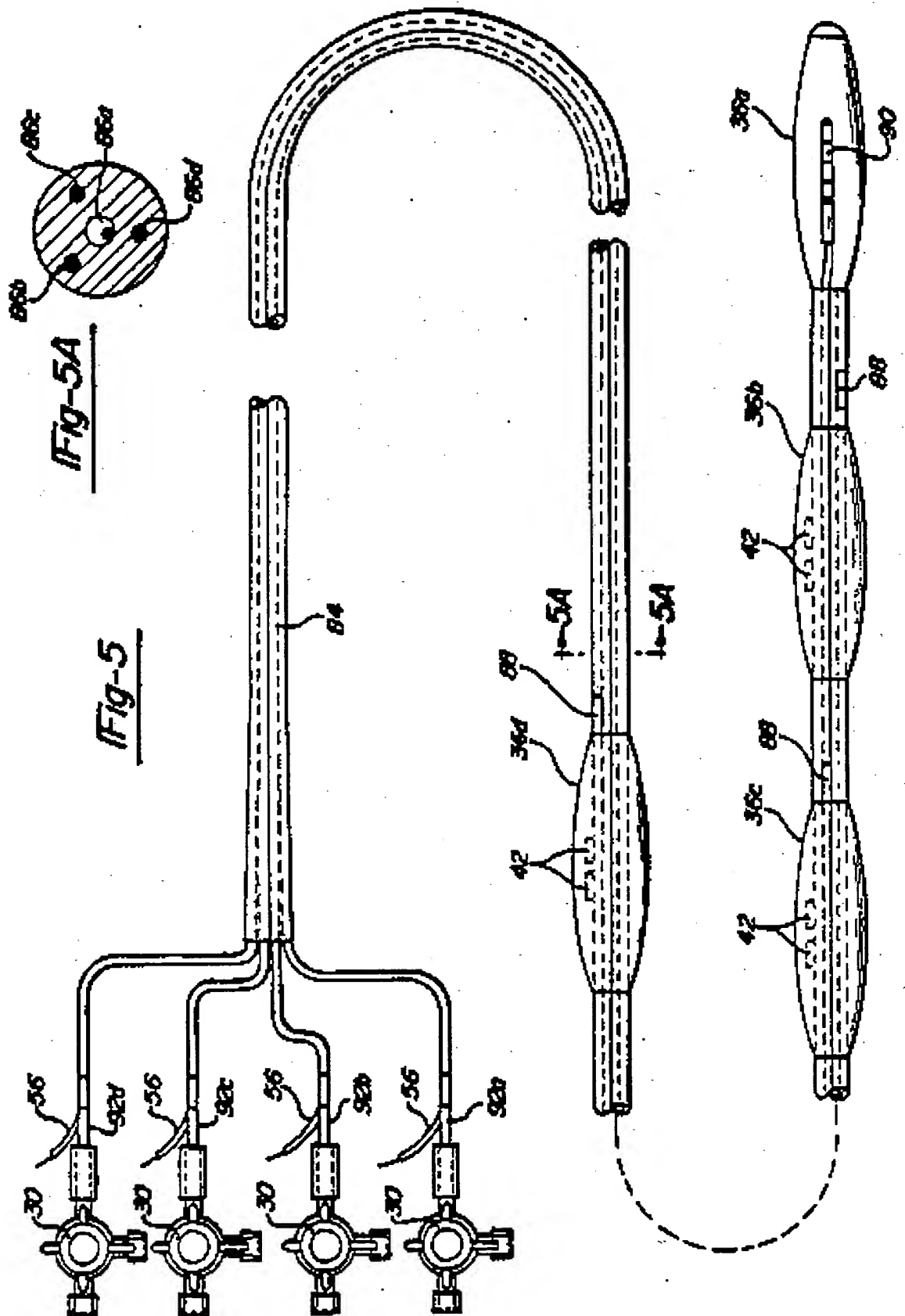
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Fig-1



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Fig-6

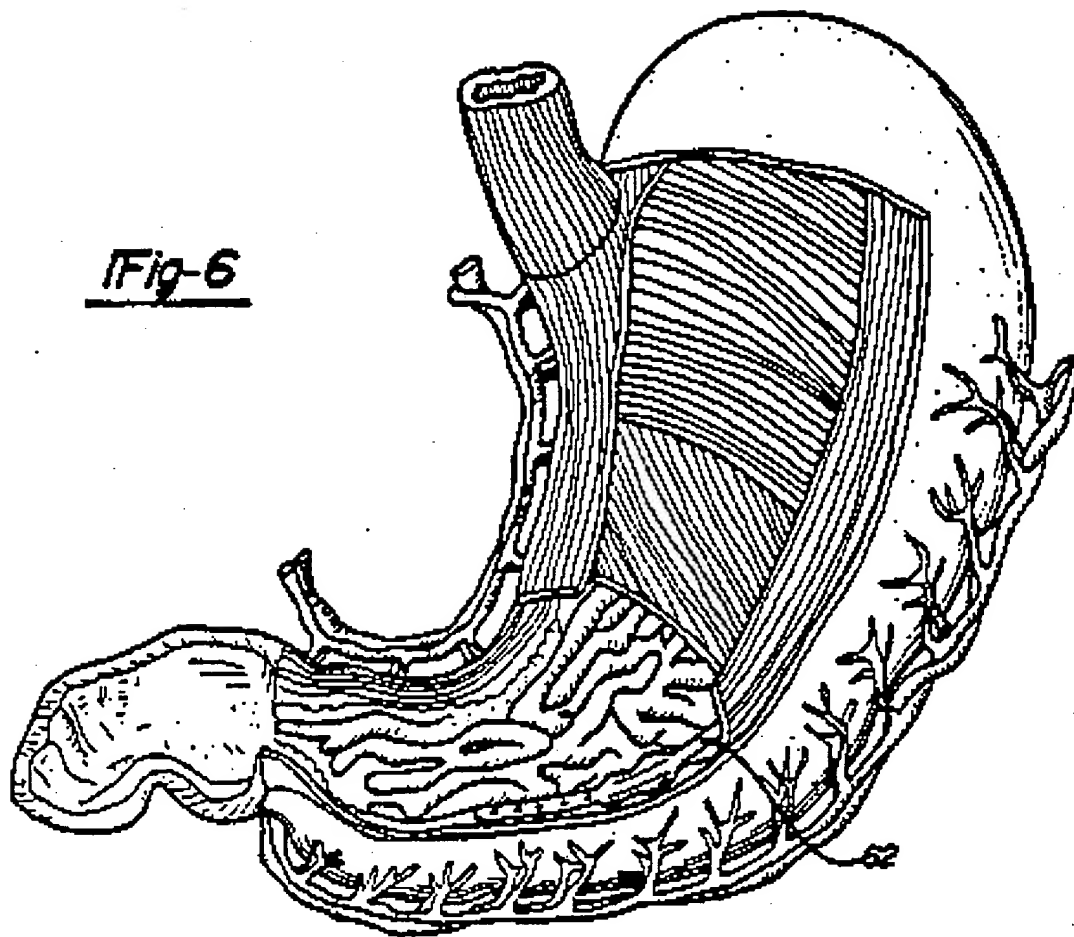
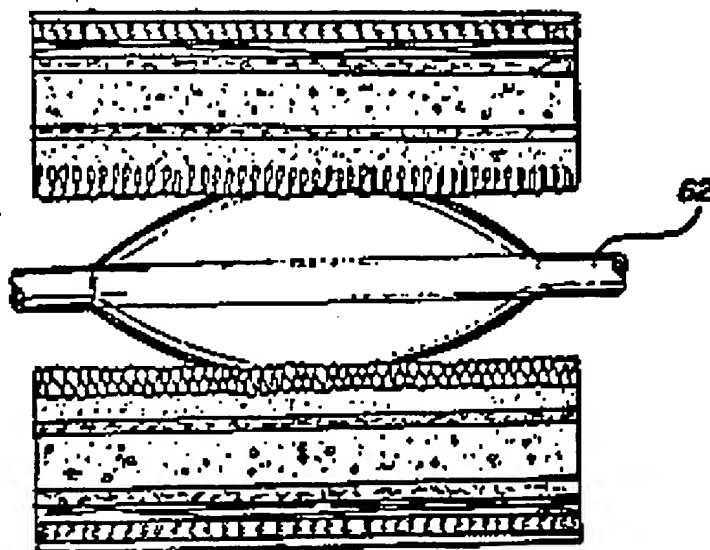
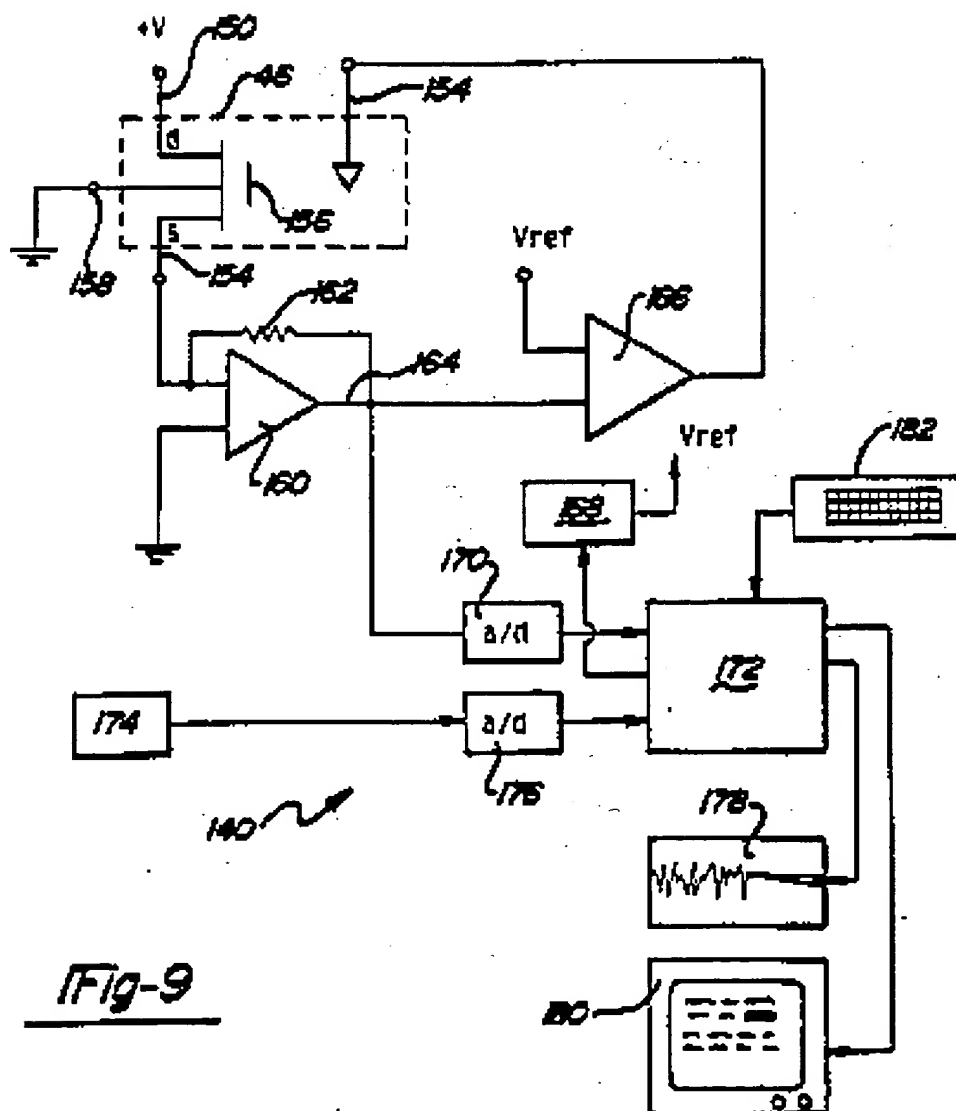
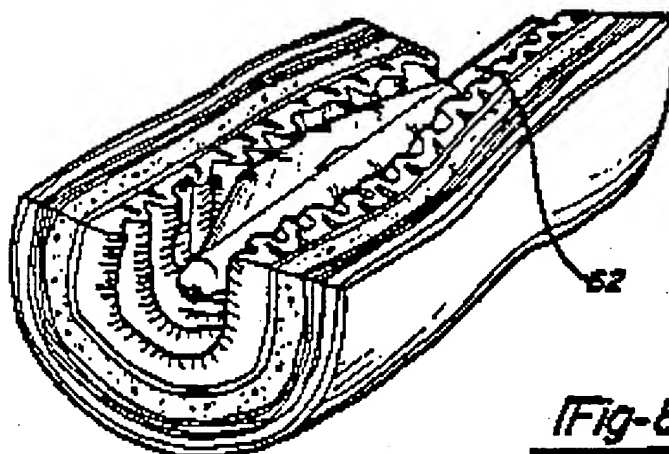


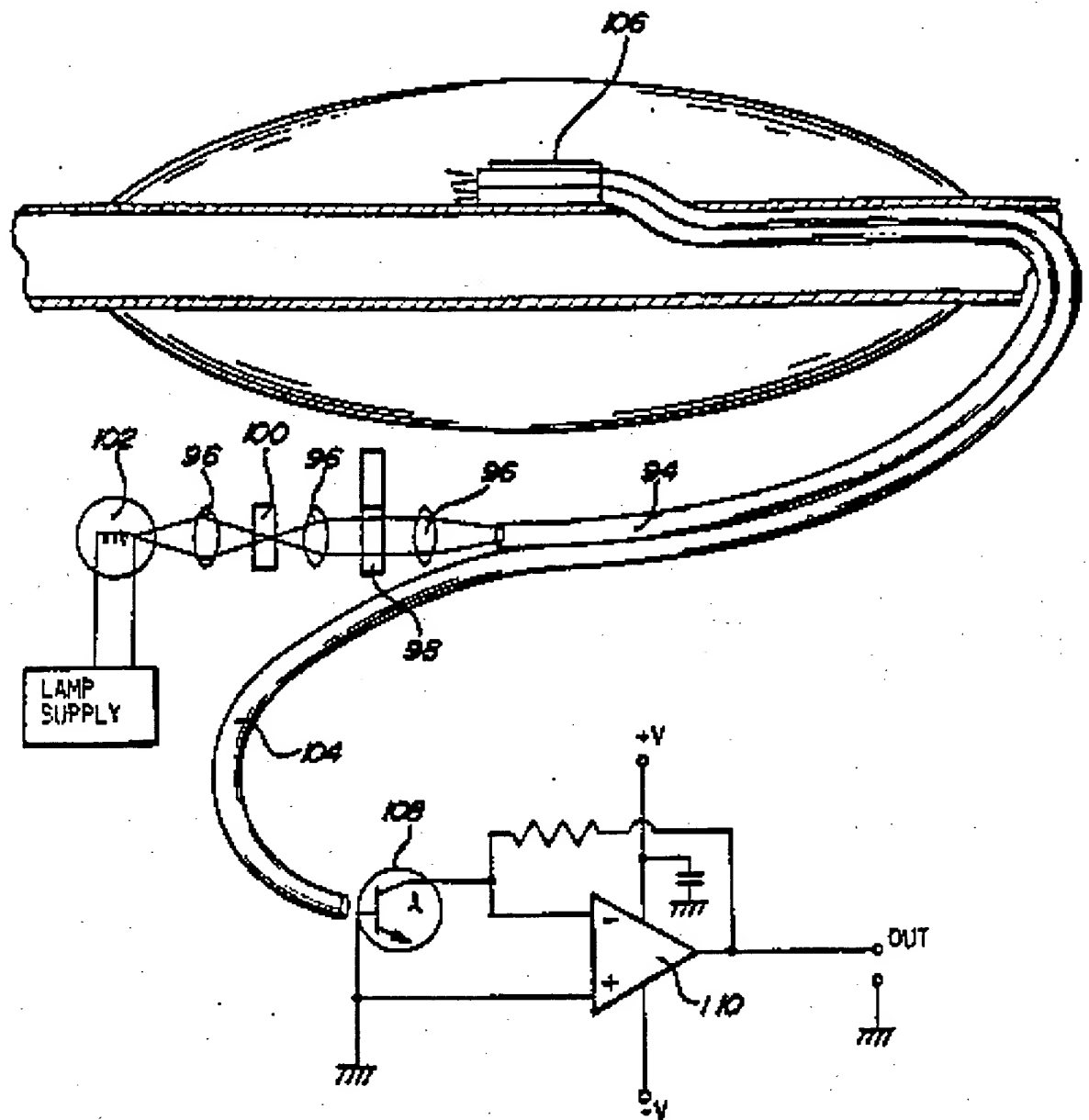
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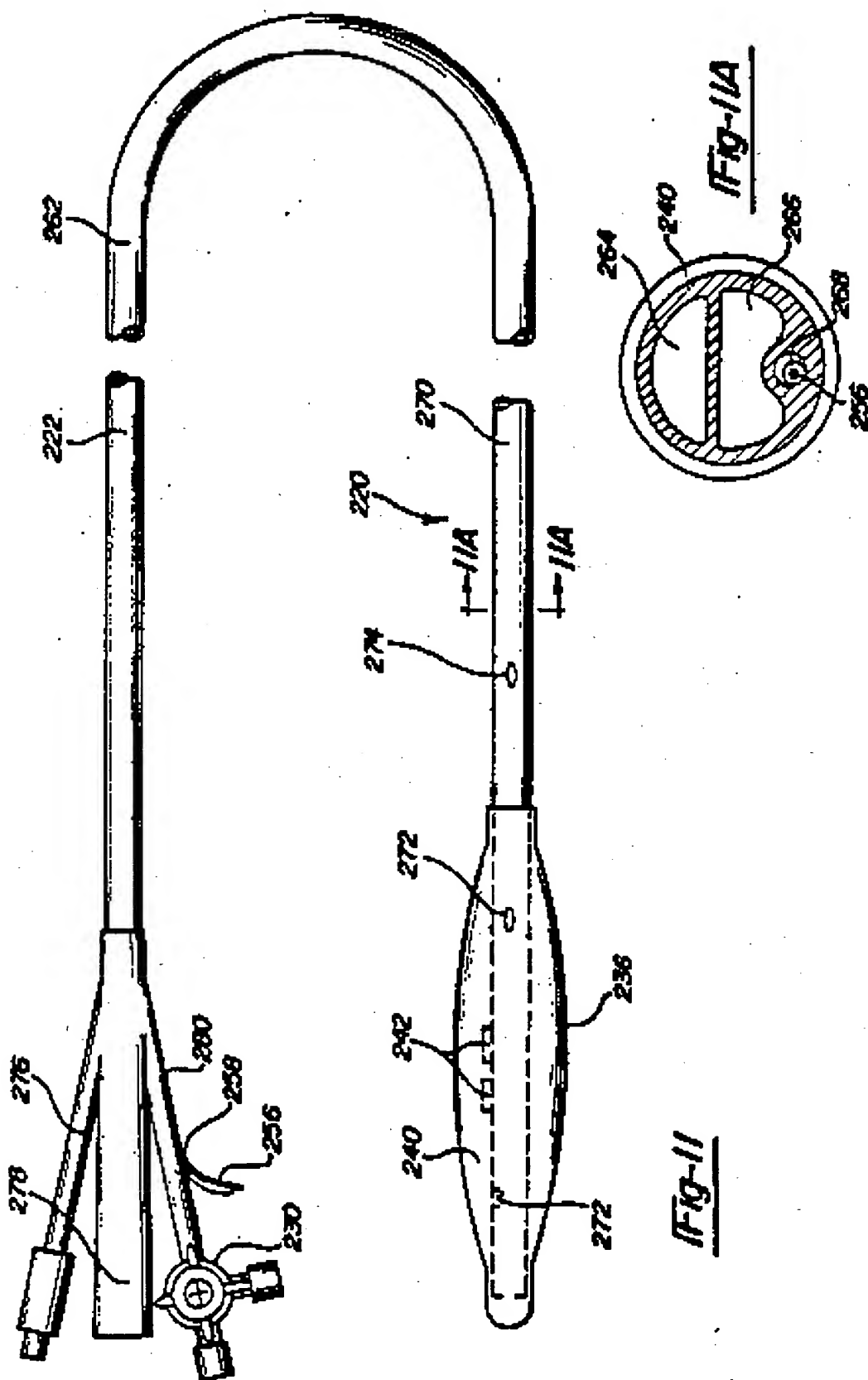
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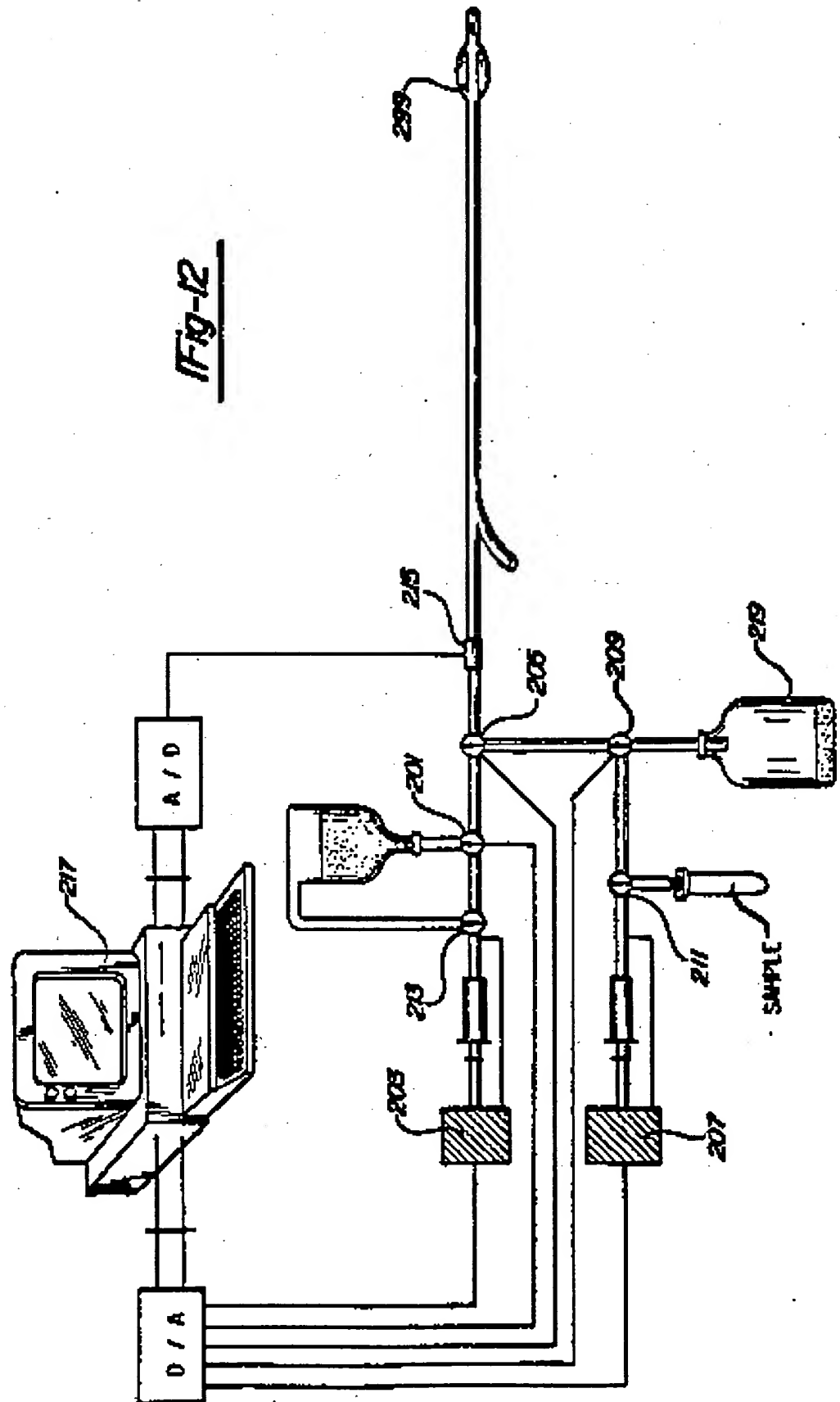
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Fig-10

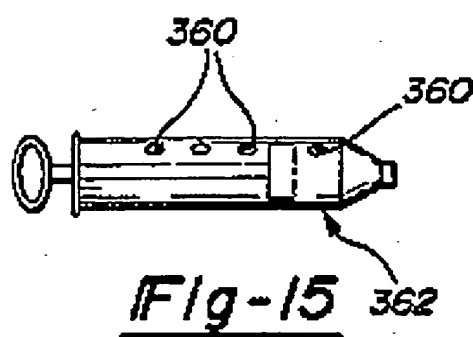
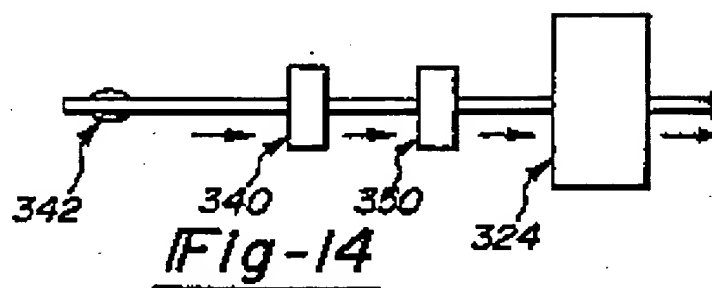
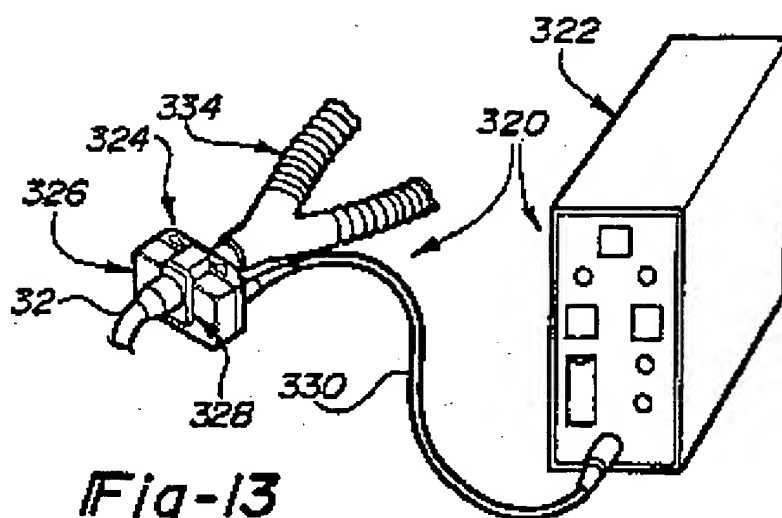
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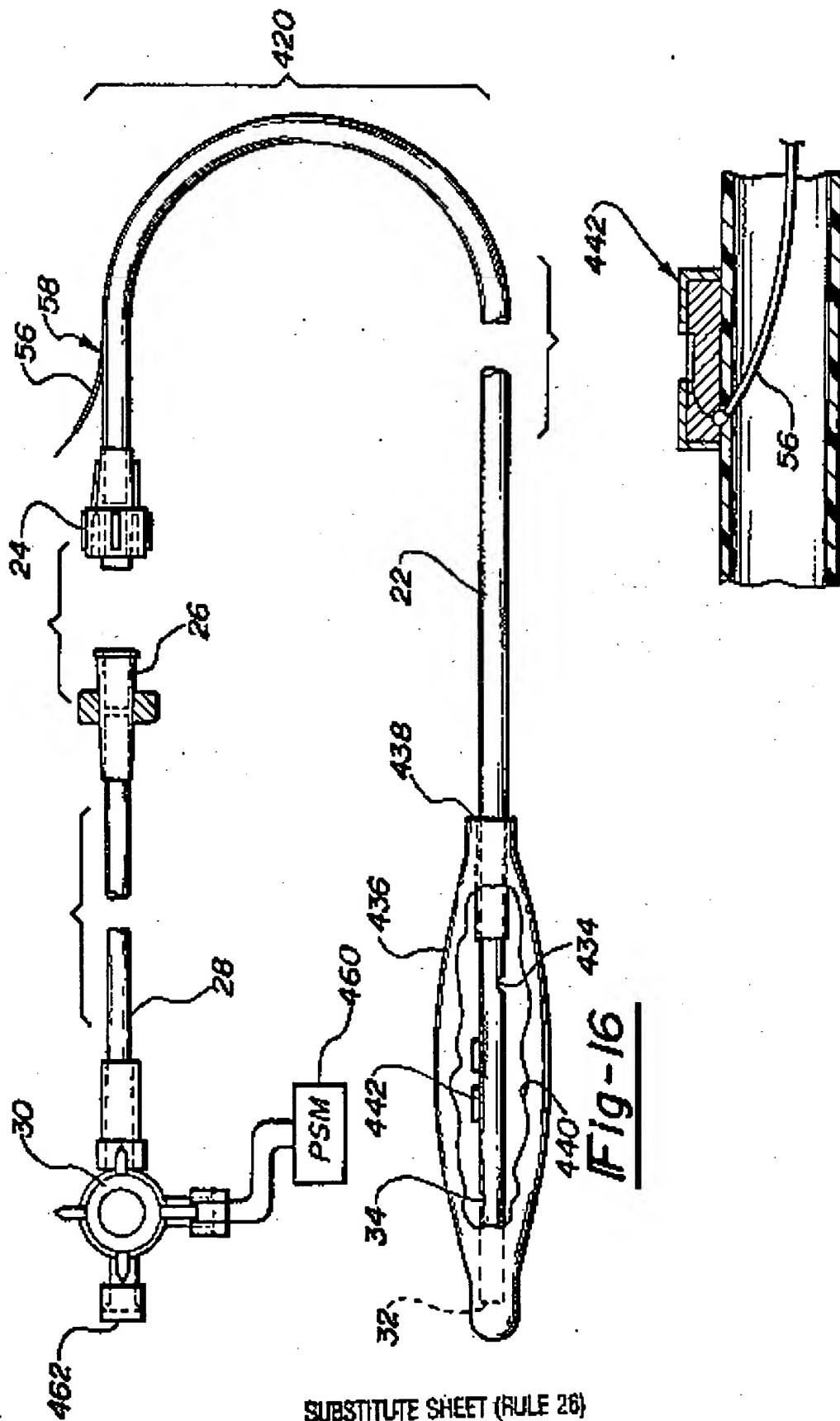
Fig-12

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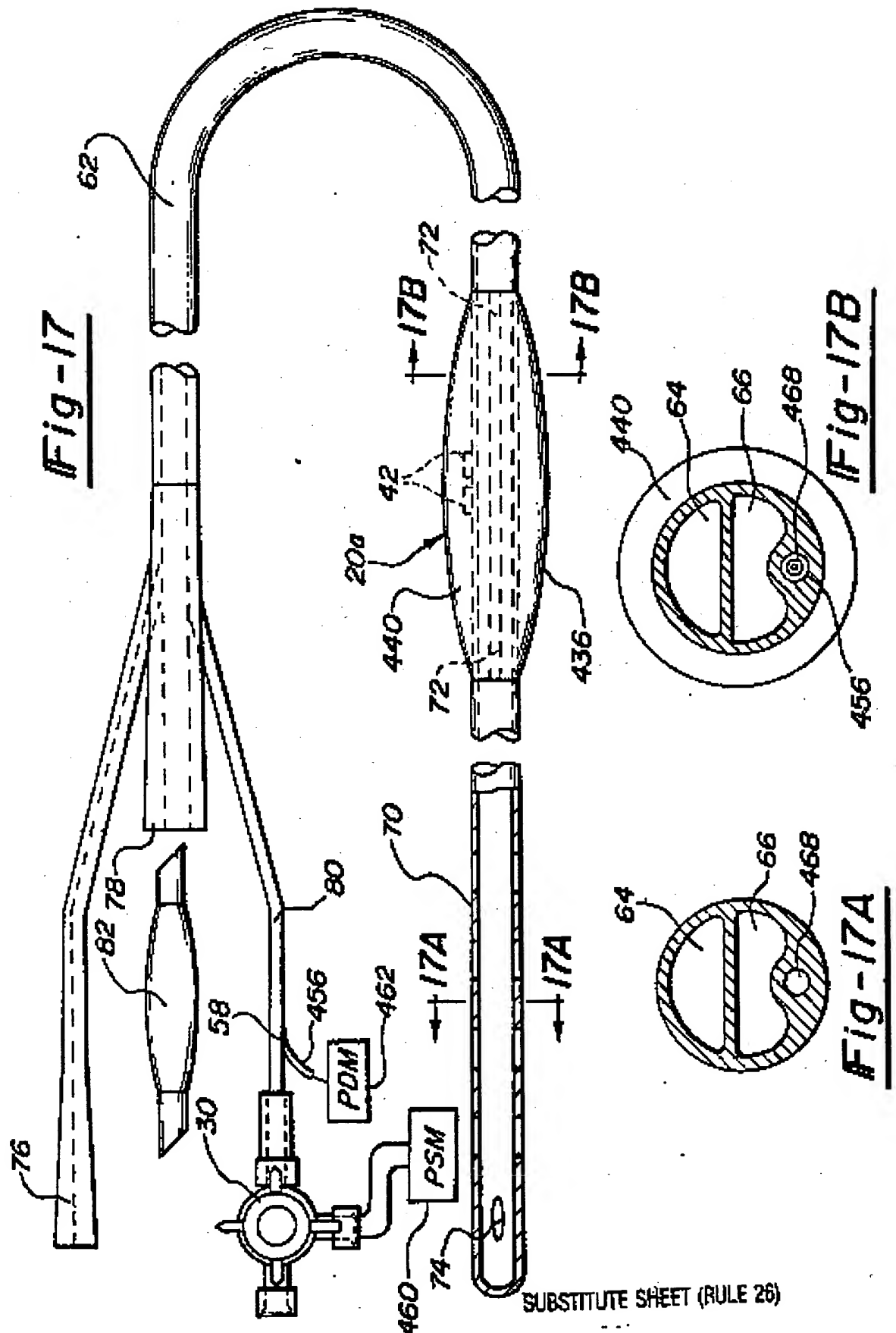
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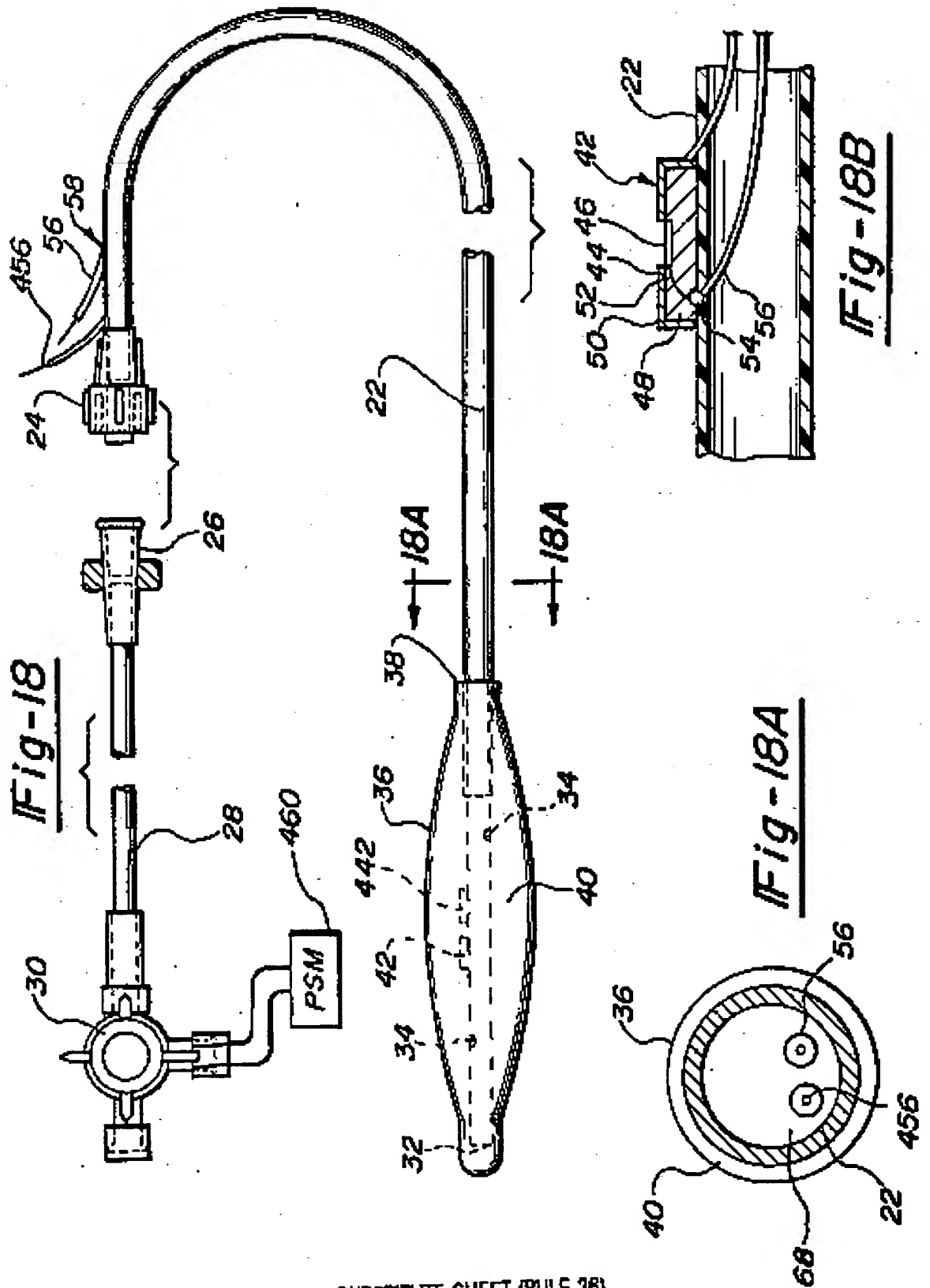
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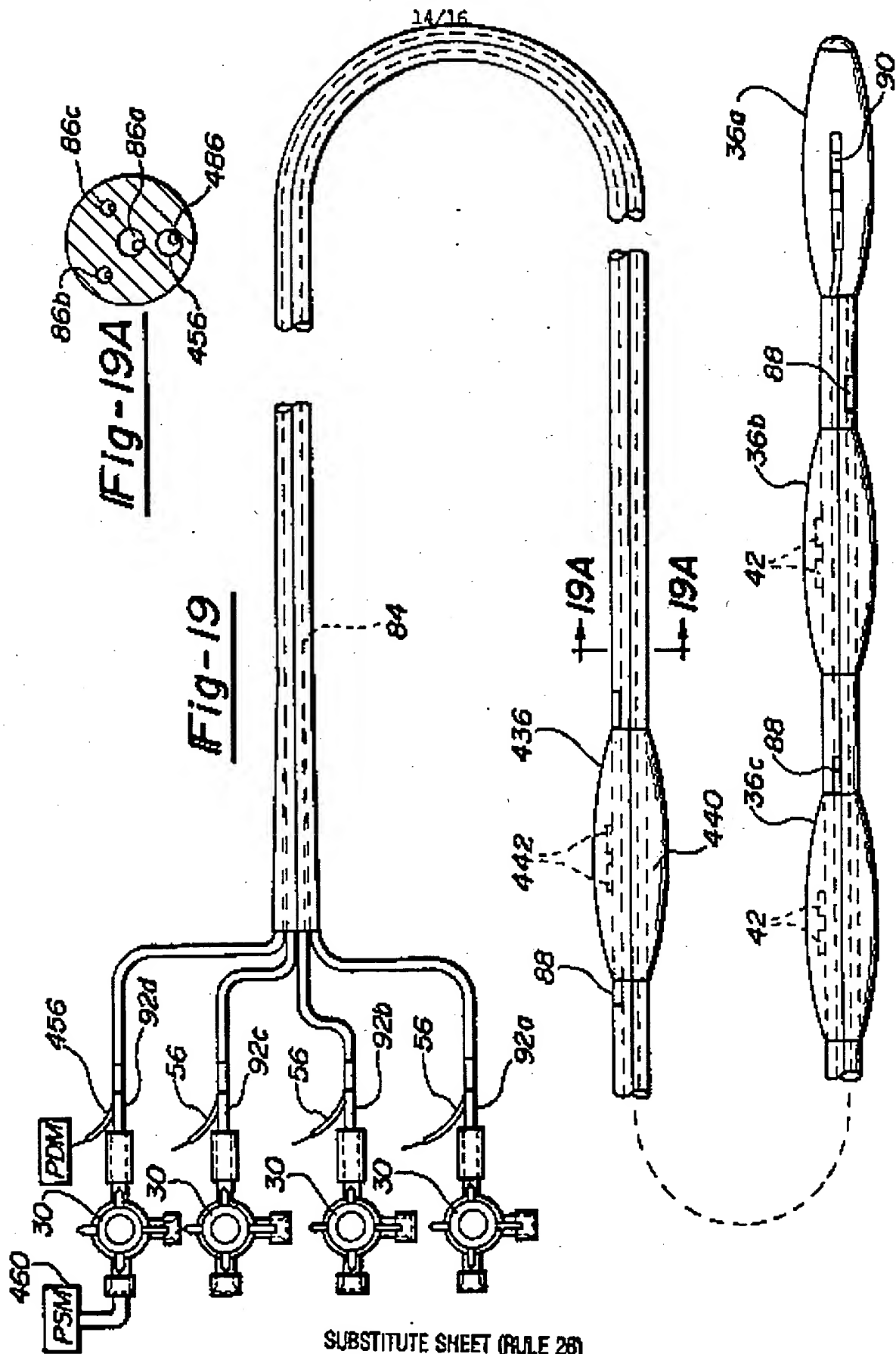
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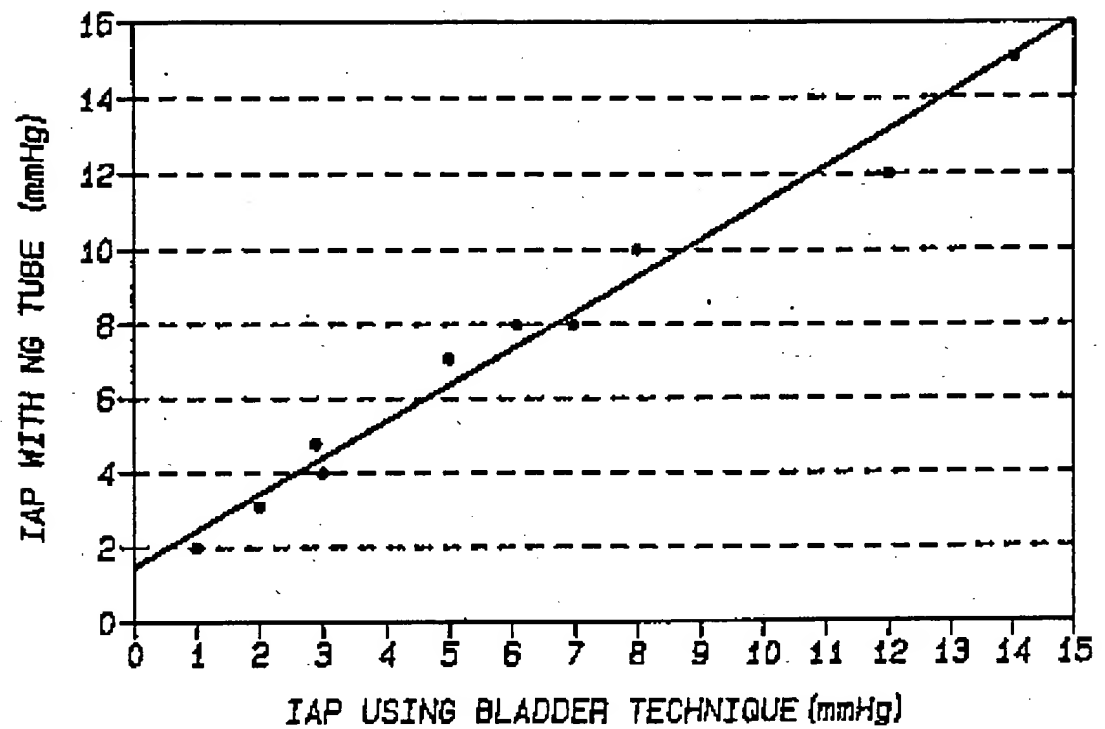


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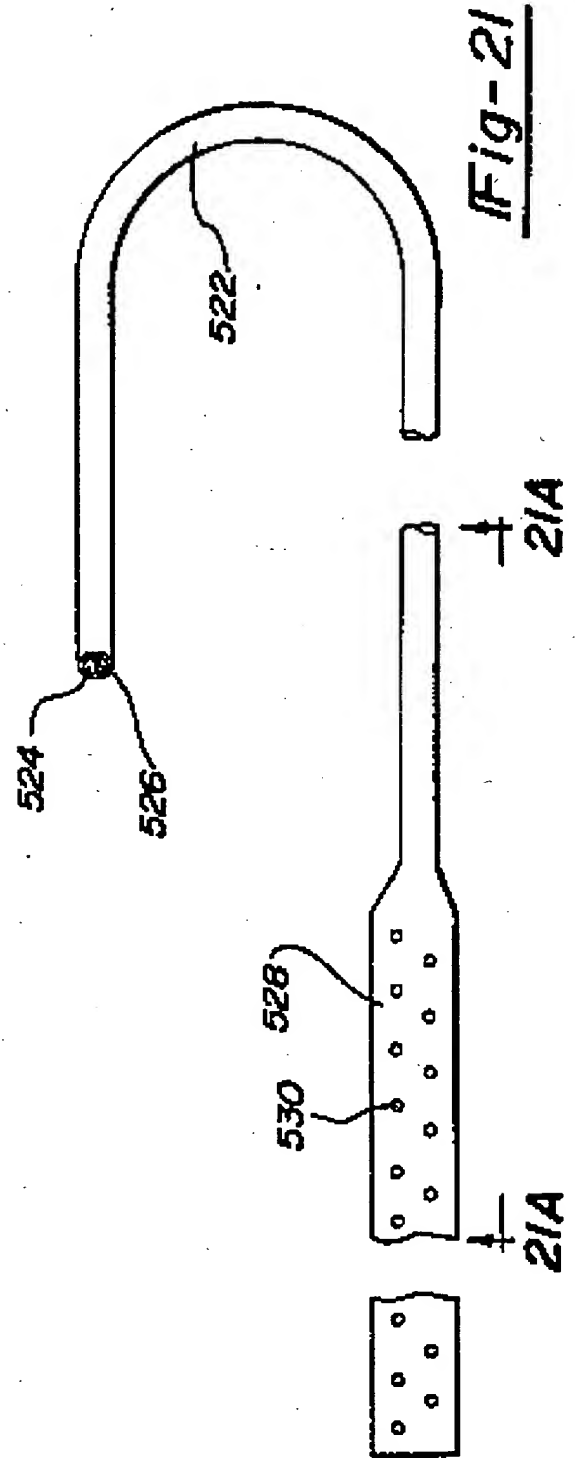
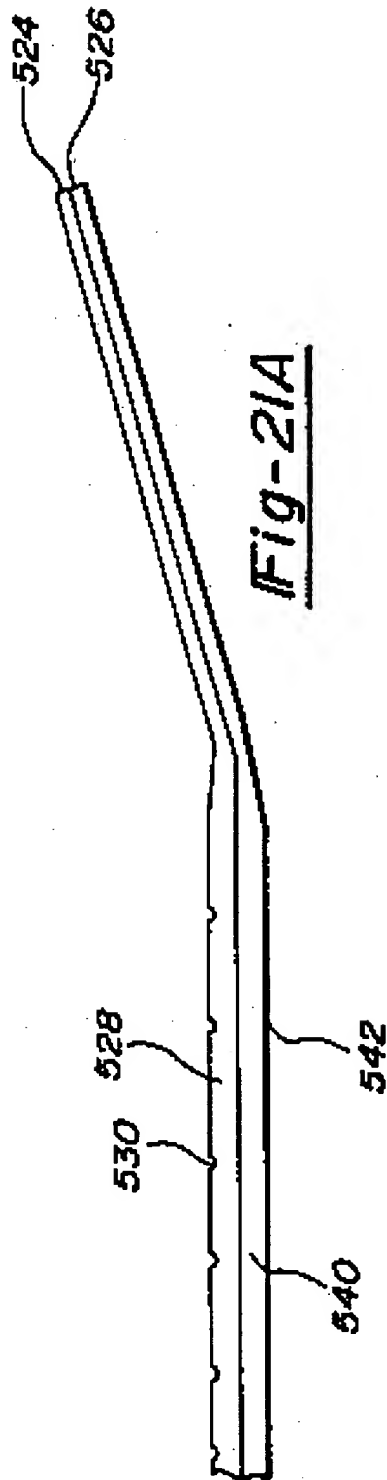


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Fig-20

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